



Green Synthesized Nanoparticles & An approach towards Antibacterial & Antimicrobial activities: A Review

**Bhabani Shankar Panda^{1*}, Mohammed Ansar Ahemad²
and Laxmi Narayan Mishra³**

^{*1}Coastal Laboratory, State Pollution Control Board, ICZMP, Bhubaneswar, Odisha, India

²Gandhi Institute For Education and Technology, Bhubaneswar, Odisha, India

³Paramananda College, Bolgarh, Khordha, Odisha, India

Abstract : Nanoscience has reformed nearly in every field of human life inferable from the one of a kind and astounding physiochemical, electrical and mechanical properties of nano-sized materials. The quantum repression impacts and large accessible dynamic surface areas are accepted to be the key components to the improved usefulness of nanostructures. The synthesis of nanomaterials or nano particles are dependent upon the particular applications, then they can suffer from some challenges such as stability in hostile environment, at that point they can experience in fundamental mechanism and modelling factors, toxicity features, expansive analysis supplies, need for skilled operatives, problem in devices accumulating and structures etc. To counter those impediments, a novel period of ‘green synthesis’ methods is gaining increasing extraordinary consideration in flow innovative work on materials science and technology. Mostly, green synthesis of nanomaterials, formed by control, fresh up, regulation and remediation method will directly help elevate their ecological cordiality. Nano-Particles have progressively been utilized in industry in the course of recent decades with utilizations shifting from food additives to drug management. Different investigations have been completed to enhance antimicrobial capacities as a result of the becoming microbial opposition towards basic germicide and antibiotics.

Keywords : Green synthesis, Antimicrobial capacities, Nano-Particles, Antibacterial, Toxicity features.

Introduction

In the most recent decade, novel synthesis methods for nanomaterials like metal nanoparticles, quantum sports or quantum dots (QDs), graphene, and their composites have been a fascinating region with regards to nanoscience^[1-8]. Nanoscience has reformed nearly in every field of human life inferable from the one of a kind and astounding physiochemical, electrical and mechanical properties of nano-sized materials. The quantum

repression impacts and large accessible dynamic surface areas are accepted to be the key components to the improved usefulness of nanostructures. These properties make them appropriate for different biomedical applications^[9-13]. Generally, there are two types of basic techniques or standards happens for synthesis of nanoparticles; topdown and bottom up methods as shows in literature(Fig.1).In relations of the size of objects, both methods are quite similar. Both the methods incline to meet in relations of the size variety of objects. In past the nanoparticles are prepared through different scope of synthesis methods like mechanical milling, faltering, lithographic and chemical etching^[14].

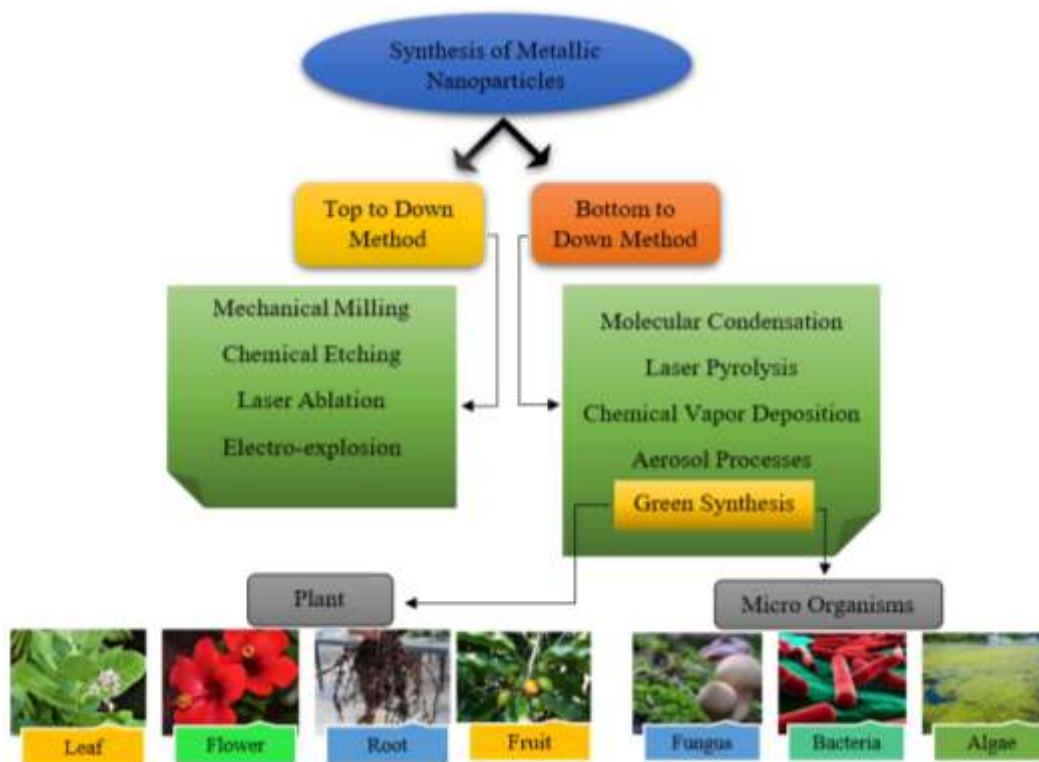


Fig. 1 Different synthesis methods accessible for the preparation of metallic nanoparticles

The synthesis of nanomaterials or nano particles are dependent upon the particular applications, then they can suffer from some challenges such as stability in hostile environment, at that point they can experience in fundamental mechanism and modelling factors, toxicity features, expansive analysis supplies, need for skilled operatives, problem in devices accumulating and structures etc.which demonstrates that the properties, conduct, and sorts of nanomaterials ought to be improved to meet the previously mentioned focuses. To counter those impediments, a novel period of ‘green synthesis’ methods is gaining increasing extraordinary consideration in flow innovative work on materials science and technology. Mostly, green synthesis of nanomaterials, formed by control, fresh up,regulation and remediation method will directly help elevate their ecological cordiality.

By Green synthesis process nanoparticles are produced through plants, bacteria, fungi, and algae takes into the enormous scope of formation of metal oxide nanoparticles free of filths^[15].These are required to stay away from the creation of undesirable or hurtful results through the development of dependable, reasonable, and eco-accommodating amalgamation techniques. The organic systems or perfect dissolvable solvents are used essentially to achieve this goal. These processes dependent on some biological precursors; which are depend on several reaction parameters such as conditions of pH, solvent, pressure and temperature.

Mostly in case of the metal oxide or metal nanoparticles synthesis, plant biodiversity has been considered for the accessibility of powerful phytochemicals in different plant separates, particularly in leaves, for example, ketones, aldehydes, flavones, amides, terpenoids, carboxylic acids, phenols, and ascorbic acids.These components are equipped for diminishing metal salts into metal nanoparticles^[16].

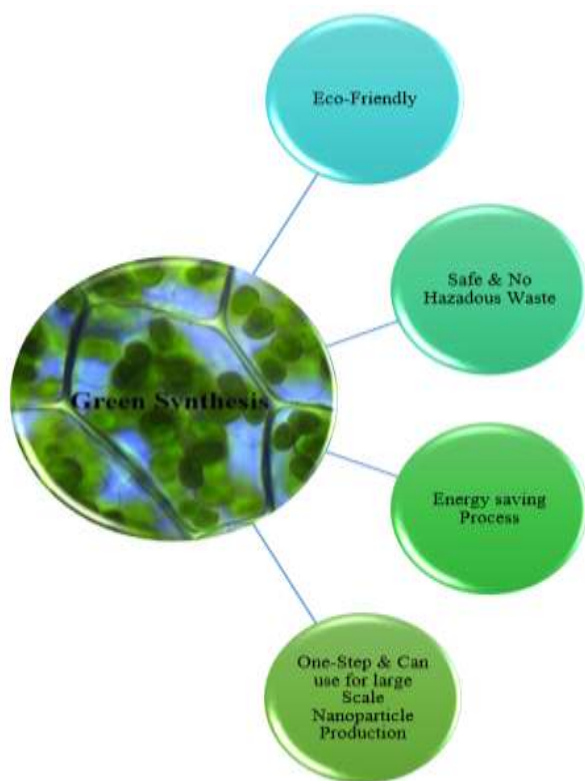


Fig. 2 Advantages of green synthesis methods

This review is about the summarization of the process (green synthesis) of metal oxide or metal nanoparticles with their focal points. Also, the role of different natural extracts of plant, algae, bacteria and fungi points of interest over other traditional segments or solvents were discussed here. The main aim of this literature study is to give an efficiently mechanisms of green synthesis procedures and their correlated segments that are profitable for researchers; Who associated with this developing field and a specific approach towards antibacterial as well as antimicrobial activities.

Generally green synthesis is of two types i.e. Solvent system-based synthesis and Biological component-based synthesis.

Solvent system-based synthesis

Water is the least expensive and most normally available dissolvable on earth. Since the appearance of nanoscience and nanotechnology, the utilization of water as a dissolvable for the union of different nanoparticles has been completed. For example, synthesis of Au and Ag nanoparticles at room temperature by using gallic acid a bifunctional atom, in a watery medium^[17]. Gold nanoparticles were delivered by means of a laser removal procedure in an aqueous solution^[18].

Some metallic nanoparticles like Al, Au, Te, Ag, Ru and Pt have been integrated in ionic fluids^[19–22]. The procedure of nanoparticle synthesis is shortened since can fill in as both a reductant and a defensive agent. Ionic Liquids can be hydrophilic or hydrophobic relying upon the idea of the cations and anions^[21, 23–26]. Lazarus et al. integrated silver nanoparticles in an ionic fluid. For the first time, Kim et al. built up a one-phase arrangement method for gold (Au) and platinum (Pt) nanoparticles by means of thiol-functionalized ionic liquids (TFILs). TFILs acted as an alleviating agent to create crystalline structures with little sizes^[28]. Bussamara et al. have made a virtual study by controlling the amalgamation of manganese oxide (Mn₃O₄) nanoparticles by means of imidazolium ionic liquids and a conventional dissolvable i.e. oleyl amine^[27]. The innumerable ILs are utilized to incorporate different metallic nanoparticles as recorded in Table 1. Sue et al. recommended that diminishing the solubility of metal oxides around the critical point can cause super inundation and a definitive arrangement of nanoparticles^[29].

Table-1:Green synthesis of metallic nanoparticles from several plant extracts

Plant origin	Nanoparticle	Size (in n.m.)	Shape	Applications	References
<i>Azadirachta indica</i> (neem)	Gold, silver and silver-gold alloys	5–35 and 50–100	Spherical, triangular or hexagonal	Detection of toxic Metals	[30]
<i>Aloe barbadensis</i> Miller (Aloe vera)	Indium oxide	5–50	Spherical	Solar cells, gas sensors	[31]
<i>Acalypha indica</i>	Silver	20–30	Spherical	Antibacterial activity against water borne pathogens	[32]
Apiin extracted from henna leaves	Silver and gold	39	Spherical, triangular or quasi-spherical	Hyperthermia of cancer cells and IR-absorbing optical coatings	[33]
<i>Avena sativa</i> (oat)	Gold	5–20	Rod-shaped	–	[34]
<i>Camellia sinensis</i> (blacktea leaf extracts)	Gold and silver	20	Spherical, prism	Catalysts, sensors	[35]
<i>Cinnamomum camphora</i> (camphor tree)	Gold and silver	55–80	Triangular, spherical (Au), and quasi-spherical(Ag)	–	[36]
<i>Coriandrum sativum</i> (coriander)	Gold	6.75–57.91	Spherical, triangular, truncated triangular or decahedral	Drug delivery, tissue/ tumour imaging, photo-thermal therapy	[37]
<i>Cymbopogon flexuosus</i> (lemongrass)	Gold	200–500	Spherical or triangular	Infrared-absorbing optical coatings	[38]
<i>Diospyros kaki</i> (persim-mon)	bimetallic gold/silver	50–500	Cubic	–	[39]
<i>Eucalyptus citriodora</i> (neelagiri)	Silver	20	Spherical	Antibacterial	[40]
<i>Eucalyptus hybrida</i> (safeda)	Silver	50–150	Crystalline or spherical	–	[41]
<i>Garcinia mangostana</i> (mangosteen)	Silver	35	Spherical	Antimicrobial activity against E. coli and Sauers	[42]
<i>Gardenia jasminoides</i> Ellis (gardenia)	Palladium	3–5	–	Nano catalysts for p-nitrotoluene hydrogenation	[43]
<i>Syzygium aromaticum</i> (clove buds)	Gold	5-100	Irregular	Detection and destruction of cancer cells	[44]
<i>Medicago sativa</i> (alfalfa)	Gold	2–40	Irregular, tetrahedral, hexagonal platelet, decahedral or icosahedral	Labelling in structural biology, paints	[45-47]
<i>Mentha piperita</i>	Silver	5–30	Spherical	To kill microbes	[48]

(pep-permint)					
<i>Medicago sativa</i> (alfalfa)	Iron oxide	2–10	Crystalline	Cancer hyperthermia, drug delivery	[49]
<i>Morus</i> (mulberry)	Silver	15–20	Spherical	Antimicrobial activity against <i>E. coli</i> , <i>B.</i> <i>subtilis</i>	[50]
<i>Ocimum sanctum</i> (tulsi; root extract)	Silver	10 ± 2 and 5 ± 1.5 nm	Spherical	Catalytic reduction	[51]
<i>Pear fruit extract</i>	Gold	200–500	Triangular or hexagonal	Catalysis, biosensing	[52]
<i>Pelargonium roseum</i> (rose geranium)	Gold	2.5–27.5	Crystalline	–	[53]
<i>Psidium guajava</i> (guava)	Gold	25–30	Spherical	–	[54]
<i>Sedum alfredii Hance</i>	Zinc oxide	53.7	Hexagonal wurtzite and pseudo- spherical	Nanoelectronics	[55]
<i>Ocimum sanctum</i> (tulsi; leaf extract)	Gold and silver	30 and 10– 20	Crystalline, hexagonal, triangular and spherical	Biolabeling, biosensor	[56]

Ag nano-particles synthesized from extraction of tea leaf were found to be unchanging after inflowing the aquatic environmental condition^[57]. In similar manner, the constancy of Ag nano-particles in presence of aqueous medium; produced utilizing plant extracts and plant metabolites was confirmed from the subsequent material^[58]. The nature and steadiness of nanoparticles were hypothetically forecast through anticipated through an unthinking comprehension of the surface complexation processes^[59]. The colloidal constancy of nanoparticles can be managed by controlling the molecule size and surface topping or through functionalization strategies or techniques^[60, 61]. Aside from surface science, other key auxiliary highlights deciding the nanomaterial poisonousness are the size, shape, and arrangement of the nanomaterials^[62].

