

Polysaccharides Based Colon Specific Drug delivery: A Review

Ravi Kumar^{*1}, M. B. Patil², Sachin R. Patil¹, Mahesh S. Paschapur³

¹ Department of Pharmaceutics, K.L.E.S's College of Pharmacy, Ankola-581314, India,

² Department of Pharmacognosy, K.L.E.S's College of Pharmacy, Ankola-581314, India,

³ Department of Pharmacology, K.L.E.S's College of Pharmacy, Ankola-581314, India,

E-mail: ravikumar300@gmail.com

ABSTRACT: Although oral delivery has become a widely accepted route of administration of therapeutic drugs, the gastrointestinal tract presents several formidable barriers to drug delivery. The delivery of drugs to the colon has a number of therapeutic implications in the field of drug delivery. In the recent times, the colon specific delivery systems are also gaining importance not only for local drug delivery of drugs but also for the systemic delivery of protein and peptide drugs. The various approaches that can be exploited to target the release of drug to colon include prodrug formation, coating with pH sensitive polymers, coating with biodegradable polymers, embedding in biodegradable matrices, hydrogel, timed release systems, osmotic and bioadhesive systems. In this review article we have made an attempt to give an overview on polysaccharide-based colon specific drug delivery system.

Key Word: Polysaccharide, Gastrointestinal tract, Hydrogel.

INTRODUCTION:

The oral route is considered to be most convenient for administration of drugs to patients. The conventional oral dosage forms normally dissolve in the stomach fluid or intestinal fluid and are absorbed from these regions of the Gastrointestinal Tract (GIT), which depend upon the physicochemical properties of the drug. Localized delivery of the drugs in the colon region is possible only when the drug is protected from the hostile environment of upper GIT. Dosage forms that deliver drugs into the colon region rather than upper GIT proffers number of advantages. Oral delivery of drugs to the colon is valuable in the treatment of diseases of colon (ulcerative colitis, Chron's disease, carcinomas and infections) whereby high local concentration can be achieved while minimizing side effects that occur because of release of drugs in the upper GIT or unnecessary systemic absorption. The colon is rich in lymphoid tissue. Uptake of antigens into the mast cells of the colonic mucosa produces rapid local production of antibodies and this helps in efficient vaccine delivery¹. Specific systemic absorption of drugs and protein/peptides in the colonic region offers interesting possibilities for the treatment of disease susceptible to diurnal rhythm such as asthma, arthritis or inflammation²⁻³. The colon is considered to be more suitable for delivery of peptides and protein in comparison to small intestine^{4,5}. Besides this low hostile environment, the colonic transit time (20-30 hours) and the colonic tissue is highly responsive to the action of

absorption enhancers^{6,7}. Colonic delivery can be accomplished by oral or rectal administration. Rectal dosage forms such as suppositories and enemas are not always effective since a high variability in the distribution of these forms is observed⁸⁻¹⁰. The GIT is divided into various regions like stomach, small intestine and large intestine. The colon serves four major functions: viz; creation of suitable environment for the growth of colonic microorganisms, storage reservoir of faecal contents, expulsion of the contents of the colon at an appropriate time, absorption of potassium and water from lumen and excretion of potassium and bicarbonate. An overview of the pH details of the GIT¹¹ is shown in Table 1. Gastric emptying of dosage forms is highly variable and depends primarily on whether the subject is fed or fasted and on the properties of the dosage form such as size and density. The transit times of small dosage forms in GI tract is shown in Table 2.

