

## Veliparuthi (*Pergularia daemia* (Forsk.) Chiov.) – As a phytomedicine: A review

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**ABSTRACT:** The plant *Pergularia daemia* has been traditionally used as anthelmintic, laxative, antipyretic expectorant and also used to treat infantile diarrhea and malarial intermittent fevers. It is widely distributed in the tropical and sub tropical regions of the world. Various phytochemical including terpenoid, flavonoids, sterols and cardenolids have been isolated and identified from the various parts of the plant (leaves, stems, shoots, roots, seeds and fruits). *P. daemia* widely used by various tribal communities in Western Ghats of India for the treatment of variety of ailments, while predominantly the roots of the plant have been used to treat liver disease and jaundice. The present review article aims towards medicinal properties, chemical constituents and other important aspects of *P. daemia*.

**Keywords:** Ethnobotanical uses, *Pergularia daemia*, pharmacological activities, phytochemistry

### INTRODUCTION

Plant and plant products are being used as a source of medicine since long. According to World Health Organization (WHO) more than 80% of the world's population, mostly in poor and less developed countries depend on traditional plant based medicines for their primary healthcare needs [1]. The efficacy and safety of herbal medicine have turned the major pharmaceutical population towards medicinal plant's research. Owing to the global trend towards improved 'quality of life', there is considerable evidence of an increase in demand from medicinal plant [2]. Use of plants for treating various ailments of both man and animal is as old practice as man himself. India is richly endowed with a wide variety of plant shaving medicinal value. These plants are widely used by all sections of the society whether directly as folk remedies or indirectly as pharmaceutical preparation of modern medicine [3]. In recent times, focus on plant research has increased all over the world and a large body of evidence has collected to show immense potential of medicinal plants used in various traditional systems (Ayurveda, Siddha and Unani) [4] and also major source of biodynamic compounds of therapeutic values [5]. Exploration of the chemical constituents of the plants and

pharmacological screening may provide us the basis for developing the lead for development of novel agents. Herbs have provided us some of the very important life saving drugs used in the armamentarium of modern medicine. Among the estimated 400,000 plant species, only 6% have been studied for biological activity, and about 15% have been investigated phytochemically [6]. This shows a need for investigation of various chemical constituents, its activity and phytopharmacological evaluation of herbal drugs.

The plant *Pergularia daemia* (Asclepiadaceae) [7] known as "Veliparuthi" in Tamil, "Uttaravaruni" in Sanskrit and "Utranajutuka" in Hindi. Traditionally the plant *P.daemia* is used as anthelmintic, laxative, antipyretic and expectorant, also used to treat infantile diarrhoea and malarial intermittent fevers [8-10]. Latex of this plant used for toothache [11]. Stem bark remedy for cold [12] and fever [13]. Aerial parts of this plant reported the various pharmacological activities like hepatoprotective [14], antifertility [15], anti-diabetic [16], analgesic, antipyretic and anti-inflammatory [17]. Phytochemically the plant has been investigated for cardenolides, alkaloid and saponins [17]. The plant was found to contain various triterpenes and steroidal compounds [18].

**PLANT REVIEW****General information** <sup>[19]</sup>

A slender, hispid, fetid- smelling perennial climber. Leaves opposite, membranous, 3-9 cm long and about as wide, broadly ovate, orbicular or deeply cordate, acute or short-acuminate at apex, pubescent beneath, petioles 2-9 cm long. Flowers greenish-yellow or dull white tinged with purple, borne in axillary, long-peduncled, drooping clusters. Fruits (follicles) lanceolate, long-pointed, about 5 cm long, covered with soft spines and seeds are pubescent, broadly ovate. Flowering may occur each year between August and January in central India, with fruits maturing from October to February. In central Indian deciduous forests, the stems typically die down in February and reappear with the onset of the rainy season.

**Habitat** <sup>[19]</sup>

A widely distributed in the tropical and sub tropical area. In India it is very commonly found in hedges through out most of cenfry to an altitude about 1000m in Himalayas and 900m in Southern India.

**Vernacular Names** <sup>[19]</sup>

*P.daemia*(Forsk) Chiv or *P.extensa* N.E.Br or *Daemia extensa* R.Br <sup>[20]</sup>

Bengali : Chagulbanti, Changulbati  
Gujarati : Amaradudheli, Chamarudheli,  
Nagaladudhi, Nagaladhdheli  
Hindi : Utranajutuka, Utran, Dudhi, Dudhibel,  
Jutuk, Sagovani