Plant leaf extract-based mechanism

For synthesis of nanoparticle facilitated by plant leaf extracts, is assorted with metal antecedent solutions at various response conditions^[63]. The parameters defining the circumstances of the leaf extracts like types of phytochemicals, metal salt concentration, temperature, pH, and phytochemical concentration; are confessed to control the pace of nanoparticle arrangement just as their yield and dependability^[64]. The plant leaf extract configuration is also a significant aspect in synthesis of nanoparticles, for example various plants contain fluctuating focus levels of phytochemicals^[65, 66]. The fundamental phytochemicals present in plants are flavones, terpenoids, sugars, ketones, amides, aldehydes and carboxylic acids which are accountable for bio-reduction of nanoparticles^[67].

FT-IR analysis of green synthesized nanoparticles with the help of plants extract inveterate that embryonic nanoparticles were frequently found to be related with proteins^[69]. Amino acids have various roles of tumbling the metal ions. Plant extracts are comprised of proteins biomolecules and carbohydrates, which performance as a reducing agent to endorse the development of metallic nanoparticles^[68]. Kesharwani et al.^[71] sheltered photographic films utilizing an emulsion of silver bromide. At the point when light hit the film, the silver bromide was sharpened; this uncovered film was put into a solution of hydroquinone, which was additionally oxidized to quinone by the activity of silver particle or ion as shown in fig.3. The silver particle was converted to silver metal, which stayed in the emulsion.

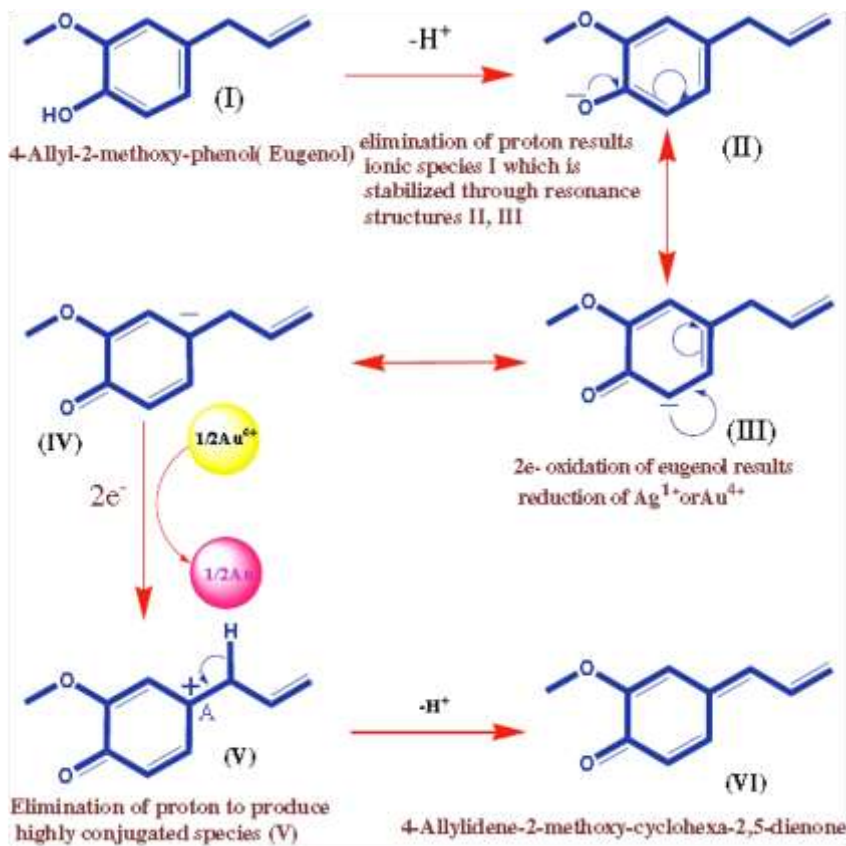


Fig. 3 Reactions for the reduction of Au and Ag particles^[70]

FT-IR information presented that the withdrawing groups ($-OH$ groups) initiating from eugenol vanish during Au and Ag nanoparticles arrangement. Alkenes, carbonyl, and chloride functional groups seemed after the development of Au nanoparticles. Some other groups such as $-OH$ (aqueous) and $R-CH$ were also discovered after and before the manufacture of Au nanoparticles^[70]. Thus, they proposed the conceivable chemical mechanism or approach as shown in Fig. 4.

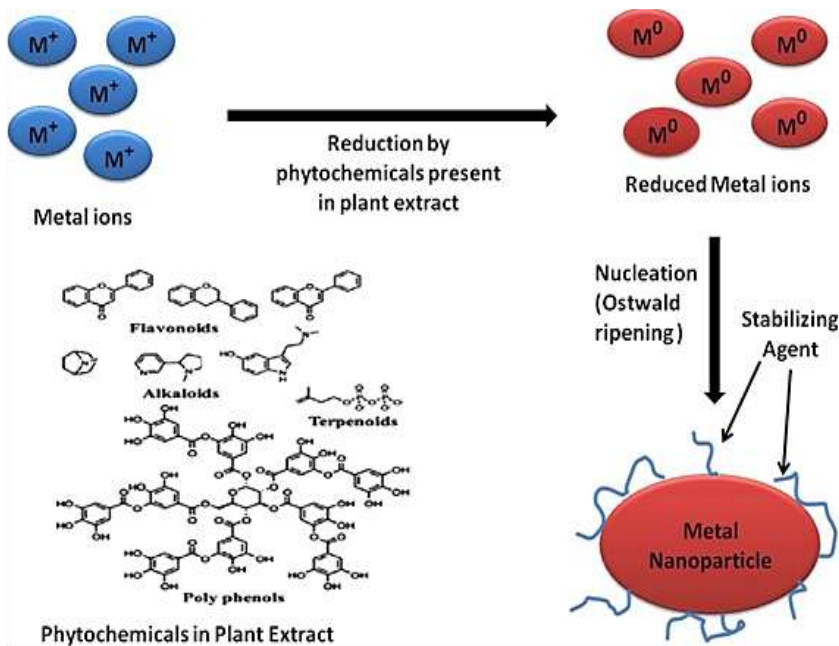


Fig. 4 Mechanism of nanoparticle formation by leaf extracts^[72]

Biological components-based synthesis

Generally chemical synthesis methodologies require high radiation, highly toxic reductants and alleviating agents, which can make malicious impacts to both humans and marine life. Interestingly, green combination of metallic nanoparticles is a one pot or single step eco-friendly bio-reduction technique that requires moderately low energy to start the reaction. This reduction technique is also cost effective^[73-79].

Bacteria

Bacterial species have been used for marketable biotechnological applications like bioremediation, genetic engineering and bioleaching^[80]. Microorganisms have the capacity to lessen metal particles and are significant candidates in preparation of nanoparticles^[81]. Prokaryotic microorganisms and actinomycetes have been extensively utilized for synthesizing metal or metal oxide nanoparticles. The bacterial synthesis of nanoparticles has been embraced because of the overall ease of controlling the bacteria such as *Escherichia coli*, *Lactobacillus casei*, *Bacillus cereus*, *Aeromonas sp.* etc.^[82].

Table-2: Synthesis of metallic nano-particles from various Bacterial species

Species	Nanoparticles	Size (in n.m.)	Shapes	Application	References
<i>E. coli</i>	Cadmium sulfide	2–5	Fluorescent labels	Wurtzite structures	[83]
<i>Bacillus cereus</i>	Silver	20–40	Spherical	Antibacterial activity against <i>Escherichia coli</i> , <i>Pseudomonas aeruginosa</i> , <i>Staphylococcus aureus</i> , <i>Salmonella typhi</i> and <i>Klebsiella pneumoniae</i> bacteria	[84]
<i>Pseudomonas proteolytica</i> , <i>Bacillus cecembensis</i>	Silver	6-13	Spherical	Antibacterial activity against <i>A. kerguelensis</i> , <i>A. gangotriensis</i> , <i>B. indicus</i> , <i>P. antarctica</i> , <i>P. proteolytica</i> , and <i>E. coli</i>	[85]
<i>Aquaspirillum magnetotacticum</i>	Iron Oxide	40–50	Octahedral prism	–	[86]
<i>Klebsiella aerogenes</i>	Cadmium sulfide	20–200	–	–	[87]
<i>Lactobacillus casei</i>	Silver	20–50	Spherical	Drug delivery, cancer treatments, bio-labeling	[88]
<i>E. coli DH 5α</i>	Gold	8–25	Spherical	Direct electrochemistry of hemoglobin	[89]
<i>Klebsiella pneumoniae</i> ,	Silver	28–122	Spherical	Optical receptors, electrical	[90]
<i>Bacillus megaterium D01</i>	Gold	< 2.5	Spherical	Catalysis, biosensing	[91]
<i>Desulfovibrio desulfuricans</i>	Gold	20–50	Spherical	Catalysis	[92]
<i>Rhodospseudomonas capsulate</i>	Gold	10–20	Cancer hyperthermia	Triangular	[93]
<i>Magnetospirillum magneto-tacticum</i>	Iron Oxide	47	–	Handle shaped cluster	[94]

Fungus

Fungus interceded biosynthesis of metal or metal oxide nanoparticles is likewise a very effectual process for the age of monodispersed nanoparticles with well-defined morphologies. These are going about as better natural operators for the arrangement of metal and metal oxide nanoparticles, because of the nearness of an assortment of intracellular enzyme^[95]. Competent fungi can incorporate bigger measures of nanoparticles contrasted with microbes or bacteria^[96]. Additionally, fungi have numerous benefits over different life forms because of the nearness of catalysts or lessening parts on their cell surfaces^[97]. Several fungal species are utilized to blend metal/metal oxide nanoparticles like silver, gold, titanium dioxide and zinc oxide.

Table-3: Synthesis of metallic nano-particles from various Fungus species

Species	Nanoparticles	Size (in n.m.)	Shapes	Application	References
<i>Verticillium</i>	Silver	21–25	Spherical	Catalysis	[98]
<i>Aspergillus fumigates</i>	Silver	5–25	Spherical	Coating for solar energy absorption and intercalation material for electrical batteries	[99]
<i>Phanerochaete chrysosporium</i>	Silver	50–200	Pyramidal	Medical textiles for antimicrobial activity	[100]
<i>Aspergillus flavus</i> TFR7	Titanium dioxide	12–15	Spherical	Plant nutrient fertilizer	[101]
<i>Fusarium solani</i>	Silver	5–35	Spherical	Biolabeling, sensors, drug delivery	[102]
<i>Penicillium brecompactum</i>	Silver	23–105	Crystalline spherical	Antimicrobial agent	[103]
<i>Penicillium fellutanum</i>	Silver	5–25	Spherical	Thin film and surface coating	[104]
<i>Trichothecium</i> sp.	Gold	10–25	Spherical, rod-like and triangular	–	[105]
<i>Trichoderma viride</i>	Silver	5–40	Spherical	Antimicrobial agent	[106]
<i>Verticillium luteoalbum</i>	Gold	< 10	Triangular, hexagonal	Optics, sensor, coatings	[107]
<i>Fusarium oxysporum</i>	Gold-silver alloy	8–14	Spherical	Biomedical field	[108]
<i>Aspergillus terreus</i>	Zinc oxide	8	Spherical	Catalysis, biosensing, drug delivery, molecular diagnostics, solar cell, optoelectronics, cell labeling, and imaging	[109]

Yeast

Yeasts are one-celled microorganisms contemporary in eukaryotic cells. There are nearly 1500 yeast species have been identified^[110]. The silver and gold nanoparticles were synthesized by a silver-tolerant yeast strain and *Saccharomyces cerevisiae* broth has been conveyed. Various species are working for the research of numerous metallic nanoparticles, as deliberated in Table 4.