TABLE1: AVERAGE pH IN THE GIT

Location	pH
Oral cavity	6.2-7.4
Oesophagus	5.0-6.0
Stomach	Fasted condition: 1.5-2.0 Fed condition: 3.0-5.0
Small intestine	Jejunum: 5.0-6.5 Ileum: 6.0-7.5
Large intestine	Right colon: 6.4 Mild colon and left colon: 6.0-7.6

Organ	Transit time(h)
Stomach	<1 (Fasting) >3 (Fed)
Small intestine	3-4
Large intestine	20-30

TABLE3: CRITERIA FOR SELECTION OF DRUGS FOR COLON SPECIFIC DRUG DELIVERY SYSTEMS:

Criteria	Nonpeptide drugs	Peptide drugs
Drugs used for local effects in colon against GIT diseases	Diclofenac, Metaprolol	Amylin, Calcitonin
Drugs poorly absorbed from upper GIT	Ibuprofen, Theophylline, Isosorbides	Cyclosporine, Desmopressin
Drugs for colon cancer	Pseudoephedrine	Glucagon, Epoetin
Drugs that degrade in stomach and small intestine	Bromophenaramine 5-Flourouracil	Gonadorelin, Insulin
Drugs that under go extensive first pass metabolism	Nimustine, Bleomycin	Sermorelin, Saloatonin
Drugs for targeting	5-Aminosalicylic- acid, Prednisolone	Vasopressin, urotilitin

Drug candidate for colonic drug delivery:

Drugs which show poor absorption from the stomach or intestine including peptide drugs, are most suitable for colon specific drug delivery systems. The criteria for selection of drugs for colon specific drug delivery systems^{12,13} is shown in Table 3. Selection of carrier for particular drug candidate depends on the physicochemical nature of the drug as well as the disease for which the system is to be used. The factors such as chemical nature, stability and partition coefficient of the drug and the type of the absorption enhancer chosen

influence the carrier selection. Choice of drug carrier depends on the functional groups of the drug molecule¹⁴. There are several ways in which colon specific drug delivery has been attempted¹⁵. This includes prodrug formation, coating with pH sensitive polymers, coating with biodegradable polymers, embedding in biodegradable matrices and hydrogel, timed-release systems, osmotic systems, and bioadhesive systems. In this review article we have made an attempt to focus on the polysaccharide based colon delivery systems.

POLYSACCHARIDES BASED APPROACHES:**TABLE 4: CHARACTERISTICS OF VARIOUS BIODEGRADABLE POLYSACCHARIDES FOR COLON TARGETED DRUG DELIVERY.**

Polymer	Chemical name	General properties	Bacterial species that degrade polysaccharide
Amylose	-1, 4 D- glucose	Unbranched constituent of starch, used as tablet excipients	Bactericides
Arabinogalactose	-1,4 and -1,3 galactose, -1,6 and -1,3 D- arabinose and D- galactose	Natural pectin, hemi cellulose Used as a thickening agent	Bifidobacterium
Chitosan	Deacetylated -1, 4- N- acetyl -D- glucosamine	Deacetylated chitin, used as a absorption enhancing agent	Bactericides
Cyclodextrins	-1, 4 D- glucose	Cyclic structures of 6,7or 8 units used as a solubilising and absorption enhancing agent	Bactericides
Chondroitin sulphate	-1,3 D- glucuronic acid and N- acetyl -D- glucosamine	Mucopolysaccharides, contains various amounts esters of sulphate at 4 or 6 position	Bactericides
Pectin	-1,4 D- galacturonic acid and 1,2 D- rhamnose with D- galactose and D- arabinose side chain	Partial methyl ester, commonly used as thickening agent	Bifidobacterium, Eubacterium,
Dextran	-1, 6 D- glucose -1,3 D- glucose	Plasma expanders	Bactericides
Guar gum	-1, 6 D-galactose -1,4 D- mannose	Galactomanan, used as a thickening agent	Bacteroides, Ruminococcus
Xylan	-1,4 D- xylose with -1,3 L- arabinose side chains	Abundant hemi cellulose of plant cell wall	Bacteroides, Bifidobacterium

Biodegradable matrix and hydrogel systems:

The inability of GIT enzymes to digest certain plant polysaccharides (pectin, xylan) is taken as an advantage to develop colon specific drug delivery systems¹⁶. The drug is embedded in the matrix core of the biodegradable polymer by compressing the blend of active drug, a degradable polymer and additives. Various polysaccharides such as pectin, guar gum, inulin, amylase, cyclodextrins etc. have been investigated for their use in colon targeted drug delivery systems. The bacterial enzymes of colon degrade the carrier polymer and release the contents for localized or systemic absorption through colon¹⁷⁻²¹. The most important fact in the development of polysaccharide derivatives for colon targeted drug delivery is the selection of a suitable biodegradable polysaccharide. As these polysaccharides are usually soluble in water, they must be made water insoluble by cross-linking or hydrophobic derivatisation.