Kannada : Haalu koratige, Hala koratige, Juttuve balli, Kurudigana balli,Alavaarana

balli, Talayarana balli

Malayalam : Veliparatti, Veliparuti

Marathi : Utaranavel, Uturhi

Oriya : Juktiruhi, Uttruri, Uturdi

Sanskrit : Uttaravaruni, Kurutakah, Yugaphala, Yugmaphala

Tami : Beliparti, Nandamani, Uthamani, Veliparuthi

Telegu : Dushtupatige, Gurtichettu, Guruti, Jittupaku

**Taxonomy classification** <sup>[21]</sup>

Kingdom : Plantae

Subkingdom : Tracheobionta

Super division: Spermatophyta

Division : Magnoliophyta

Class : Magnoliopsida

Subclass : Asteridae

Order : Gentianales

Family : Asclepiadaceae

Genus : Pergularia

Species : *P. daemia* (Forsk) Chiv

**Ethnomedical Information**

Aerial parts of the plant used for snake bite <sup>[22]</sup>. Entire plant used as an anthelmintic <sup>[23]</sup>, emmenagogue <sup>[24]</sup>, emetic <sup>[25, 26]</sup>, antiseptic <sup>[27]</sup>, emetic expectorant <sup>[27]</sup>, expectorant <sup>[25,26,28]</sup> and antivenin <sup>[29]</sup> and used to facilitate parturition <sup>[30]</sup>, while used in Ayurvedic medicine for delayed childbirth <sup>[31]</sup>, amenorrhea <sup>[23]</sup>, asthma, snakebite, rheumatic swellings <sup>[26]</sup> and used to treat post-partum hemorrhage <sup>[31]</sup>. Latex of this plant used for boils and

sores <sup>[32]</sup>. Dried leaf used as an emetic <sup>[33]</sup>, antirheumatic <sup>[34]</sup> and used for bronchitis <sup>[33]</sup>, amenorrhea, dysmenorrheal <sup>[35, 36]</sup>, asthma <sup>[25]</sup>, healing cuts and wounds <sup>[37]</sup>, while used to treat whooping cough <sup>[38]</sup> and to facilitate parturition <sup>[36]</sup>. Fresh leaf used as fish poison <sup>[39]</sup>, while leaf juice used for amenorrhea, dysmenorrheal, catarrhal infections, infantile diarrhea <sup>[25]</sup> and used reduce the body pain <sup>[40,41]</sup>. Dried root used as an abortifacient <sup>[42]</sup>, emetic, bronchitis <sup>[33]</sup> and used for cough, asthma and constipation <sup>[39]</sup>, while fresh root used as an abortifacient <sup>[43, 44]</sup> and used to treat gonorrhoea <sup>[45]</sup>. Shoots used to treat whooping cough <sup>[46]</sup>. Stem bark has been used to treat malaria <sup>[47]</sup> and twig used as an antipyretic and appetizer <sup>[48]</sup>.

**CLAIMS AND REPORTS****Pharmacological/Biological activity**

**Suresh Kumar and Mishra, 2008a;** ethanol extract and its ethanol fraction from aerial parts of *P. daemia* exhibited significant hepatoprotective effect against CCl<sub>4</sub> induced hepatotoxicity in rats. Glutamic oxaloacetic transaminase, glutamic pyruvic transaminase, alkaline phosphatase, total bilirubin, total cholesterol, total protein and albumin in serum indicated hepatoprotective effect of the ethanol extract and its ethanol fraction. Histopathological examination of liver sections confirmed that, pre-treatment with ethanol extract and its ethanol fraction prevented hepatic damage induced by CCl<sub>4</sub>. The results were comparable with the standard hepatoprotective drug silymarin. The extract and its fraction showed no signs of toxicity up to a dose level of 2000 mg/kg. It is suggested that, the presence of flavonoids in ethanol extract and its ethanol fraction may be responsible for hepatoprotective properties. High Performance Thin Layer Chromatography profile of flavonoids of bio-active extracts was developed using quercetin-3-glucoside as a marker. Results indicate hepatoprotective properties of ethanol extract of *P. daemia* <sup>[49]</sup>.

**Suresh Kumar and Mishra, 2008b;** studied on the hepatoprotective effect of acetone and ethanol sub fractions of ethanolic fraction obtained from total ethanol extract was carried out using carbon tetrachloride-induced toxicity in primary cultured rat hepatocytes. In vitro activity was assessed by determining the change in hepatocyte viability and other biochemical parameters such as glutamic oxaloacetic transaminase, glutamic pyruvic transaminase and total protein. Acetone and ethanol sub fractions showed significant (P<0.05) protective effect by restoring altered parameters in the selected in vitro model. The flavonoids present in acetone and ethanol sub fractions of total alcohol extract from *P. daemia* may be responsible for significant hepatoprotective properties. The results justify the claims of *P. daemia* in folk medicine as a hepatoprotective agent <sup>[50]</sup>.