Table-4: Synthesis of metallic nano-particles from various Yeast species

Species	Nanoparticles	Size (in n.m.)	Shapes	Application	References
<i>Saccharimycetes cerevisiae</i> broth	Gold, silver	4–15	Spherical	Catalysis	[111]
MKY3	Silver	2–5	Hexagonal	Coatings for solar energy absorption and intercalation material for electrical batteries	[112]

Antibacterial activity

Nano-Particles have progressively been utilized in industry in the course of recent decades with utilizations shifting from food additives^[113] to drug management^[114]. The ceaseless rise of bacterial opposition has provoked the exploration network to create novel antibiotic agents. Among the most auspicious of these novel antibiotic agents are metal nano-particles, which have indicated solid antibacterial activity in a devastating number of studies.

Structure of Bacterial Cell Wall

Most of the microorganisms or bacteria can be isolated into two separate arrangements dependent on their cell divider structure; Gram-positive and Gram-negative. Gram-positive microorganisms contain a thick layer of peptidoglycan in their cell dividers or cell walls, whereas Gram-negative bacteria have a thin peptidoglycan layer with a supplementary outer film comprising of lipopolysaccharide. This supplementary film in Gram-negative bacteria indicates that there is also an extra membrane layer named as periplasm (Fig. 3). Numerous investigations have discovered that Gram-positive bacteria are increasingly impervious to nano-particles systems of activity^[115–119]. In the case of Gram-negative bacteria, such as *Escherichia coli*, bacterial cells are enclosed by a coating of peptidoglycan (~ 8 n.m. thick) and lipopolysaccharide (1–3 μm thick). This procedure may simplify the entrance of unrestricted ions from nano-particles into the cell.

Another likely purpose behind Gram-negative defencelessness to NPs is that Gram-negative bacteria are covered with lipopolysaccharide particles, which convey a negative charge. These negatively charged particles have a higher attraction for the positive ions that a large portion of the NPs discharge, prompting a development and expanded take-up of particles, which at that point cause intracellular harm.

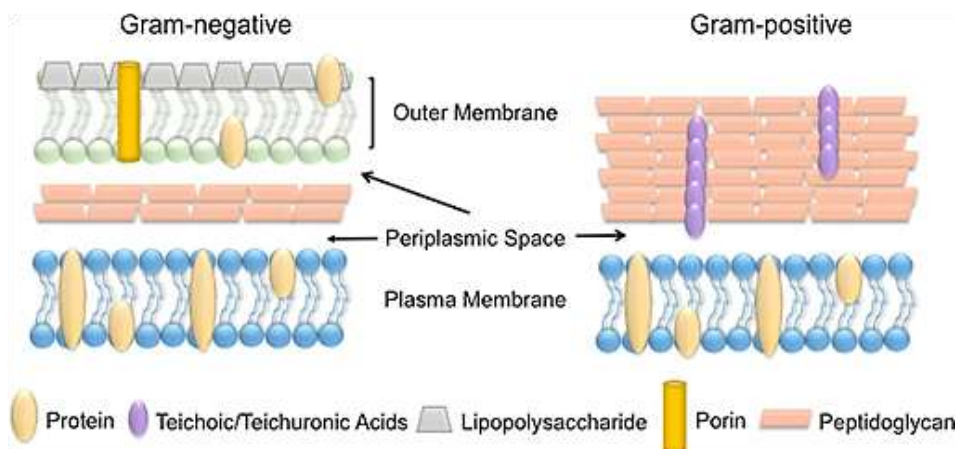


Fig. 3 Comparison of bacterial Gram-positive and Gram-negative cell wall structure^[195]

The Gram-positive and Gram-negative bacteria an adversely charged cell divider, a trademark that is estimated to influence the cooperation between the cell dividers of the microscopic organisms and NPs or particles discharged from them. Studies acted in Gram-microorganisms, for example, *Salmonella typhimurium* indicated that the cell divider is populated with a mosaic of anionic surfaces areas instead of a consistent layer^[120]. Thus, an expected authoritative of a high number of NPs on these negative anionic spaces may increase the central harmfulness on account of the moderately high nano particle focuses in these zones. Also, consolidated investigations of electrophoretic versatility and numerical computations verified that *E. coli* is more adversely charged and inflexible than *S. aureus*^[121].

An exemption to the run of the mill impacts of cell film charge and cell structure is substantial metal safe microscopic organisms. Barely any investigations detailed that these microorganisms are unaffected when presented to metallic NPs, which demonstrated antibacterial action against non-overwhelming metal safe microscopic organisms. For instance, when both Gram-negative *E. coli* and *Cupriavidus metallidurans* strains were presented to TiO₂, Al₂O₃, and carbon nanotube NPs, *E. coli* was delicate and slaughtered by all NPs tried, while *C. metapleurans* was safe notwithstanding being likewise a Gram-negative bacterium, showing that this bacterium is acclimated with being in a domain with substantial metal pressure^[122]. Curiously, transmission electron microscopy investigation demonstrated that the various sorts of TiO₂-NPsutilized in this examination carried on in an alternate way. For instance, TiO₂ Al₂, which was orchestrated utilizing laser pyrolysis^[123].

The Gram-negative bacterium *Shewanella oneidensis* has comparably been demonstrated to have the option to decrease overwhelming metal particles when rewarded with CeO₂ NPs. It was likewise seen as impervious to nano-particle action, though *E. coli* and *Bacillus subtilis* were delicate^[124]. In rundown, all things considered, microbes adjusted to situations tainted with overwhelming metals (metal stresses) are better ready to adapt to NP introduction either by (a) changing the peptidoglycan layer, (b) initiating qualities answerable for cell divider or film fix, or (c) particle sequestration by metabolites or proteins.

In Table-5 the physical and concoction qualities of NPs talked about in this survey are summarized. Naturally, smaller NPs have higher antibacterial action^[122,125-130]. However, few investigations have demonstrated that bigger NPs are progressively powerful, showing that size alone isn't the most significant factor of their poisonousness nature or toxicity^[131,132]. Different components can incorporate the definition procedure, the earth, the bacterial guard instrument and the physical qualities of the nano-particles.

Table-5:Size, Shape, Strain and antibacterial activities of some nano-particles

NP type	Size (nm)	Shape	Strain	Exposure time	Activity	Remarks/purpose	References
Ag	17.5	NR	<i>P. aeruginosa</i> ATCC 27317	4h	G=3.7fold reduction	Citrate capped	[119]
	9–21	NR	Nitrifying bacteria	NR	EC ₅₀ = 0.14 µg/mL	Inhibition of nitrification	[133]
	9.5 26 79	Spherical	<i>S. mutants</i>	24 h	MIC = 4 µg/mL MIC = 8 µg/mL MIC = 4 µg/mL		[129]
	18 80	Spherical	<i>E. coli</i>	8h	MIC = 50 µg/mL MIC = 200 µg/mL		[130]
	13.5	Spherical	<i>E. coli</i> O157:H8, <i>S. aureus</i> ATCC 19636	24h	MIC = > 3.3 nM		[134]
	5–15	Spherical	<i>L. monocytogenes</i> ISP 6508	24h	99.9% killing at 5 wt%	Polyethylene modified	[135]
	9.2	Spherical	<i>E. coli</i> K12 MG 1655	16h	MIC = 2 nM	Oxidized	[136]

		al				particles	
Bio-Ag	2-10	NR	<i>K. pneumonia</i> ATCC 700603 <i>P. mirabilis</i> (collec- tion), <i>S. infantis</i> (collection) <i>P. aeruginosa</i> ATCC 10145 <i>S. aureus</i> ATCC 6338	24h	Z = 2 mm at 100 µg/mL Z = 0 mm at 100 µg/mL Z = 10 mm at 100 µg/mL Z = 8 mm at 100 µg/mL	Synthesized from Actinobacteria CGG 11n supernatant	[137]
Ag/CeO ₂		Rod	<i>E. coli</i> ATCC 8099		G = ~ threefold reduction (100 µg/mL)	Used 1% wt%	[138]
		Cube			G = fourfold reduction (100 µg/mL)		[138]
		Particles			G = ~ 3.5 fold reduction (100 µg/mL)		[138]
		Cube			G = ~ fourfold reduction (100 µg/mL)		[138]
Al ₂ O ₃	11	Spherical	<i>E. coli</i> MG 1655	24h	MIC = 106 µg/mL		[122]
Au	50, 100		<i>E. faecium</i> VRE4 <i>S. oneidensis</i> MR-1		MIC = 32 µg/mL	COOH-, quaternary amine (NMe ₃ +), and methyl-conju- gated (CH ₃ -) NP attachment study	[139]
CeO ₂	6	Square	<i>B. subtilis</i> ATCC 6333 <i>E. coli</i> ATCC 700926	24h	Z = ~ 3.3 mm Z = ~ 0.2 mm		
	22	Ovoid, rectangular, triangular	<i>B. subtilis</i> ATCC 6333 <i>E. coli</i> ATCC 700926	24h	Z = ~ 2.2 mm Z = ~ 1.8 mm		[124]
	40	Heterogeneous	<i>B. subtilis</i> ATCC 6333 <i>E. coli</i> ATCC 700926	24h	Z = ~ 3 mm Z = ~ 1.0 mm		

Cu₂O	40	Heterogeneous	E. coli	18h	MBC = 0.1 mM	Tryptophan-capped	[140]
MgO	4	Square, polyhedral	<i>E. coli</i> C3000, <i>B. megaterium</i> ATCC 14581 <i>B. subtilis</i> ATCC 6333	1h	NG at 250 mg 48% killed	Agar overlay with aero gel	[141]
	20	Amorphous	<i>E. coli</i> XL-1 blue			Metabolic pathway regulation study	[142]
Mg(OH ₂)-MgCl ₂	12.9	Flake	E. coli		88% killed at 100 µg/mL	Co-precipitated with MgCl ₂	[143]
TiO ₂	12	Spherical	<i>E. coli</i> MG 1655	24h	MIC = 100 µg/mL		[122]
	25	Spherical	<i>E. coli</i> MG 1655	24h	MIC = 100 µg/mL		[122]
	< 100	Elongated	<i>E. coli</i> MG 1655	24 h	MIC = 100 µg/mL		[122]
ZnO	12	Spherical	E. coli	24h	Z = 31 mm	Thiol-capped	[125]
	19	Sphere-like		3h	MIC = 50 µg/mL		[144]

The small nano-particles will in general be more harmful than large nano-particles can be clarified by the smaller NPs relative bigger surface region to volume proportion when contrasted with bigger NPs. This can significantly build the creation of ROS, which subsequently can harm and inactivate fundamental biomolecules, including DNA, proteins, and lipids^[145].

Interaction of NPs with intra or extracellular compounds and DNA

It is assumed that, the concentration of nano particle decreases as the NPs interrelate and muddle with organic constituents in the way of life stock and harmed cell parts^[147]. The ZnO-NP toxicity changed significantly relying upon the media wherein they were suspended, recommending that a complexation among Zn²⁺ and explicit particles of the stock happens with a decrease in the antibacterial toxicity^[148]. Different ligands are likewise ready to respond with Ag⁺ and AgNPs, diminishing antibacterial movement because of diminished accessibility as showed by their official to Cl⁻, S²⁻, cysteines and phosphates, which are abundant in aquatic environmental condition^[148]. Moreover, bacteria treated with CuO-NP and Ag-NP demonstrated that bacterial discharge of exopolysaccharides collaborated with the NPs, extracellularly catching the NPs and diminishing toxicity^[149, 148]. NPs sized between 1–12 n.m. appear to have the option to enter into the bacterial intracellular condition^[150, 151]. The NPs discharge ions, which board several sites simultaneously in once inside the cell. Ag-NPs are normally used to explore protein-restricting properties because of their affinity for thiol groups^[152, 153]. According to a proteomic study, it has been demonstrated that around 65% of *E. coli* proteins

bound to Ag-NPs are catalysts^[154]. Among the catalysts or enzymes with a comparative high affinity for Ag-NPs are tryptophanase, liquor dehydrogenase, and cytochrome C, as exhibited in a period subordinate response, recommending a various levelled authoritative to proteins. The non-enzymatic proteins that Ag-NPs tie to are engaged with film honesty, for example, layer porins (OmpA and OmpB), chaperonins, and periplasmic peptide binding proteins^[154]. The high affinity of the periplasmic peptide restricting protein towards Ag-NPs may clarify why these NPs collect in the periplasmic region of the bacteria^[155]. As mentioned earlier, Ag-NPs and all the more explicitly Ag⁺, respond with thiol groups^[152, 150, 153]. Thiol is the functional group on the amino acid cysteine. Cysteine is very vital in biological reactions due to disulphide linking which is decisive for proper protein portable and function, just as, its nucleophilic job in synergist responses. While adding cysteine to a blend of Ag⁺ and microbes, the antibacterial movement of Ag⁺ is killed, showing a cooperation of Ag⁺ with thiol groups^[153,154]. It is imperative to feature that there are thiol bunches in fundamental pathways, for example, respiratory and cell divider combination chemicals, which speak to expected areas of Ag⁺ binding^[155]. The theory that Ag⁺ ties to the DNA was affirmed after the perception that bacterial DNA was dense when both *E. coli* and *S. aureus* species were presented to Ag⁺, prompting a resulting cell augmentation capture^[156]. High-resolution imaging uncovered a low sub-atomic weight area (low density region) shaped in the focal point of the microscopic organisms, recommending this is an instrument of protection utilized by the bacterial cell because of Ag⁺ exposure. This phenomenon recommends that the bacterium faculties either an unsettling influence in the cell layer or the nearness of a danger, for example, Ag⁺ and gathers its DNA to shield it from potential incoming damage^[156]. Surprisingly, when Ag-NPs were utilized instead of Ag⁺ in *E. coli* cells, the build-up didn't happen^[157]. This proposes the bacterial cell may detect the nearness of an edge of Ag⁺ to enact the referenced protection mechanism. Many studies presenting cells to NPs found that the DNA was damaged^[155, 158, 159]. This damage included atomic discontinuity^[160] or physical attachment of the Ag-NPs to the DNA, most likely due to the high affinity of Ag⁺ to phosphates exceptionally bountiful in the DNA molecule^[157].

Effect of NP on DNA replication and repair

Escherichia coli exposed to TiO₂-NPs downregulated genes *dna-X* and *hol-B*, associated with DNA replication^[161]. Downregulation of genes involved in induction of purines (*guaC*), pyrimidines (*pyrC*), and glutaredoxin, an amino acid cofactor (*grxA*), designates the downregulation of DNA synthesis as a retort to TiO₂-NP revelation. This suggests that the cell is under stress and not prioritizing DNA synthesis^[161]. NPs were likewise tried and different DNA fix qualities were animated, including: *recN*, *mutT*, *nfo*, *uvrA*, *uvrD*, *umuD*, *polB*, and *ssb*. This implies that the DNA is harmed upon introduction to metal NPs, yet different systems are activated to react to the harm. Interestingly, the quality *recA* is communicated during DNA harm and presents as an Ag⁺ rewarded phenotype when downregulated^[147]. It is indistinct whether Ag⁺ legitimately downregulates the quality to forestall DNA fix or in the event that it is a consequence of other harmfulness components. For example, *E. coli* cells treated with Ag-NPs didn't endure any worldwide protein change, be that as it may, explicit protein bunches demonstrated an adjustment in guideline. The Ag-NPs have selectivity when official to protein gatherings, yet don't tie enough to change protein-protein communications on a worldwide scale in the cells^[162]. The combination of ROS (Role of reactive Oxygen Species) production, gene regulation changes, cell wall dispersion, and metabolite requisite are contests for variation and existence, and the bacteria fail to create an outline instantaneously against all of the communications (Fig.7).

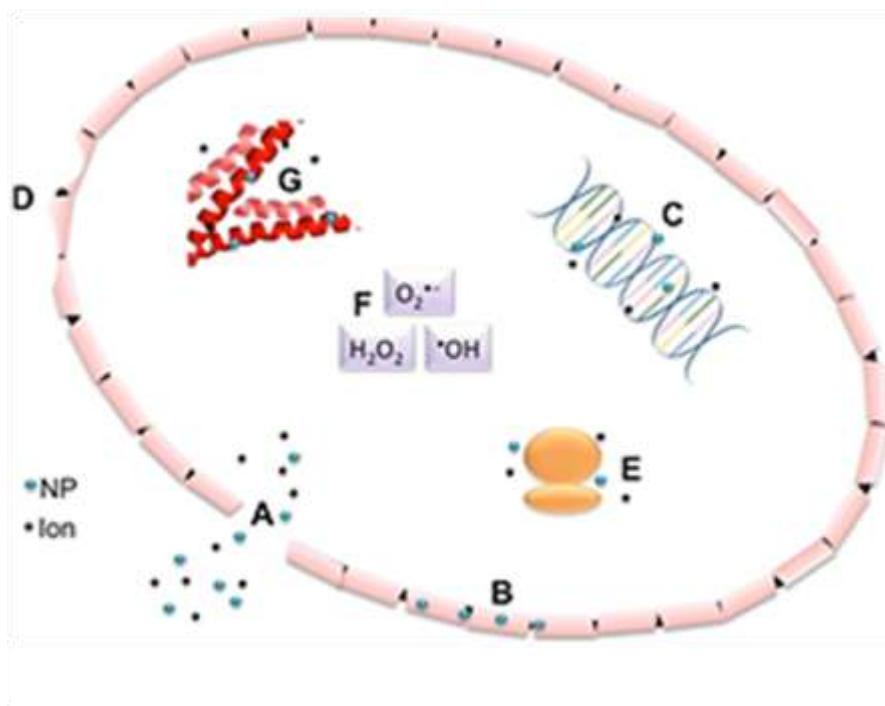


Fig. 7A proposed model indicating the mechanisms of action of Ag-NPs exposed to Gram-negative *E. coli* cell. (A) Breakdown of cell wall permitting intracellular mechanisms to leave the cell. (B) Ag-NPs entering periplasmic space, beginning a separation of the cytosol from membrane. (C) Collaboration of Ag-NPs with DNA. Embarrassment can cause ROS production. (D) Cell pits happening after exposure. (E) Reserve of proper ribosome function, leading to ROS production, malformation or suppression of proteins, unsuitable DNA purpose. (F) ROS production. (G) Communication with proteins, explicitly cysteine^[195].

The multi-target movement brought about by NPs would be perfect to treat and execute multi-medicate safe microscopic organisms, as they likely would not have the option to mount numerous resistances without a moment's delay^[163]. Before future application can be investigated, more examination ought to be done to increase a further comprehension of how the antibacterial framework capacities upon presentation to nanoparticles, with clarification of guessed action and examination concerning new likely potential mechanisms.

Antimicrobial activity

Different investigations have been completed to enhance antimicrobial capacities as a result of the becoming microbial opposition towards basic germicide and antibiotics. Conferring to *in vitro* antimicrobial investigation, the metallic nanoparticles viably impede the few microbial species^[164]. The antimicrobial viability of the metallic nanoparticles relies on two vital parameters: (a) material utilized for the synthesis of the nanoparticles and (b) their molecule size. Over the time, microbial protection from antimicrobial medications has developed progressively raised and is hence a substantial threat to general wellbeing. For instance, antimicrobial drug resistant bacteria contain methicillin-safe, sulfonamide-resistant, penicillin-resistant, and vancomycin-resistant properties^[165].

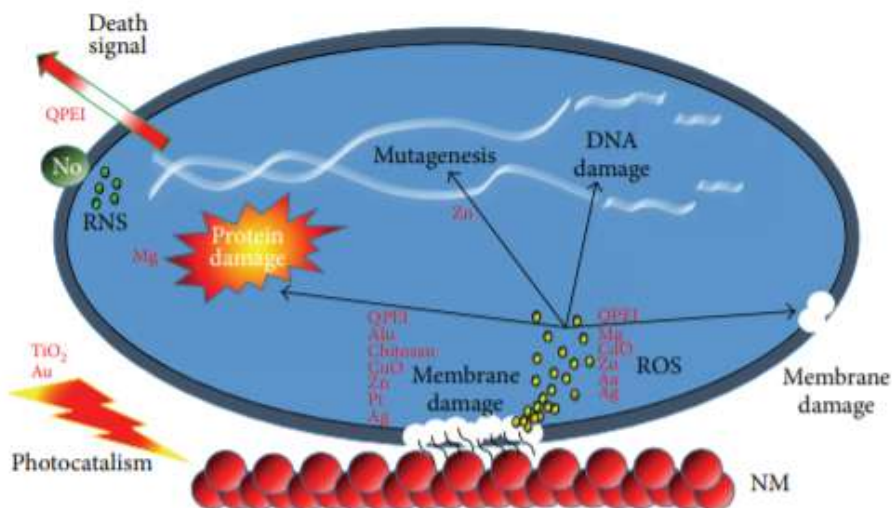


Fig. 4. Scheme displaying antibacterial method of activity of nano-metal [146].

Antibiotics face numerous multilevel difficulties, for example, combatting multidrug-resistant strains and biofilms. The adequacy of antibiotic is probably going to diminish quickly in view of the medication obstruction abilities of microbes. Hence, even when microscopic organisms are treated with huge dosages of anti-toxins, sicknesses will persist in living creatures. Biofilms are additionally a significant method of giving multidrug obstruction against overwhelming amounts of anti-infection living beings. Biofilms are also a vital way of providing multidrug confrontation against heavy doses of antibiotics. Medication obstruction happens for the most part in irresistible ailments, for example, lung contamination and gingivitis [166]. The most encouraging methodology for lessening or keeping away from microbial drug obstruction is the application of nanoparticles. Because of different mechanisms, metallic nanoparticles can exclude or overpower the multidrug-resistance and bio-film arrangement.

Different nanoparticles utilize various components simultaneously to fight organisms [e.g., metal-containing nanoparticles, NO-releasing nanoparticles (NO NPs), and chitosan-containing nanoparticles (chitosan NPs)]. Nanoparticles can fight drug opposition since they work utilizing various mechanisms. Therefore, microorganisms should at the same time have various quality changes in their cell to defeat the nanoparticle.

Table-6: Multiple Mechanism of antimicrobial action for various metallic nanoparticles [167]

S.NO.	Nanoparticles	Multiple mechanisms
1.	Nitric oxide-releasing nanoparticles (NO-NPs)	NO forms reactive nitrogen oxide intermediates (RNOS) by reacting with superoxide (O_2^-) (a) RNOS cause direct nitrosative damage to DNA, including causing strand breaks, formation of abasic sites and depleting the Fe in a bacterial cell (b) RNOS inactivate zinc metalloproteins, which results in inhibition of microbial cellular respiration (c) RNOS also cause lipid peroxidation
2.	Chitosan-containing nanoparticles	(a) Due to its positive charge, chitosan binds with DNA in bacterial and fungal cells, thereby inhibiting transcription of mRNA resulting in protein translation (b) Chitosan also decreases the activities of metalloproteins

3.	Silver-containing nanoparticles (Ag-NPs)	The antimicrobial activity of silver (Ag) is due to its $-Ag^+$ ions (a) Ag^+ inhibits the electron transport chain of microbes (b) Ag^+ damages DNA and RNA by binding with them (c) Ag^+ also inhibits cell division by inhibiting DNA replication (d) Ag^+ ions form ROS, which are toxic to both bacterial cells and eukaryotic host cells
4.	Zinc oxide-containing nanoparticles (ZnO-NPs)	(a) ZnO NPs destroy both lipids and the proteins of the membrane, which can cause cell death (b) ZnO NPs also form Zn^{2+} ions and ROS, including hydrogen peroxide (H_2O_2), which damage the bacterial cell
5.	Copper-containing nanoparticles	(a) Copper interacts with amine and carboxyl groups, which are present on microbes such as <i>B. subtilis</i> (b) Higher concentrations of Cu^{2+} ions can produce ROS
6.	Titanium dioxide-containing nanoparticles (TiO_2 -NPs)	(a) In the photocatalysis process TiO_2 NPs generate ROS, including hydrogen peroxide (H_2O_2) and hydroxyl radicals ($\cdot OH$), upon exposure to near-UV and UVA radiation
7.	Magnesium-containing nanoparticles	(a) MgX_2 NPs also cause lipid peroxidation of the microbial cell envelope by generating (b) MgF_2 NPs can cause lipid peroxidation and a drop in cytoplasmic pH, which raises membrane potential

However, immediate multiple biological gene alterations in the same cell are doubtful^[167]. Numerous mechanisms detected in nanoparticles are deliberated in Table 5. Silver nanoparticles are the most appreciated inorganic nanoparticles, and they are used as efficient antimicrobial, antifungal, antiviral, and anti-inflammatory specialists^[168]. According to a literature survey or overview, the antimicrobial possibility of silver nanoparticles can be defined in the accompanying manners: (1) denaturation of the bacterial external film or outer membrane^[169], (2) generation-gaps in the bacterial cell layer prompting discontinuity of the cell film^[170, 171], and (3) collaborations between Ag NPs and disulphide or sulfhydryl gatherings of chemicals upset metabolic procedures; this progression prompts cell death^[172]. The shape-dependent antimicrobial action was likewise analysed. According to Pal et al.^[173], shortened triangular nanoparticles are profoundly responsive in nature on the grounds that their high-particle thickness surfaces have improved antimicrobial activity.

Catalytic activity

4-Nitrophenol and its subordinates are utilized to make herbicides, bug sprays, and engineered dyestuffs, and they can significantly harm the biological system as normal natural poisons of wastewater. Because of its harmful and inhibitory nature, 4-nitrophenol is an extraordinary ecological concern. Therefore, the decrease of these contaminations is urgent. The 4-nitrophenol reduction product, 4-aminophenol, has been applied in assorted fields as a transitional for paracetamol, sulphur colours, elastic cancer prevention agents, arrangement of dark/white film engineers, consumption inhibitors, and antecedents in antipyretic and pain-relieving drugs^[174, 175]. The simplest and most effective way to diminish 4-nitrophenol is to present $NaBH_4$ as a reductant and a metal compound/catalyst like Au NPs^[176], Ag NPs^[177], CuO NPs^[178], and Pd NPs^[179] respectively.