**Flyman and Afolayan, 2007;** studied the implication of the mineral ratios of *P. daemia* in human diets. The Ca/Fe, Ca/K, Ca/Mg, and Ca/Zn ratios were 1.7, 0.3, 1.6, and 9.7, respectively. The Fe/Zn ratio was 5.6, while P/Ca ratio was 0.3 [51].

**Suresh Kumar and Mishra, 2007;** studied on the hepatoprotective effect of acetone and ethanolic sub fractions of ethanolic fraction obtained from total alcoholic extract was carried out using carbon tetrachloride- induced liver damage in wistar albino rats. Acetone sub fraction showed significant ( $P < 0.05$ ) protective effect by lowering serum levels of various biochemical parameters in the selected model. These biochemical observations were supplemented by histopathological examination of liver sections. Silymarin was used as positive control. The presence of flavonoid compounds in the ethanolic sub fraction of alcohol extract of *P. daemia* may be responsible for significant hepatoprotective properties. The results justify use of *P. daemia* as a hepatoprotective agent [52].

**Suresh Kumar and Mishra, 2006;** reported hepatoprotective effect of crude ethanolic and aqueous extracts from the aerial parts of *P. daemia*. The aqueous and ethanolic extracts obtained from aerial parts of *P. daemia* were evaluated for hepatoprotective activity in rats by inducing liver damage by carbon tetrachloride. The ethanolic extract at an oral dose of 200 mg/kg exhibited a significant ( $P < 0.05$ ) protective effect by lowering serum levels of glutamic oxaloacetic transaminase, glutamic pyruvic transaminase, alkaline phosphatase, total bilirubin and total cholesterol and increasing the levels of total protein and albumin levels as compared to silymarin used as a positive control. These biochemical observations were supplemented by histopathological examination of liver sections. The activity may be a result of the presence of flavonoid compounds. Furthermore, the acute toxicity of the extracts showed no signs of toxicity up to a dose level of 2000 mg/kg. Thus it could be concluded that ethanolic extract of *P. daemia* possesses significant hepatoprotective properties [14].

**Hebbar et al., 2004;** reported the ethnomedicine of Dharwad district of Karnataka in southern India. It was revealed that *P. daemia* was used to treat tooth ache [11].

**Kohler et al., 2002;** reported lipophilic fraction obtained from stem bark was showed antimalarial activity against *Plasmodium falciparum* and the  $IC_{50}$  was found to be  $> 50 \mu\text{g/ml}$  [47].

**Wahi et al., 2002;** studied the ethanol and aqueous extract of entire plant 200 mg/kg showed antihyperglycemic (antidiabetic) activity on rat (both the sex) by alloxan induced hyperglycemia method [16].

**Golam Sadik et al., 2001a;** reported ethanolic extract of *P. daemia* and its steroidal fraction 200 mg/kg body

weight showed significant anti fertility activity in preimplantation stage of female mice [15].

**Golam Sadik et al., 2001b;** studied the oral administration of the alkaloidal fraction of ethanol extract at a dose of 200 mg/kg body weight showed significant anti fertility activity in preimplantation stage of female mice [53].

**Srinivasan et al., 2001;** reported the 0.3 ml of aqueous extract of whole plant was inactive against various bacteria and fungal such as *Chromobacterium violaceum*, *Escherichia coli*, *Enterococcus faecalis*, *Klebsiella pneumonia*, *Pseudomonas aeruginosa*, *Proteus mirabilis*, *Staphylococcus aureus*, *Salmonella paratyphi*, *Salmonella typhi*, *Bacillus subtilis*, *Aspergillus niger*, *Aspergillus flavus*, *Aspergillus fumigates* and *Candida albicans* by agar plate(well) method [54].

**Perumal Samy and Ignacimuthu, 2000;** studied the antibacterial activity of various extracts of leaf by disc diffusion method and found methanol extract active against *B. subtilis*, *S. aureus* and *E. coli* at 10 mg/ml concentration [55].

**Qureshi et al., 1997;** studied the sensitivity of the keratinophilic fungi in *P. daemia* extract by dry-weight method [56].

**Valsaraj et al.,1997;** studied the antibacterial activity of ethanol (80%) extract of leaf and stem and exhibited activity against various bacteria strain such as *Pseudomonas aeruginosa*, *Bacillus subtilis*, *Escherichia coli* and *Staphylococcus aureus* at a concentration of 25 mg/ml, while ethanol (95%) extract of plant inactive against *Mycobacterium tuberculosis* by agar plate method [57].