The UV-visible region of 4-nitrophenol was considered by a sharp SPR band at 400 n.m. as a nitrophenolate ion was created within the sight of NaOH. The expansion of Ag NPs (Which was synthesized from extract of *Chenopodium aristatum L. stem*) to the response medium prompted a quick rot reaction medium led to a fast decay in the absorption intensity at 400 n.m., which was simultaneously joined by the presence of a wide band at 313 n.m., showing the development of 4-aminophenol^[180] (Fig. 5).

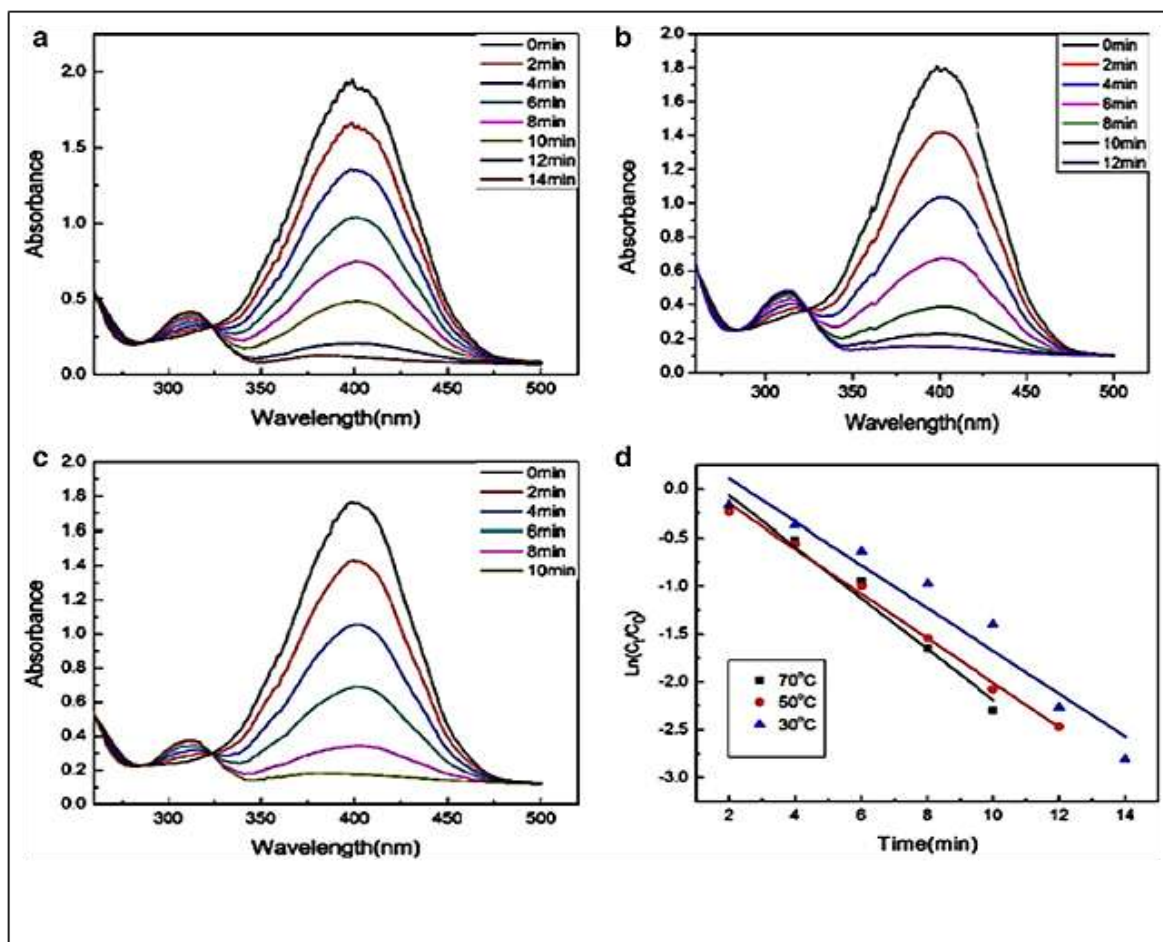


Fig. 5 UV-visible spectra illustrating *Chenopodium aristatum L.* stem extract synthesized Ag NP-mediated catalytic drop of 4-NP to 4-AP at three unlike temperatures (30 °C, 50 °C and 70 °C). Decrease in the absorption intensity of the characteristic nitrophenolate band at 400 n.m. conveyed by associated entrance of a wider absorption band at 313 n.m. designates the formation of 4-AP^[190]

Removal of pollutant dyes

Both cationic dyes and anionic dyes are key class of natural toxins used in different applications^[181]. Organic dyes assume a basic role because of their immense interest in paper factories, textiles, plastic, leather, food, printing, and pharmaceuticals enterprises. In textile businesses, about 60% of dyes are expended in the assembling procedure of pigmentation for some textures^[182]. After the texture procedure, about 15% of dyes are squandered and are released into the hydrosphere, and they speak to a significant wellspring of contamination because of their unmanageability nature^[183]. The pollutants from these assembling units are the most significant sources of biological contamination. Therefore, the management of sewages containing dyes is one of the unnerving challenges in the field of environmental chemistry^[184].

In the nano technology, semiconductor nanomaterials have better photocatalytic movement relative than the massive materials. Metal oxide semiconductor nanoparticles (like WO_3 , ZnO , SnO_2 , TiO_2 , and CuO) have been applied specially for the photocatalytic action of manufactured dyes^[185-188]. The benefits of these nano photocatalysts like ZnO and TiO_2 nanoparticles are ascribable to their high surface area a region to mass proportion to upgrade the adsorption of natural pollutants/poisons. The surface energy of the nanoparticles rises because of the large number of surface sensitive sites accessible on the nanoparticle surfaces. This leads to arise in rate of pollutant elimination at low concentrations. Accordingly, a lower extent of nano-catalyst will be mandatory to treat polluted water comparative to the bulk material^[189-192]. Like metal oxide nanoparticles, metal nanoparticles too display heightened photocatalytic deprivation of several pollutant dyes like synthesized silver nanoparticles from *Z. armatum* leaf extract were used for the corruption of different pollutant dyes^[193] (Fig. 6).

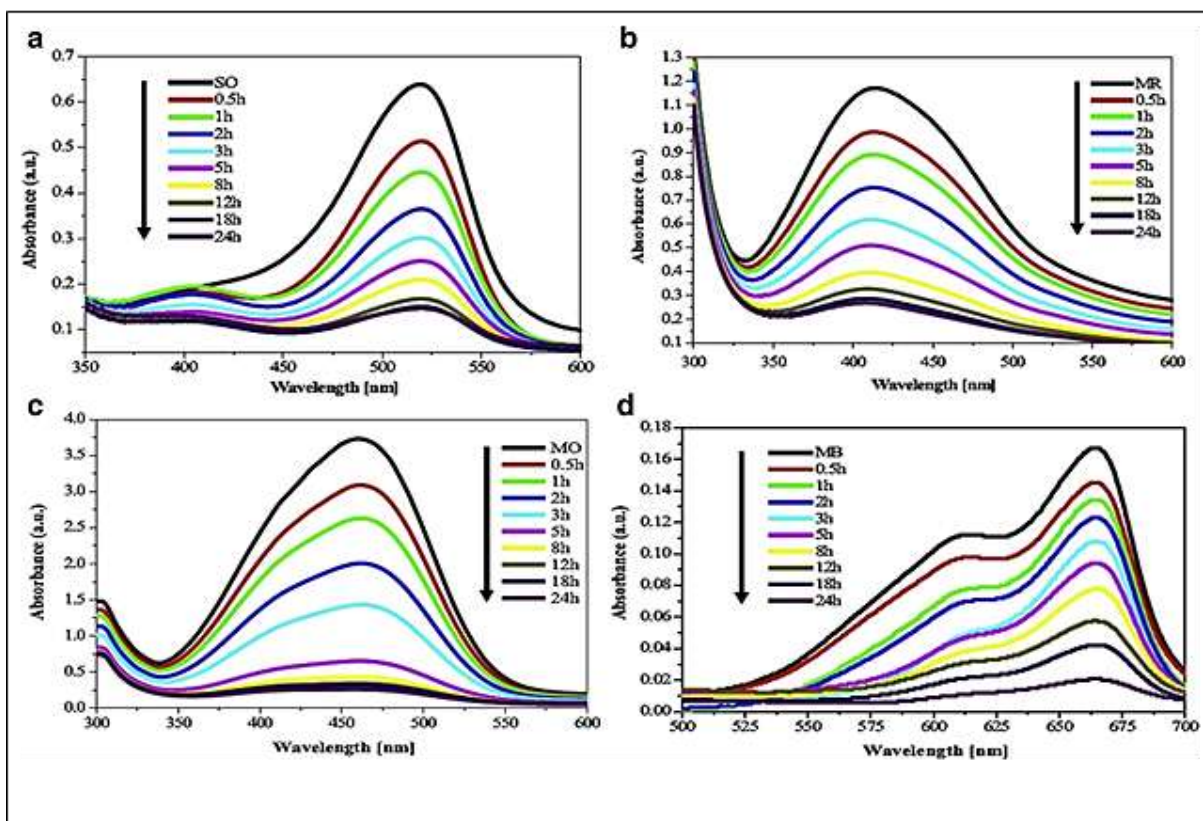


Fig. 6 Graphic representation for the decrease of safranin, methyl red, methyl orange, and methylene blue dyes by means of silver NPs manufactured from *Z. armatum* leaf extract by metallic nanoparticles [194]

Conclusion

The “Green synthesis” of metal and metal oxide nanoparticles, mechanisms and their efficient literature review will help to come across the present difficulties in green synthesis. In summary, future innovative work of forthcoming materials or nanoparticles synthesis should be focused on the way to stretching out research facility-based work to a modern scale by considering customary or present issues, particularly health and environmental impacts. It is also manifest in the literature that both NPs and specific ions shows durable antibacterial activities. In spite of the fact that the different pathways that appear to be at the same time initiated by NPs make clarification a difficult task, they are additionally the motivation behind why nano-particle presentation is so effective. Before future application can be investigated, more examination ought to be done to increase a further comprehension of how the antibacterial framework capacities upon presentation to NPs, with clarification of guessed action and examination concerning new likely potential mechanisms.

Future Possibilities

Biosynthesis of metals and their oxide materials or nanoparticles utilizing marine algae and marine plants is a region or area that remains unknown. Similarly, sufficient prospects remain for the investigation of new green preliminary techniques dependent on biogenic synthesis. Standardized practices in NP manufacture should be considered for maximal approval amongst future studies related to it, which should incorporate a cytotoxicity investigation and afiery reaction. Moreover, the rising number of multiple-drug unaffected bacterial strains should be tended to testing clinical confines as opposed to customary strains from microbial assortments.

References

1. Huang X, El-Sayed IH, Qian W, El-Sayed MA. Cancer cell imaging and photothermal therapy in the near-infrared region by using gold nanorods. *J Am Chem Soc.* 2006;128:2115–20.

2. Kim JS, Kuk E, Yu KN, et al. Antimicrobial effects of silver nanoparticles. *Nanomed Nanotechnol Biol Med.* 2007;3:95–101.
3. Livage J, Henry M, Sanchez C. Sol–gel chemistry of transition metal oxides. *Prog Solid State Chem.* 1988;18:259–341.
4. O’Neal DP, Hirsch LR, Halas NJ, et al. Photo-thermal tumor ablation in mice using near infrared-absorbing nanoparticles. *Cancer Lett.* 2016;209:171–6.
5. Sastry M, Ahmad A, Khan MI, Kumar R. Biosynthesis of metal nanoparticles using fungi and actinomycete. *Curr Sci.* 2003;85:162–70.
6. Su X-Y, Liu P-D, Wu H, Gu N. Enhancement of radiosensitization by metal-based nanoparticles in cancer radiation therapy. *Cancer Biol Med.* 2014;11:86–91.
7. Oskam G. Metal oxide nanoparticles: synthesis, characterization and application. *J Sol–gel Sci Technol.* 2006;37:161–4.
8. Hofmann MR, Martin ST, Choi W, Bahnemann DW. Environmental applications of semiconductor photocatalysis. *Chem Rev.* 1995;95:69–96.
9. Laurent S, Forge D, Port M, et al. Magnetic iron oxide nanoparticles: synthesis, stabilization, vectorization, physicochemical characterizations, and biological applications. *Chem Rev.* 2008;108:2064–110.
10. Pelgrift RY, Friedman AJ. 2013 Nanotechnology as a therapeutic tool to combat microbial resistance. *Adv. Drug Deliv. Rev.* 65, 1803 – 1815.
11. Raza RA, Kanwal Z, Rauf A, Sabri AN, Riaz S, Naseem S. 2016 Size- and shape-dependent antibacterial studies of silver nanoparticles synthesized by wet chemical routes. *Nanomaterials* 6, 74.
12. Kanwal Z, Raza MA, Riaz S, Manzoor S, Tayyeb A, Sajid I, Naseem S. 2019 Synthesis and characterization of silver nanoparticle-decorated cobalt nanocomposites (Co@AgNPs) and their density-dependent antibacterial activity. *R. Soc. open sci.* 6: 182135.
13. Bohara RA, Pawar SH. 2015 Innovative developments in bacterial detection with magnetic nanoparticles. *Appl. Biochem. Biotechnol.* 176, 1044– 1058.
14. Cao G. *Nanstructures and nanomaterials—synthesis, properties and applications.* Singapore: World Scientific; 2004.
15. Yuvakkumar R, Suresh J, Nathanael A J, Sundrarajan M and Hong S I 2014 *Mater. Sci. Eng. C* 41 17.
16. Doble M, Kruthiventi AK. *Green chemistry and engineering.* Cambridge: Academic Press; 2007.
17. Yoosaf K, Ipe BI, Suresh CH, Thomas KG. In situ synthesis of metal nanoparticles and selective naked-eye detection of lead ions from aqueous media. *J Phys Chem C.* 2007;111:12839–47.
18. Sylvestre J, Poulin S, Kabashin AV, et al. Surface chemistry of gold nanoparticles produced by laser ablation in aqueous media. *J Phys Chem B.* 2004;108:16864–9.
19. Er H, Yasuda H, Harada M, et al. Formation of silver nanoparticles from ionic liquids comprising N-alkylethylenediamine: effects of dissolution modes of the silver(I) ions in the ionic liquids. *Colloids Surf A Physicochem Eng Asp.* 2017;522:503–13.
20. Srivastava V. In situ generation of Ru nanoparticles to catalyze CO₂ hydrogenation to formic acid. *Catal Lett.* 2014;144:1745–50.
21. Vollmer C, Redel E, Abu-Shandi K, et al. Microwave irradiation for the facile synthesis of transition-metal nanoparticles (NPs) in ionic liquids (ILs) from metal-carbonyl precursors and Ru-, Rh-, and Ir-NP/IL dispersions as biphasic liquid-liquid hydrogenation nanocatalysts for cyclohexene. *Chem A Eur J.* 2010;16:3849–58.
22. Zhang H, Cui H. Synthesis and characterization of functionalized ionic liquid-stabilized metal (gold and platinum) nanoparticles and metal nanoparticle/carbon nanotube hybrids. *Langmuir.* 2009;25:2604–12.
23. Zhang ZC. Catalysis in ionic liquids. *Adv Catal.* 2006;49:153–237.
24. Dupont J, De Souza RF, Suarez PAZ. Ionic liquid (molten salt) phase organometallic catalysis. *Chem Rev.* 2002;102:3667–92.
25. Van Rantwijk F, Sheldon RA. Biocatalysis in ionic liquids. *Chem Rev.* 2007;107:2757–85.
26. Welton T. Ionic liquids in catalysis. *Coord Chem Rev.* 2004;248:2459–77.
27. Bussamara R, Melo WWM, Scholten JD, et al. Controlled synthesis of Mn₃O₄ nanoparticles in ionic liquids. *Dalton Trans.* 2013;42:14473.
28. Kim K-S, Demberelnyamba D, Lee H. Size-selective synthesis of gold and platinum nanoparticles using novel thiol-functionalized ionic liquids. *Langmuir.* 2004;20:556–60.
29. Sue K, Adschiri T, Arai K. Predictive model for equilibrium constants of aqueous inorganic species at subcritical and supercritical conditions. *Ind Eng Chem Res.* 2002;41:3298–306.

30. Shankar SS, Rai A, Ahmad A, Sastry M. Rapid synthesis of Au, Ag, and bimetallic Au core Ag shell nanoparticles using Neem (*Azadirachta indica*) leaf broth. *J Colloid Interface Sci.* 2004;1:1.
31. Maensiri S, Laokul P, Klinkaewnarong J, et al. Indium oxide (in $2O_3$) nanoparticles using aloe vera plant extract: synthesis and optical properties. *J Optoelectron Adv Mater.* 2008;10:161–5.
32. Krishnaraj C, Jagan EG, Rajasekar S, et al. Synthesis of silver nanoparticles using *Acalypha indica* leaf extracts and its antibacterial activity against water borne pathogens. *Colloids Surf B Biointerfaces.* 2010;1:1.
33. Kasthuri J, Veerapandian S, Rajendiran N. Biological synthesis of silver and gold nanoparticles using apiin as reducing agent. *Colloids Surf B Biointerfaces.* 2009;68:55–60.
34. Armendariz V, Herrera I, Peralta-Videa JR, et al. Size controlled gold nanoparticle formation by *Avena sativa* biomass: use of plants in nanobiotechnology. *J Nanoparticle Res.* 2004;6:377–82.
35. Mondal S, Roy N, Laskar RA, et al. Biogenic synthesis of Ag, Au and bimetallic Au/Ag alloy nanoparticles using aqueous extract of mahogany (*Swietenia mahogany* JACQ.) leaves. *Colloids Surfaces B Biointerfaces.* 2011;82:497–504.
36. Huang Q, Li D, Sun Y, et al. Biosynthesis of silver and gold nanoparticles by novel sundried *Cinnamomum camphora* leaf. *Nanotechnol.* 2007;1:1.
37. Narayanan KB, Sakthivel N. Coriander leaf mediated biosynthesis of gold nanoparticles. *Mater Lett.* 2008;62:4588–90.
38. Shankar SS, Rai A, Ahmad A, Sastry M. Controlling the optical properties of lemongrass extract synthesized gold nanotriangles and potential application in infrared-absorbing optical coatings. *Chem Mater.* 2005;17:566–72.
39. Song JY, Kim BS. Biological synthesis of bimetallic Au/Ag nanoparticles using Persimmon (*Diopyros kaki*) leaf extract. *Korean J Chem Eng.* 2008;25:808–11.
40. Ravindra S, Murali Mohan Y, Narayana Reddy N, Mohana Raju K. Fabrication of antibacterial cotton fibres loaded with silver nanoparticles via “green approach”. *Colloids Surf A Physicochem Eng Asp.* 2010;367:31–40.
41. Dubey M, Bhaduria S, Kushwah BS. Green synthesis of nanosilver particles from extract of *Eucalyptus hybrida* (Safeda) leaf. *Dig J Nanomater Biostruct.* 2009;4:537–43.
42. Veerasamy R, Xin TZ, Gunasagaran S, et al. Biosynthesis of silver nanoparticles using mangosteen leaf extract and evaluation of their antimicrobial activities. *J Saudi Chem Soc.* 2010.
43. Jia L, Zhang Q, Li Q, Song H. The biosynthesis of palladium nanoparticles by antioxidants in *Gardenia jasminoides* Ellis: long lifetime nanocatalysts for p-nitrotoluene hydrogenation. *Nanotechnology.* 2009.
44. Raghunandan D, Bedre MD, Basavaraja S, et al. Rapid biosynthesis of irregular shaped gold nanoparticles from macerated aqueous extracellular dried clove buds (*Syzygium aromaticum*) solution. *Colloids Surf B Biointerfaces.* 2010;79:235–40.
45. Gardea-Torresdey JL, Gomez E, Peralta-Videa JR, et al. Alfalfa sprouts: a natural source for the synthesis of silver nanoparticles. *Langmuir.* 2003.
46. Gardea-Torresdey JL, Parsons JG, Gomez E, et al. Formation and growth of Au nanoparticles inside live alfalfa plants. *Nano Lett.* 2002;2:397–401.
47. Gardea-Torresdey JL, Tiemann KJ, Gamez G, et al. Gold nanoparticles obtained by bio-precipitation from gold(III) solutions. *J Nanoparticle Res.* 1999;1:397–404.
48. Parashar UK, Saxena PS. Bioinspired synthesis of silver nanoparticles. *J Nanomater.* 2009;4:159–66.
49. Herrera-Becerra R, Zorrilla C, Rius JL, Ascencio JA. Electron microscopy characterization of biosynthesized iron oxide nanoparticles. *Appl Phys A Mater Sci Process.* 2008;91:241–6.
50. Singh J, Singh N, Rathi A, et al. Facile approach to synthesize and characterization of silver nanoparticles by using mulberry leaves extract in aqueous medium and its application in antimicrobial activity. *J Nano-structures.* 2017;7:134–40
51. Singh J, Mehta A, Rawat M, Basu S. Green synthesis of silver nanoparticles using sun dried tulsi leaves and its catalytic application for 4-nitrophenol reduction. *J Environ Chem Eng.* 2018;6:1468–74.
52. Ghodake GS, Deshpande NG, Lee YP, Jin ES. Pear fruit extract-assisted room-temperature biosynthesis of gold nanoplates. *Colloids Surf B Biointerfaces.* 2010;75:584–9.
53. Shankar SS, Ahmad A, Pasricha R, Sastry M. Bioreduction of chloroaurate ions by geranium leaves and its endophytic fungus yields gold nanoparticles of different shapes. *J Mater Chem.* 2003;13:1822.

54. Raghunandan D, Basavaraja S, Mahesh B, et al. Biosynthesis of stable polyshaped gold nanoparticles from microwave-exposed aqueous extracellular anti-malignant guava (*Psidium guajava*) leaf extract. *NanoBiotechnology*. 2009;5:34–41.
55. Qu J, Luo C, Hou J. Synthesis of ZnO nanoparticles from Zn-hyper- accumulator (*Sedum alfredii* Hance) plants. *IET Micro Nano Lett*. 2011;6:174–6.
56. Philip D, Unni C. Extracellular biosynthesis of gold and silver nanoparticles using Krishna tulsi (*Ocimum sanctum*) leaf. *Phys E Low Dimens Syst Nanostructures*. 2011;43:1318–22.
57. Sun Q, Cai X, Li J, et al. Green synthesis of silver nanoparticles using tea leaf extract and evaluation of their stability and antibacterial activity. *Colloids Surf A Physicochem Eng Asp*. 2014;444:226–31.
58. Sadeghi B, Gholamhoseinpoor F. A study on the stability and green synthesis of silver nanoparticles using *Ziziphora tenuior* (Zt) extract at room temperature. *Spectrochim Acta Part A Mol Biomol Spectrosc*. 2015;134:310–5.
59. Fukushi K, Sato T. Using a surface complexation model to predict the nature and stability of nanoparticles. *Environ Sci Technol*. 2005;39:1250–6.
60. Sharma VK, Siskova KM, Zboril R, Gardea-Torresdey JL. Organic-coated silver nanoparticles in biological and environmental conditions: fate, stability and toxicity. *Adv Colloid Interface Sci*. 2014;204:15–34.
61. Tejamaya M, Römer I, Merrifield RC, Lead JR. Stability of citrate, PVP, and PEG coated silver nanoparticles in ecotoxicology media. *Environ Sci Technol*. 2012;46:7011–7.
62. Virkutyte J, Varma RS. Green synthesis of metal nanoparticles: biodegradable polymers and enzymes in stabilization and surface functionalization. *Chem Sci*. 2011;2:837.
63. Mittal AK, Chisti Y, Banerjee UC. Synthesis of metallic nanoparticles using plant extracts. *Biotechnol Adv*. 2013;31:346–56.
64. Dwivedi AD, Gopal K. Biosynthesis of silver and gold nanoparticles using *Chenopodium album* leaf extract. *Colloids Surf A Physicochem Eng Asp*. 2010;369:27–33.
65. Mukunthan KS, Balaji S. Cashew apple juice (*Anacardium occidentale* L.) speeds up the synthesis of silver nanoparticles. *Int J Green Nanotechnol*. 2012;4:71–9.
66. Li X, Xu H, Chen ZS, Chen G. Biosynthesis of nanoparticles by microorganisms and their applications. *J Nanomater*. 2011
67. Prathna TC, Mathew L, Chandrasekaran N, et al. Biomimetic synthesis of nanoparticles: science, technology and applicability. *Biomimetics Learn Nat*. 2010.
68. Iravani S. Green synthesis of metal nanoparticles using plants. *Green Chem*. 2011;13:2638.
69. Zayed MF, Eisa WH, Shabaka AA. *Malva parviflora* extract assisted green synthesis of silver nanoparticles. *Spectrochim Acta Part A Mol Biomol Spectrosc*. 2012;98:423–8.
70. Singh AK, Talat M, Singh DP, Srivastava ON. Biosynthesis of gold and silver nanoparticles by natural precursor clove and their functionalization with amine group. *J Nanoparticle Res*. 2010;12:1667–75.
71. Kesharwani J, Yoon KY, Hwang J, Rai M. Phytofabrication of silver nanoparticles by leaf extract of *Datura metel*: hypothetical mechanism involved in synthesis. *J Bionanosci*. 2009;3:39–44.
72. Makarov VV, Love AJ, Sinitsyna OV, Makarova SS, Yaminsky IV, Taliansky ME et al. “Green” nanotechnologies: synthesis of metal nanoparticles using plants. *Acta Naturae*. 2014;6:35–44.
73. Dahoumane SA, Yéprémian C, Djédiat C, et al. Improvement of kinetics, yield, and colloidal stability of biogenic gold nanoparticles using living cells of *Euglena gracilis* microalga. *J Nanoparticle Res*. 2016.
74. El-Rafe HM, El-Rafe MH, Zahran MK. Green synthesis of silver nanoparticles using polysaccharides extracted from marine macro algae. *Carbohydr Polym*. 2013;96:403–10.
75. Husen A, Siddiqi KS. Plants and microbes assisted selenium nanoparticles: characterization and application. *J Nanobiotechnol*. 2014;12:28.
76. Khan M, Al-Marri AH, Khan M, et al. Green approach for the effective reduction of graphene oxide using *Salvadora persica* L. root (Miswak) extract. *Nanoscale Res Lett*. 2015;10:1–9. <https://doi.org/10.1186/s11671-015-0987-z>.
77. Patel V, Berthold D, Puranik P, Gantar M. Screening of cyanobacteria and microalgae for their ability to synthesize silver nanoparticles with antibacterial activity. *Biotechnol Reports*. 2015;5:112–9. <https://doi.org/10.1016/j.btre.2014.12.001>.
78. Siddiqi KS, Husen A. Fabrication of metal nanoparticles from fungi and metal salts: scope and application. *Nanoscale Res Lett*. 2016;11:1–15.