**Elango et al., 1985;** studied the antibacterial activity of ethanol (80%) extract of whole plant and it was exhibited activity at a concentration of 80  $\mu\text{g/ml}$  against *Proteus mirabilis*, *Escherichia coli*, *Staphylococcus aureus* and *Pseudomonas aeruginosa*, agar disc diffusion method. 1 to 3 mg of the extract showed cardiovascular effects on heart of frog, while the high dose of extract was blocked heart. The extract 10 mg/kg showed uterine stimulant effect on female guinea pig, while 1 to 2.5 mg of the extract exhibited smooth muscle stimulant activity on guinea pig ileum [25].

**Runnebaum et al., 1984;** reported the ethanol extract of leaf not showed anti-implantation and abortifacient effect on female pregnant rat at a dose level of 200 mg/kg [58].

**Prakash et al., 1978;** reported aqueous ethanolic extract of leaf not showed embryotoxic effect on pregnant rat (related to fertility regulation), at a dose of 100 mg/kg [59].

**Anonymous, 1976:** reported aqueous ethanolic extract of fruit, leaf and stem exhibited cytotoxicity (CA-9KB) by cell culture method and the ED<sub>50</sub> was 20µg/ml<sup>[60]</sup>.

**Ogunlana and Ramstad, 1975;** reported the 50% methanol (1:1) extract of flower and leaf inactive against various bacteria including *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Staphylococcus albus*, *Bacillus subtilis* and *Proteus species* by broth culture method<sup>[61]</sup>.

**Dhar et al., 1973;** studied the toxicity (LD<sub>50</sub>) of aqueous ethanolic extract of whole plant and it was showed more than 1 gm/kg on mouse. Cytotoxic activity (CA-9KB) of the extract inactive (ED<sub>50</sub>> mcg/ml) and hypotensive activity of the extract showed active at 50 mg/kg (IV route) dog, while antispasmodic activity of the extract tested on guinea pig ileum and hypothermic activity of the extract showed active at dose level of 500 mg/kg (mouse, IP)<sup>[62]</sup>.

**Ghatak and De, 1961;** studied the toxicity assessment (LD<sub>50</sub>) of ethanol extract of whole plant and it was found to be 6, 5, 6 and 4 mg/kg, IV on guinea pig, rabbit, dog and cat respectively<sup>[31]</sup>.

**Ghatak and De, 1961;** reported ethanol extract of whole plant was showed uterine stimulant effect on normal and pregnant female animals such as rat, rabbit, guinea pig, and cat at a dose of 1-2 mg /kg, IV and also it showed intestine smooth muscle stimulant activity on cat, rabbit and guinea pig at a dose of 0.067 mg/ml, while bronchoconstrictor and respiratory depressant activities active at a dose of 2 mg/kg, IV on cat, rabbit and guinea pig. The ethanol extract of whole plant (0.1 mg/ml) exhibited cardiac depressant activity on rabbit and guinea pig heart (Perfusion method) and it was showed hypertensive activity at dose of 1-2 mg/kg, IV on dog and cat<sup>[31]</sup>.

**Rakhit et al., 1959;** reported the aqueous extract of whole plant was showed uterine stimulant effect on female guinea pig<sup>[63]</sup>.

**Gupta et al., 1950;** reported alkaloid fraction of the whole plant was showed uterine stimulant effect on female cat (0.75 mg / animal, IV), while the same fraction was exhibited intestine smooth stimulant activity on cat (3mg/kg, IV) and also hyperglycemic activity on monkey (3mg/kg, IV). Oral dose of 3 mg/ kg on cat was showed gastric secretory stimulation activity<sup>[64]</sup>.

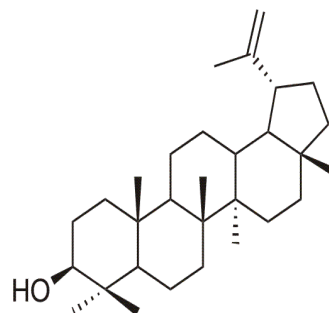
**Heal et al., 1950;** studied the insecticide activity of aqueous extract of entire plant and it was exhibited strong insecticide activity against *Periplaneta Americana*, *Blatella germanica* and *Oncopeltus fasciatus* at a dose level of 40 ml/kg<sup>[65]</sup>.

**Dutta and Ghosh, 1947a;** reported aqueous extract of entire plant was showed uterine stimulant effect on female cat<sup>[23]</sup>.

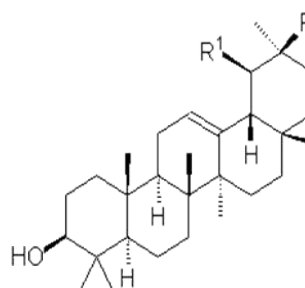
**Dutta and Ghosh, 1947b;** reported ethanol (95%) extract of entire plant was showed uterine stimulant effect on female rat<sup>[36]</sup>.

### Phytochemistry

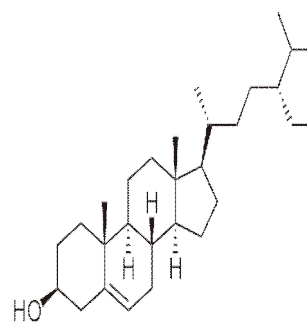
**Anjaneyulu et al., 1998; Seshadri and Vydeeswaran,1971; Rakhit et al., 1959; Raman and Barua, 1958;** reported to contain β-sitosterol, lupeol, lupeol acetate, α, β-amyrin and its acetate in entire plant and root<sup>[18, 28, 63, 66]</sup>.



Lupeol

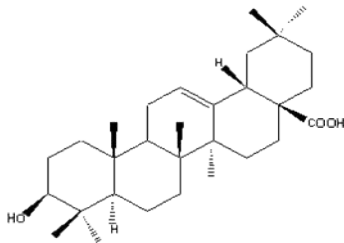


R = H, R<sup>1</sup> = CH<sub>3</sub> = α-amyrin  
R = CH<sub>3</sub>, R<sup>1</sup> = H = β-amyrin

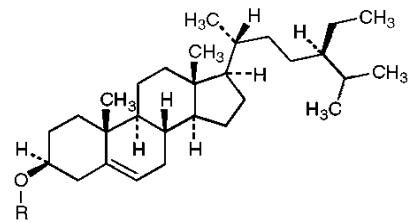


β-sitosterol

*Anjaneyulu et al., 1998*; isolated lupeol-3-beta trans crotonate and oleanolic acid acetate from dried whole plant [18].



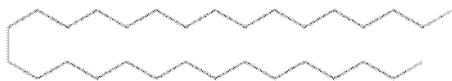
Oleanolic acid



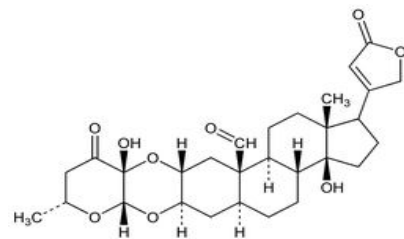
Daucosterol

*Mittal et al., 1962*; reported to contain various cardenolide such as calotoxin, calotropagenin, dihydro calotropagenin, calotropin and uscharidin in seed, while coroglaucigenin, corotoxigenin, uscharidin and uzarigenin in stem [33].

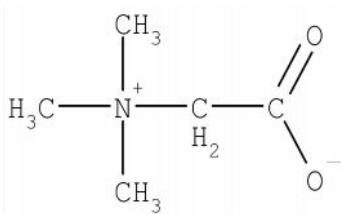
*Rakhit et al., 1959*; isolated betaine, hentriacontane and pentacosanoic acid from entire plant, while reported to contain magnesium and potassium carbonate, daemia extensa polypeptide, Ca, Mg and K oxalate [63].



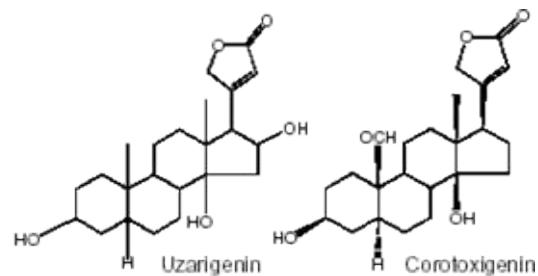
Hentriacontane



Uscharidin



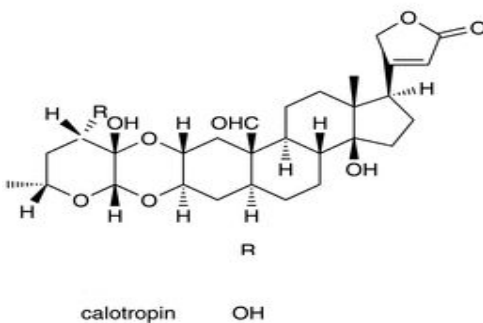
Betaine



Uzarigenin

Corotoxigenin

*Seshadri and Vydeswaran, 1971*; reported to contain calactin, calotropin, corotoxigenin, daucosterol and sucrose in root [28].

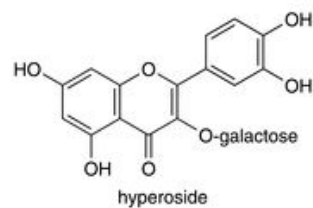


calotropin

OH

*Dutta and Ghosh, 1947a, b*; reported to contain daemia extensa polypeptide, daemia extensa glucoside, Inorganic salts such as KCl and KNO<sub>3</sub> in entire plant [23, 36].