79. Wadhvani SA, Shedbalkar UU, Singh R, Chopade BA. Biogenic selenium nanoparticles: current status and future prospects. *Appl Microbiol Biotechnol*. 2016;100:2555–66.
80. Gericke M, Pinches A. Microbial production of gold nanoparticles. *Gold Bull*. 2006;39:22–8.
81. Irvani S. Bacteria in nanoparticle synthesis: current status and future prospects. *Int Sch Res Not*. 2014;2014:1–18.
82. Thakkar KN, Mhatre SS, Parikh RY. Biological synthesis of metallic nano- particles. *Nanomed Nanotechnol Biol Med*. 2010;6:257–62.
83. Chen Y-L, Tuan H-Y, Tien C-W, et al. Augmented biosynthesis of cad- mium sulfde nanoparticles by genetically engineered *Escherichia coli*. *Biotechnol Prog*. 2009;25:1260–6.
84. Sunkar S, Nachiyar CV. Biogenesis of antibacterial silver nanoparticles using the endophytic bacterium *Bacillus cereus* isolated from *Garcinia xanthochymus*. *Asian Pac J Trop Biomed*. 2012;2:953–9.
85. Shivaji S, Madhu S, Singh S. Extracellular synthesis of antibacterial silver nanoparticles using psychrophilic bacteria. *Process Biochem*. 2011;46:1800–7.
86. Mann S, Frankel RB, Blakemore RP. Structure, morphology and crystal growth of bacterial magnetite. *Nature*. 1984;310:405–7.
87. Holmes JD, Smith PR, Richardson DJ, et al. Energy-dispersive X-ray analysis of the extracellular cadmium sulfde crystallites of *Klebsiella aerogenes*. *Arch Microbiol*. 1995;163:143–7.
88. Korbekandi H, Irvani S, Abbasi S. Optimization of biological synthesis of silver nanoparticles using *Lactobacillus casei* subsp. *casei*. *J Chem Technol Biotechnol*. 2012;87:932–7.
89. Du L, Jiang H, Liu X, Wang E. Biosynthesis of gold nanoparticles assisted by *Escherichia coli* DH5 α and its application on direct electrochemistry of hemoglobin. *Electrochem Commun*. 2007;9:1165–70.
90. Ahmad N, Sharma S, Alam MK, et al. Rapid synthesis of silver nanopar- ticles using dried medicinal plant of basil. *Colloids Surf B Biointerfaces*. 2010;81:81–6.
91. Wen L, Lin Z, Gu P, et al. Extracellular biosynthesis of monodispersed gold nanoparticles by a SAM capping route. *J Nanoparticle Res*. 2009;11:279–88.
92. Deplanche K, Macaskie LE. Biorecovery of gold by *Escherichia coli* and *Desulfovibrio desulfuricans*. *Biotechnol Bioeng*. 2008;99:1055–64.
93. He S, Guo Z, Zhang Y, et al. Biosynthesis of gold nanoparticles using the bacteria *Rhodospseudomonas capsulata*. *Mater Lett*. 2007;61:3984–7.
94. Philipse AP, Maas D. Magnetic colloids from magnetotactic bacteria: chain formation and colloidal stability. *Langmuir*. 2002;18:9977–84.
95. Chen Y-L, Tuan H-Y, Tien C-W, et al. Augmented biosynthesis of cad- mium sulfde nanoparticles by genetically engineered *Escherichia coli*. *Biotechnol Prog*. 2009;25:1260–6.
96. Mohanpuria P, Rana NK, Yadav SK. Biosynthesis of nanoparticles: technological concepts and future applications. *J Nanoparticle Res*. 2008;10:507–17.
97. Narayanan KB, Sakthivel N. Synthesis and characterization of nanogold composite using *Cylindrocladium foridanum* and its heteroge- neous catalysis in the degradation of 4-nitrophenol. *J Hazard Mater*. 2011;189:519–25.
98. Mukherjee P, Ahmad A, Mandal D, et al. Fungus-mediated synthesis of silver nanoparticles and their immobilization in the mycelial matrix: a novel biological approach to nanoparticle synthesis. *Nano Lett*. 2001;1:515–9.
99. Bhainsa KC, D'Souza SF. Extracellular biosynthesis of silver nanoparticles using the fungus *Aspergillus fumigatus*. *Colloids Surf B Biointerfaces*. 2006;47:160–4.
100. Vigneshwaran N, Ashtaputre NM, Varadarajan PV, et al. Biological syn- thesis of silver nanoparticles using the fungus *Aspergillus favus*. *Mater Lett*. 2007;61:1413–8.
101. Raliya R, Biswas P, Tarafdar JC. TiO₂ nanoparticle biosynthesis and its physiological efect on mung bean (*Vigna radiata* L.). *Biotechnol Rep*. 2015;5:22–6.
102. Ingle A, Rai M, Gade A, Bawaskar M. *Fusarium solani*: a novel biological agent for the extracellular synthesis of silver nanoparticles. *J Nanoparti- cle Res*. 2009;11:2079–85.
103. Shaligram NS, Bule M, Bhambure R, et al. Biosynthesis of silver nano- particles using aqueous extract from the compactin producing fungal strain. *Process Biochem*. 2009;44:939–43.
104. Kathiresan K, Manivannan S, Nabeel MA, Dhivya B. Studies on silver nanoparticles synthesized by a marine fungus, *Penicillium fellutanum* isolated from coastal mangrove sediment. *Colloids Surf B Biointerfaces*. 2009;71:133–7.
105. Ahmad A, Senapati S, Khan MI, et al. Extra-/intracellular biosynthesis of gold nanoparticles by an alkalotolerant fungus, *Trichothecium* sp. *J Biomed Nanotechnol*. 2005;1:47–53.

106. Fayaz AM, Balaji K, Girilal M, et al. Biogenic synthesis of silver nanoparticles and their synergistic effect with antibiotics: a study against gram-positive and gram-negative bacteria. *Nanomed Nanotechnol Biol Med*. 2010.
107. Gericke M, Pinches A. Microbial production of gold nanoparticles. *Gold Bull*. 2006;39:22–8.
108. Senapati S, Ahmad A, Khan MI, et al. Extracellular biosynthesis of bimetallic Au–Ag alloy nanoparticles. *Small*. 2005;1:517–20.
109. Raliya R, Tarafdar JC. Biosynthesis and characterization of zinc, magnesium and titanium nanoparticles: an eco-friendly approach. *Int Nano Lett*. 2014;4:93.
110. Yurkov AM, Kemler M, Begerow D. Species accumulation curves and incidence-based species richness estimators to appraise the diversity of cultivable yeasts from beech forest soils. *PLoS ONE*. 2011;1:1.
111. Mourato A, Gadanho M, Lino AR, Tenreiro R. Biosynthesis of crystalline silver and gold nanoparticles by extremophilic yeasts. *Bioinorg Chem Appl*. 2011;1:1.
112. Kowshik M, Vogel W, Urban J, et al. Microbial synthesis of semiconductor PbS nanocrystallites. *Adv Mater*. 2002;14:815–8.
113. Weir A, Westerhof P, Fabricius L, von Goetz N. Titanium dioxide nanoparticles in food and personal care products. *Environ Sci Technol*. 2012;46:2242–50.
114. Singh R, Lillard JW Jr. Nanoparticle-based targeted drug delivery. *Exp Mol Pathol*. 2009;86:215–23.
115. Feng QL, Wu J, Chen GQ, Cui FZ, Kim TN, Kim JO. A mechanistic study of the antibacterial effect of silver ions on *Escherichia coli* and *Staphylococcus aureus*. *J Biomed Mater Res*. 2000;52:662–8.
116. Kim T-H, Kim M, Park H-S, Shin US, Gong M-S, Kim H-W. Size-dependent cellular toxicity of silver nanoparticles. *J Biomed Mater Res A*. 2012;100:1033–43.
117. Mukha IP, Eremenko AM, Smirnova NP, Mikhienkova AI, Korchak GI, Gorchev VF, et al. Antimicrobial activity of stable silver nanoparticles of a certain size. *Appl Biochem Microbiol*. 2013;49:199–206.
118. Cavassin ED, de Figueiredo LFP, Otoch JP, Seckler MM, de Oliveira RA, Franco FF, et al. Comparison of methods to detect the in vitro activity of silver nanoparticles (AgNP) against multidrug resistant bacteria. *J Nano-biotechnol*. 2015;13:64.
119. Dorobantu LS, Fallone C, Noble AJ, Veinot J, Ma G, Goss GG, et al. Toxicity of silver nanoparticles against bacteria, yeast, and algae. *J Nanopart Res*. 2015;17:172.
120. Magnusson K-E, Bayer ME. Anionic sites on the envelope of *Salmonella typhimurium* mapped with cationized ferritin. *Cell Biophys*. 1982;4:163–75.
121. Sonohara R, Muramatsu N, Ohshima H, Kondo T. Difference in surface properties between *Escherichia coli* and *Staphylococcus aureus* as revealed by electrophoretic mobility measurements. *Biophys Chem*. 1995;55:273–7.
122. Simon-Deckers A, Loo S, Mayne-L'hermite M, Herlin-Boime N, Menguy N, Reynaud C, et al. Size, composition and shape-dependent toxicological impact of metal oxide nanoparticles and carbon nanotubes toward bacteria. *Environ Sci Technol*. 2009;43:8423–9.
123. Pignon B, Maskrot H, Guyot Ferreol V, Leconte Y, Coste S, Gervais M, et al. Versatility of laser pyrolysis applied to the synthesis of TiO₂ nanoparticles—application to UV attenuation. *Eur J Inorg Chem*. 2008;2008:883–9.
124. Pelletier DA, Suresh AK, Holton GA, McKeown CK, Wang W, Gu B, et al. Effects of engineered cerium oxide nanoparticles on bacterial growth and viability. *Appl Environ Microbiol*. 2010;76:7981–9.
125. Padmavathy N, Vijayaraghavan R. Enhanced bioactivity of ZnO nanoparticles—an antimicrobial study. *Sci Technol Adv Mater*. 2008;9:35004.
126. Martinez-Gutierrez F, Olive PL, Banuelos A, Orrantia E, Nino N, Sanchez EM, et al. Synthesis, characterization, and evaluation of antimicrobial and cytotoxic effect of silver and titanium nanoparticles. *Nanomedicine*. 2010;6:681–8.
127. Choi O, Hu Z. Size dependent and reactive oxygen species related nanosilver toxicity to nitrifying bacteria. *Environ Sci Technol*. 2008;42:4583–8.
128. Ivask A, El Badawy A, Kaweeteerawat C, Boren D, Fischer H, Ji Z, et al. Toxicity mechanisms in *Escherichia coli* vary for silver nanoparticles and differ from ionic silver. *ACS Nano*. 2014;8:374–86.
129. Pérez-Díaz MA, Boegli L, James G, Velasquillo C, Sánchez-Sánchez R, Martínez-Martínez R-E, et al. Silver nanoparticles with antimicrobial activities against *Streptococcus* mutants and their cytotoxic effect. *Mater Sci Eng C*. 2015;55:360–6.