*Sinha and Dogra, 1985; Sankara Subramanian and Nair 1968*; reported to contain hyperoside (flavonol) in dried stem, while flavonoids and saponins in fresh shoots and flowers [67, 68].



hyperoside

## CONCLUSION

The plant *P. daemia* (Veliparuthi) has a wide array of pharmacological activities. It is widely used in various traditional system of medicine as a medicine. It has been used since centuries as an analgesic, antipyretic and anti-inflammatory, abortifacient, in treatment of diarrhea and malarial intermittent fever. Recent research carried out

indicates its other uses such as hepatoprotective, antifertility and anti-diabetic. The plant *P. daemia* is an important source of various types of compounds with diverse chemical structures as well as pharmacological activities. However, very less work has been done on this plant and there is a wide scope for investigation.

**Figure 1. Aerial Parts of *P. daemia***



**Figure 2. Roots of *P. daemia***





## REFERENCES

1. WHO, IUCN, WWF. Guidelines on the conservation of medicinal plants. Switzerland: IUCN Gland; 1993.
2. Kotnis MS, Patel P, Menon SN, Sane RT. Rene protective effect of *Hemidesmus indicus*, a herbal drug used in Gentomicin-induced renal toxicity. *Nephrology (Carlton)* 2004; 3: 142-152.
3. Bhagwati Uniyal. Utilization of medicinal plants by the rural women of Kulu, Himachal Pradesh. *Indian J Trad Knowledge* 2003; 2(4): 366-370.
4. Dahanukar SA, Kulkarni AR, Rege NN. Pharmacology of medicinal plants and natural products. *Indian J Pharmacol* 2000; 32: S81-S118.
5. Harsha VH, Hebbar SS, Hegde GR, Shripatti V. Ethnomedical knowledge of plants used by Kunabi tribe of Karnataka in India. *Fitoterapia* 2002; 73(4): 281-287.
6. Cragg GM, Newman DJ, Sander KM. Natural Products in Drug Discovery and Development. *J.Nat Prod* 1997; 60: 52-60.
7. Khare CP. *Indian Medicinal Plants (An Illustrated Dictionary)*. New York: Springer Science and Business Media; 2007.p. 472.
8. Kirtikar KR, Basu BD. *Indian Medicinal Plants, Vol.III*. Dehardun: International Book Distributors; 1999.p. 1616 – 1617.
9. Nadkarani Ak. *Indian Materia Medica, Vol.I*. Bombay; Popular Prakashan Pvt Ltd; 1976. p. 430.
10. Anonymous. *Indian Medicinal Plants (a compendium of 500 species)*, Vol.IV. Hyderabad: Orient Longman Ltd; 1995. p. 386 -389.
11. Hebbar SS, Harsha VH, Shripathi V, Hegde GR. Ethnomedicine of Dharward district in Karnataka, India-plants used in oral health care. *J Ethnopharmacol* 2004; 94: 261-266.
12. Dokosi OB. *Herbs of Ghana*. Accra: Ghana University Press; 1998. p. 313 -314.
13. Bruce TBF. *Personal communications-phytotherapist*. Accra: Ghana; 1998&2000.
14. Sureshkumar SV, Mishra SH. Hepatoprotective effect of extracts of *Pergularia daemia* Forsk. *J Ethnopharmacol* 2006; 107: 164-168.
15. Golam Sadik, Gafur MA, Shah Alam Bhuiyan M, Khurshid Alam AHM, Helal U Biswas M, Parvez Hassan, Abdul Mannan, Omar Faruk Khan M, Chowdhury AKA. Antifertility activity of *P. daemia*. *The Sciences* 2001a; 1(1): 22-24.
16. Wahi AK, Ravi J, Hemalatha S, Singh PN. Anti diabetic activity of *Daemia extensa*. *J Nat Remed* 2002; 2(1): 80-83.
17. Sathish CJ, Sharma RA, Jain R, Macalo N, Capasso F, Vijayvergia R, Mittal C. Ethnopharmacological evaluation of *P. daemia* (Forsk.) Chivo. *Phytother Res* 1998; 12: 378-380.
18. Anjaneyulu ASN, Raju DVSN, Srinivasa Rao S. Chemical evaluation of *P. daemia*. *Indian J Chem* 1998; 37B: 318-320.
19. Parrotta A John. *Healing plants of peninsular India*. Wallingford, U K: CABI Publishing; 2001. p. 131-132.
20. Rastogi RP and Mehrotra BN. *Compendium of Indian medicinal plants. Vol2 (1970-1979)*. New Delhi, India: Central Drug Research, Lucknow and NISCAIR; 2006. p. 521.
21. <http://plants.usda.gov>
22. Selvanayagam ZE, Gnavavendhan SG, Balakrishna K, Rao RB, Ali SU. Survey of medicinal plants with anti-snake venom activity in Chengalpattu district, Tamilnadu, India. *Fitoterapia* 1995; 66 (6): 488-494.
23. Dutta A, Ghosh S. Chemical examination of *Daemia extensa*. I. *J Amer Pharm Ass Sci Ed* 1947a; 36: 250-252.
24. Berhault J. *Flore illustree Du Senegal. I. Dicotyls (Acanthaceae-Avicenniaceae)*. Dakar ; Govt Senegal: 1971.
25. Elango V, Ambujavalli L, Amala Basker E, Sulochana N. Pharmacological and microbiological studies on *Pergularia extensa*. *Fitoterapia* 1985; 56 (5): 300-302.
26. Singh VP, Sharma SK, Khare VS. Medicinal plants from Ujjain district Madhya Pradesh - part II. *Indian Drugs Pharm Ind* 1980;5: 7-12.
27. Arseculeratne SN, Gunatilaka AAL, Panabokke RG. Studies on medicinal plants of Srilanka. Part 14: Toxicity of some traditional medicinal herbs. *J Ethnopharmacol* 1985; 13 (3): 323-3.
28. Seshadri TR, Vydeeswaran S. Chemical constituents of *Daemia extensa* (roots). *Curr sci* 1971; 40: 594-595.
29. Selvanayahgam ZE, Gnavavendhan SG, Balakrishna K, Rao RB. Anti-snake venom botanicals from ethnomedicine. *J Herbs Spices Med Plants* 1994; 2 (4): 45-100.
30. Gupta JC, Roy PK, Dutta A. Pharmacological action of an active constituent isolated from *Daemia extensa* (syn. *Pergularia extensa*). *Indian J Med Res* 1946; 34: 181.
31. Ghatak BJR, De NN. Pharmacological action of *Daemia extensa*. *J Sci Ind Res-C* 1961; 20: 51-53.

32. Girach RD, Aminuddin, Siddioui PA, Khan SA. Traditional plant remedies among the Kondh of district Dhenkanal (orissa). *Int J Pharmacog* 1994; 32 (3): 274-283.
33. Mittal OP, Tammz C, Reichstein T. Glycosides and aglycons. CCXXVII. The glycosides of *Pergularia extensa*. *Helv Chim Acta* 1962; 45: 907.
34. Kakrani HKN, Saluja AK. Traditional treatment through gerbs in Kutch district, Gujarat state, India. Part II. Analgesic, anti-inflammatory, antirheumatic, antiarthritic plants. *Fitoterapia* 1994; 65 (5): 427-430.
35. De laszlo H, Henshaw PS. Plant materials used by primitive peoples to affect fertility. *Science* 1954; 119: 626-631.
36. Dutta A, Ghosh S. *Daemia extensa*. *Indian J Pharmacy* 1947b; 9: 58-60.
37. Pushpangadan P, Atal CK. Ethno-medico-botanical investigations in Kerala I. Some primitive tribals of Western Ghats and their herbal medicine. *J Ethnopharmacol* 1984; 11 (1): 59-77.
38. Reddy MB, Reddy KR, Reddy MN. A Survey of plant crude drugs of Anantapur district, Andhra Pradesh, India. *Int J Crude Drug Res* 1989; 27(3): 145-155.
39. Watt JM, Breyer-Brandwijk MG. The medicinal and poisonous plants of Southern and Eastern Africa. 2nd ed. London: E. + S. Livingstone, Ltd; 1962.
40. Reddy MB, Reddy KR, Reddy MN. A survey of medicinal plants of Chenchu tribes of Andhra Pradesh, India. *Int J Crude Drug Res* 1988; 26(4): 189-196.
41. John D. One hundred useful raw drugs of the Kani tribes of Trivandrum forest division, Kerala, India. *Int J Crude Drug Res* 1984; 22(1): 17-39.
42. Kokwaro JO. Medicinal plants of East Africa. Nairobi: East Africa Literature Bureau; (1976).
43. Kokwaro JO. A review of research on plants for fertility regulation in Africa. Proc who symposium on plant-derived products for fertility regulation. Seoul, Korea. February 1981. p. 8.
44. Kokwaro JO. A review of research on plants for fertility regulation in Africa. *Korean J Pharmacog* 1981; 12(3): 149-152.
45. Samuelsson G, Farah MH, Claeson P, Hagos M, Thulin M, Hedberg O, Warfa AM, Hassan AO, Elmi AH, Abdurahman AD, Elmi AS, Abdi YA, Alin MH. Inventory of plants used in traditional medicine in Somalia. I. Plants of the families Acanthaceae-Chenopodiaceae. *J Ethnopharmacol* 1991; 35(1): 25-63.
46. Nagaraju N, Rao KN. A survey of plant crude drugs of Rayalaseema, Andhra Pradesh, India. *J Ethnopharmacol* 1990; 29(2): 137-158.
47. Kohler I, Jenett siems K, Kraft C, Siems K, Abbiw D, Bienzle U, Eich E. Herbal remedies traditionally used against malaria in Ghana: Bioassay-guided fractionation of *Microglossa pyridolia* (asteraceae). *Z Nature Forsch Ser C* 2002; 57C (11/12): 1022-1027.
48. Gill LS, Akinwumi C. Nigerian folk medicine: Practices and beliefs of the Ondo people. *J Ethnopharmacol* 1986; 18(3): 259-266.
49. Suresh Kumar SV, Mishra SH. Hepatoprotective effect of *P. daemia* (Forsk.) ethanol extract and its fraction. *Indian J Exp Biol.* 2008a Jun; 46(6): 447-52.
50. Suresh Kumar SV, Mishra SH. In-vitro evaluation of hepatoprotective activity of *P. daemia* Forsk. *Pharma Mag* 2008b; Vol 4(16)8: 298-302.
51. Flyman MV, Afolayan AJ. The implication of the mineral ratios of *Cucumis myriocarpus* Naud. and *P. daemia* (Forsk.) Chiov. in human diets. *J Med Food.* 2007 Sep; 10(3): 548-51.
52. Suresh Kumar SV, Mishra SH. Hepatoprotective activity of extracts from *P. daemia* Forsk. against carbon tetrachloride-induced toxicity in rats. *Pharma Mag* 2007; Vol 3(11): 187-191.
53. Golam Sadik MD, Gafur MA, Shah Alam Bhuiyan M, Motiur Rahman, Helal U Biswas. Antifertility Activity of the alkaloidal fraction of *P. daemia*. *The Sciences* 2001b; 1(4): 217-219.
54. Srinivasan D, Nathan S, Suresh T, Perumalsamy PL. Antimicrobial activity of certain Indian medicinal plants used in folkloric medicine. *J Ethnopharmacol* 2001; 74: 217-220.
55. Perumal Samy R, Ignacimuthu S. Antibacterial activity of some folklore plants used by tribals in Western Ghats of India. *J Ehanopharmacol* 2000; 69: 63-71.
56. Qureshi S, Rai MK, Agrawal SC. In vitro evaluation of inhibitory nature of extracts of 18-plant species of Chhindwara against 3-keratinophilic fungi. *Hindustan Antibiot Bull.* 1997 Feb-Nov; 39(1-4): 56-60.
57. Valsaraj R, Pushpangadan P, Smitt UW, Adsersen A, Nyman U. Antimicrobial screening of selected medicinal plants from India. *J Ethnopharmacol* 1997; 58(2): 75-83.
58. Runnebaum B, Rabe T, Kiesel L, Prakash AO. Biological evaluation of some medicinal plant extracts for contraceptive efficacy in females. Future aspects in contraception. Part 2. Female contraception. Boston, USA: MTP press, Ltd; 1984. p. 115-128.
59. Prakash AO, Gupta RB, Mathur R. Effect of oral administration of forty-two indigenous plant extracts on early and late pregnancy in albino rats. *Probe* 1978; 17(4): 315-323.