130. Cui L, Chen P, Chen S, Yuan Z, Yu C, Ren B, et al. In situ study of the antibacterial activity and mechanism of action of silver nanoparticles by surface-enhanced Raman spectroscopy. *Anal Chem.* 2013;85:5436–43.
131. El Badawy AM, Silva RG, Morris B, Scheckel KG, Suidan MT, Tolaymat TM. Surface charge-dependent toxicity of silver nanoparticles. *Environ Sci Technol.* 2011;45:283–7.
132. Sohm B, Immel F, Bauda P, Pagnout C. Insight into the primary mode of action of TiO₂ nanoparticles on *Escherichia coli* in the dark. *Proteomics.* 2015;15:98–113.
133. Choi O, Hu Z. Size dependent and reactive oxygen species related nanosilver toxicity to nitrifying bacteria. *Environ Sci Technol.* 2008;42:4583–8.
134. Kim JS, Kuk E, Yu KN, Kim J-H, Park SJ, Lee HJ, et al. Antimicrobial effects of silver nanoparticles. *Nanomedicine.* 2007;3:95–101.
135. Tamayo LA, Zapata PA, Vejar ND, Azócar MI, Gulppi MA, Zhou X, et al. Release of silver and copper nanoparticles from polyethylene nanocomposites and their penetration into *Listeria monocytogenes*. *Mater Sci Eng C.* 2014;40:24–31.
136. Lok C-N, Ho C-M, Chen R, He Q-Y, Yu W-Y, Sun H, et al. Silver nanoparticles: partial oxidation and antibacterial activities. *J Biol Inorg Chem.* 2007;12:527–34.
137. Railean-Plugaru V, Pomastowski P, Rafnska K, Wypij M, Kupczyk W, Dahm H, et al. Antimicrobial properties of biosynthesized silver nanoparticles studied by flow cytometry and related techniques. *Electrophoresis.* 2016;37:752–61.
138. Wang L, He H, Yu Y, Sun L, Liu S, Zhang C, et al. Morphology-dependent bactericidal activities of Ag/CeO₂ catalysts against *Escherichia coli*. *J Inorg Biochem.* 2014;135:45–53.
139. Jahnke JP, Cornejo JA, Sumner JJ, Schuler AJ, Atanassov P, Ista LK. Conjugated gold nanoparticles as a tool for probing the bacterial cell envelope: the case of *Shewanella oneidensis* MR-1. *Biointerphases.* 2016;11:11003.
140. Meghana S, Kabra P, Chakraborty S, Padmavathy N. Understanding the pathway of antibacterial activity of copper oxide nanoparticles. *RSC Adv.* 2015;5:12293–9.
141. Stoimenov PK, Klinger RL, Marchin GL, Klabunde KJ. Metal oxide nanoparticles as bactericidal agents. *Langmuir.* 2002;18:6679–86.
142. Leung YH, Ng AMC, Xu X, Shen Z, Gethings LA, Wong MT, et al. Mechanisms of antibacterial activity of MgO: non-ROS mediated toxicity of MgO nanoparticles towards *Escherichia coli*. *Small.* 2014;10:1171–83.
143. Pan X, Wang Y, Chen Z, Pan D, Cheng Y, Liu Z, et al. Investigation of antibacterial activity and related mechanism of a series of nano-Mg(OH)₂. *ACS Appl Mater Interfaces.* 2013;5:1137–42.
144. Li M, Zhu L, Lin D. Toxicity of ZnO nanoparticles to *Escherichia coli*: mechanism and the influence of medium components. *Environ Sci Technol.* 2011;45:1977–83.
145. Karakoti AS, Hench LL, Seal S. The potential toxicity of nanomaterials: the role of surfaces. *JOM.* 2006;58:77–82.
146. Nurit B., Yael Hour-H., Avi Domb, Khan W., and Ronen H. Alternative Antimicrobial Approach: Nano-Antimicrobial Material; Evidence-Based Complementary and Alternative Medicine Volume 2015, 246012, 16.
147. Ivask A, El Badawy A, Kaweeteerawat C, Boren D, Fischer H, Ji Z, et al. Toxicity mechanisms in *Escherichia coli* vary for silver nanoparticles and differ from ionic silver. *ACS Nano.* 2014;8:374–86.
148. Li M, Zhu L, Lin D. Toxicity of ZnO nanoparticles to *Escherichia coli*: mechanism and the influence of medium components. *Environ Sci Technol.* 2011;45:1977–83.
149. Kim JS, Kuk E, Yu KN, Kim J-H, Park SJ, Lee HJ, et al. Antimicrobial effects of silver nanoparticles. *Nanomedicine.* 2007;3:95–101.
150. Wang L, He H, Yu Y, Sun L, Liu S, Zhang C, et al. Morphology-dependent bactericidal activities of Ag/CeO₂ catalysts against *Escherichia coli*. *J Inorg Biochem.* 2014;135:45–53.
151. Pal S, Tak YK, Song JM. Does the antibacterial activity of silver nanoparticles depend on the shape of the nanoparticle? A study of the Gram-negative bacterium *Escherichia coli*. *Appl Environ Microbiol.* 2007;73:1712–20.
152. Mukha IP, Eremenko AM, Smirnova NP, Mikhienkova AI, Korchak GI, Gorchev VF, et al. Antimicrobial activity of stable silver nanoparticles of a certain size. *Appl Biochem Microbiol.* 2013;49:199–206.
153. Liu N, Chen X-G, Park H-J, Liu C-G, Liu C-S, Meng X-H, et al. Effect of MW and concentration of chitosan on antibacterial activity of *Escherichia coli*. *Carbohydr Polym.* 2006;64:60–5.

154. Wigginton NS, de Titta A, Piccapietra F, Dobias J, Nesatyy VJ, Suter MJF, et al. Binding of silver nanoparticles to bacterial proteins depends on surface modifications and inhibits enzymatic activity. *Environ Sci Technol*. 2010;44:2163–8.
155. Choi O, Hu Z. Size dependent and reactive oxygen species related nanosilver toxicity to nitrifying bacteria. *Environ Sci Technol*. 2008;42:4583–8.
156. Wang L, He H, Yu Y, Sun L, Liu S, Zhang C, et al. Morphology-dependent bactericidal activities of Ag/CeO₂ catalysts against *Escherichia coli*. *J Inorg Biochem*. 2014;135:45–53.
157. Liu N, Chen X-G, Park H-J, Liu C-G, Liu C-S, Meng X-H, et al. Effect of MW and concentration of chitosan on antibacterial activity of *Escherichia coli*. *Carbohydr Polym*. 2006;64:60–5.
158. Feng QL, Wu J, Chen GQ, Cui FZ, Kim TN, Kim JO. A mechanistic study of the antibacterial effect of silver ions on *Escherichia coli* and *Staphylococcus aureus*. *J Biomed Mater Res*. 2000;52:662–8.
159. Soni D, Bafana A, Gandhi D, Sivanesan S, Pandey RA. Stress response of *Pseudomonas* species to silver nanoparticles at the molecular level. *Environ Toxicol Chem*. 2014;33:2126–32.
160. Morones JR, Elechiguerra JL, Camacho A, Holt K, Kouri JB, Ramírez JT, et al. The bactericidal effect of silver nanoparticles. *Nanotechnology*. 2005;16:2346.
161. Jahnke JP, Cornejo JA, Sumner JJ, Schuler AJ, Atanassov P, Ista LK. Conjugated gold nanoparticles as a tool for probing the bacterial cell envelope: the case of *Shewanella oneidensis* MR-1. *Biointerphases*. 2016;11:11003.
162. Holt KB, Bard AJ. Interaction of silver(I) ions with the respiratory chain of *Escherichia coli*: an electrochemical and scanning electrochemical microscopy study of the antimicrobial mechanism of micromolar Ag⁺. *Biochemistry*. 2005;44:13214–23.
163. Kumar A, Pandey AK, Singh SS, Shanker R, Dhawan A. Engineered ZnO and TiO₂ nanoparticles induce oxidative stress and DNA damage leading to reduced viability of *Escherichia coli*. *Free Radic Biol Med*. 2011;51:1872–81.
164. Sohm B, Immel F, Bauda P, Pagnout C. Insight into the primary mode of action of TiO₂ nanoparticles on *Escherichia coli* in the dark. *Proteomics*. 2015;15:98–113.
165. Lok C-N, Ho C-M, Chen R, He Q-Y, Yu W-Y, Sun H, et al. Proteomic analysis of the mode of antibacterial action of silver nanoparticles. *J Proteome Res*. 2006;5:916–24.
166. Dizaj SM, Lotfpour F, Barzegar-Jalali M, et al. Antimicrobial activity of the metals and metal oxide nanoparticles. *Mater Sci Eng C*. 2014;44:278–84.
167. Fair RJ, Tor Y. Antibiotics and bacterial resistance in the 21st century. *Perspect Med Chem*. 2014.
168. Jayaraman R. Antibiotic resistance: an overview of mechanisms and a paradigm shift. *Curr Sci*. 2009;96:1475–84.
169. Pelgrift RY, Friedman AJ. Nanotechnology as a therapeutic tool to combat microbial resistance. *Adv Drug Deliv Rev*. 2013;65:1803–15.
170. Zinjarde S. Bio-inspired nanomaterials and their applications as antimicrobial agents. *Chron Young Sci*. 2012;3:74. <https://doi.org/10.4103/2229-5186.94314>.
171. Lok C, Ho C, Chen R, et al. Proteomic analysis of the mode of antibacterial action of silver nanoparticles. *J Proteome Res*. 2006;5:916–24.
172. Egger S, Lehmann RP, Height MJ, et al. Antimicrobial properties of a novel silver-silica nanocomposite material. *Appl Environ Microbiol*. 2009;75:2973–6.
173. Tak YK, Pal S, Naoghare PK, et al. Shape-dependent skin penetration of silver nanoparticles: does it really matter. *Sci Rep*. 2015.
174. Panigrahi S, Basu S, Praharaj S, et al. Synthesis and size-selective catalysis by supported gold nanoparticles: study on heterogeneous and homogeneous catalytic process. *J Phys Chem C*. 2007;111:4596–605.
175. Woo Y, Lai DY. Aromatic amino and nitro-amino compounds and their halogenated derivatives. In: Bingham E, Cohn B, Powell CH, editors. *Patty's toxicology*. Wiley; 2012.
176. Lim SH, Ahn E-Y, Park Y. Green synthesis and catalytic activity of gold nanoparticles synthesized by *Artemisia capillaris* water extract. *Nanoscale Res Lett*. 2016;11:474.
177. Rostami-Vartooni A, Nasrollahzadeh M, Alizadeh M. Green synthesis of perlite supported silver nanoparticles using *Hamamelis virginiana* leaf extract and investigation of its catalytic activity for the reduction of 4-nitrophenol and Congo red. *J Alloys Compd*. 2016;680:309–14.
178. Sharma JK, Akhtar MS, Ameen S, et al. Green synthesis of CuO nanoparticles with leaf extract of *Calotropis gigantea* and its dye-sensitized solar cells applications. *J Alloys Compd*. 2015;632:321–5.

179. Gopalakrishnan R, Loganathan B, Dinesh S, Raghu K. Strategic green synthesis, characterization and catalytic application to 4-nitrophenol reduction of palladium nanoparticles. *J Clust Sci.* 2017;28:2123–31.
180. Yuan CG, Huo C, Gui B, et al. Green synthesis of silver nanoparticles using *Chenopodium aristatum* L. stem extract and their catalytic/anti- bacterial activities. *J Clust Sci.* 2017;28:1319–33.
181. Habibi MH, Rezvani Z. Photocatalytic degradation of an azo textile dye (C.I. Reactive Red 195 (3BF)) in aqueous solution over copper cobaltitene nanocomposite coated on glass by Doctor Blade method. *Spectro- chim Acta Part A Mol Biomol Spectrosc.* 2015;147:173–7.
182. Carmen Z, Daniel S. Textile organic dyes—characteristics, polluting effects and separation/elimination procedures from industrial effluents—a critical overview. *Organic pollutants ten years after the Stockholm convention—environmental and analytical update.* London: InTech; 2012.
183. Ratna PBS. Pollution due to synthetic dyes toxicity and carcinogenicity studies and remediation. *Int J Environ Sci.* 2012;3:940–55.
184. Wesenberg D, Kyriakides I, Agathos SN. White-rot fungi and their enzymes for the treatment of industrial dye effluents. *Biotechnol Adv.* 2003;22:161–87.
185. Devi HS, Singh TD. Synthesis of copper oxide nanoparticles by a novel method and its application in the degradation of methyl orange. *Adv Electron Electr Eng.* 2014;4:83–8.
186. Bhuyan T, Mishra K, Khanuja M, et al. Biosynthesis of zinc oxide nanoparticles from *Azadirachta indica* for antibacterial and photocatalytic applications. *Mater Sci Semicond Process.* 2015;32:55–61.
187. Stan M, Popa A, Toloman D, et al. Enhanced photocatalytic degradation properties of zinc oxide nanoparticles synthesized by using plant extracts. *Mater Sci Semicond Process.* 2015;39:23–9.
188. Thandapani K, Kathiravan M, Namasivayam E, et al. Enhanced larvicidal, antibacterial, and photocatalytic efficacy of TiO₂ nanohybrids green synthesized using the aqueous leaf extract of *Parthenium hysterophorus*. *Environ Sci Pollut Res.* 2017;25:1–12.
189. Astruc D. *Nanoparticles and catalysis.* Weinheim: Wiley; 2008.
190. Dror I, Baram D, Berkowitz B. Use of nanosized catalysts for transformation of chloro-organic pollutants. *Environ Sci Technol.* 2005;39:1283–90.
191. Pradeep T, Anshup. Noble metal nanoparticles for water purification: a critical review. *Thin Solid Films.* 2009;517:6441–78.
192. Tsuda A, Konduru NV. The role of natural processes and surface energy of inhaled engineered nanoparticles on aggregation and corona formation. *NanoImpact.* 2016;2:38–44.
193. Jyoti K, Singh A. Green synthesis of nanostructured silver particles and their catalytic application in dye degradation. *J Genet Eng Biotechnol.* 2016;14:311–7.
194. Karthiga D, Anthony SP. Selective colorimetric sensing of toxic metal cations by green synthesized silver nanoparticles over a wide pH range. *RSC Adv.* 2013;3:16765–74.
195. Slavin Y. N., Asnis J., Häfeli Urs O. and Bach H., Metal nanoparticles: understanding the mechanisms behind antibacterial activity Slavin et al. *J Nanobiotechnol* (2017) 15:65.