60. Anonymous. Unpublished data. National Cancer Institute: Nat cancer inst central files; 1976.
61. Ogunlana EO, Ramstad E. Investigations into the antibacterial activities of local plants. *Planta med* 1975; 27: 354.
62. Dhar ML, Dhar MN, Dhawan BN, Mehrotra BN, Srimal RC, Tandon JS. Screening of Indian plants for biological activity. Part IV. *Indian J Exp Biol* 1973; 11: 43-54.
63. Rakhit S, Dhar MM, Anand N, Dhar ML. Chemical investigations on *Daemia extensa*. *J Sci Ind Res-B* 1959; 18: 422-426.
64. Gupta JC, Roy PK, Ray GK. Pharmacological action of an active constituent isolated from *Daemia extensa* (*Pergularia extensa*). Part II.. Further Comparative study with pituitrin. *Indian J Med Res* 1950; 38: 75-82.
65. Heal RE, Rogers EF, Wallace RT, Starnes O. A survey of plants for insecticidal activity. *Lloydia* 1950; 13(1): 89-162.
66. Raman SP, Barua AK. Chemical investigation of *Daemia extensa*. *J Amer Pharm Ass Sci Ed* 1958; 47: 559-560.
67. Sinha SKP, Dogra JVV. A survey of the plants of Bhagalpur and Santhal pargana for saponins, flavonoids and alkaloids. *Int J Crude Drug Res* 1985; 23(2): 77-86.
68. Sankara Subramanian S, Nair AGR. Flavonoids of some asclepiadaceous plants. *Phytochemistry* 1968; 7: 1703-1704.

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