Very important is an optimal proportion of the hydrophobic and hydrophilic parts and the number of free hydroxy groups in the polymeric molecule. The general properties of polysaccharides used in colon targeted drug delivery are shown in Table 4.

Pectin: Pectin an anionic polysaccharide extracted from plant primary cell wall was used by Ashford *et al*,²². Depending on the plant source and preparation they contain varying degree of methyl ester substituents²³. Excessive solubility of pectin in water creates problem in fabrication of colon targeted delivery systems. Pectin alone is unable to protect the load of drug as GI fluids penetrates into and releases the drug by diffusion. This problem can be manipulated through choice of suitable pectin type or the presence of additives^{24,25}. Coating of pectin remains unaffected in presence of

gastric and small intestinal enzymes but is completely digested in presence of colonic bacterial enzymes. Pectin in the form of compression coat was evaluated for drug targeting to colon²⁶. Compression coated core tablets of 5- Amino Salicylic acid (5-ASA) were prepared using pectin and HPMC²⁷. Munjeri *et.al.*²⁸, investigated amidated pectin for colonic drug delivery using indomethacin and sulfamethoxazole as model drugs. Walkerly *etal.*²⁹⁻³¹ used biodegradable coating containing pectin and ethyl cellulose for colon specific drug delivery. Ashford *etal.*³² evaluated high and low methoxy pectin for colonic drug delivery. Rubinstein *et.al.*³³ developed colonic specific drug delivery system using calcium pectinate by using calcium pectinate using indomethacin as model drug. Atyabi *etal.*³⁴ developed and evaluated Bovine serum albumin-loaded pectinate beads for colonic peptide delivery system. Ahmed *etal.*³⁵ studied the effect of simulated gastrointestinal conditions on drug release from pectin/ethyl cellulose as film coating for drug delivery to the colon. Bourgeois *etal.*³⁶ evaluated the pectin beads for colonic delivery of lactamases. Hiorth *etal.*³⁷ studied the immersion coating of pellets with calcium pectinate and Chitosan. Kosaraju *etal.*³⁸ Sande *etal.*³⁹ developed pectin-based oral drug delivery to the colon. Zhang *etal.*⁴⁰ developed calcium pectinate capsules for colonic drug delivery. Prabhashankar *etal.*⁴¹ studied formulation and roentgenographic studies of naproxen-pectin-based matrix tablets for colon drug delivery. Zhambito *etal.*⁴² studied matrices for site-specific controlled-delivery of 5-fluorouracil to descending colon. Fishman *etal.*⁴³ developed pectin-based systems for colon-specific drug delivery via oral route. Sinha *etal.*⁴⁴ formulated and evaluated colonic drug delivery of 5-fluorouracil. Ahrabi *etal.*⁴⁵ developed pectin matrix tablets for colonic delivery of model drug ropivacaine. Sriamornsak *etal.*⁴⁶ developed composite film-coated tablets intended for colon-specific delivery of 5-aminosalicylic acid by using deesterified pectin. El-Gibaly *etal.*⁴⁷ developed oral delayed-release system based on Zn-pectinate gel (ZPG) microparticles as an alternative carrier to calcium pectinate beads for colonic drug delivery. Turkoglu *etal.*⁴⁸ developed the *in vitro* evaluation of pectin-HPMC compression coated 5-aminosalicylic acid tablets for colonic delivery. Mura P⁴⁹ developed enteric-coated pectin-based matrix tablets for colonic delivery of theophylline. The chemical structure of pectin is shown in Figure 1.

Inulin:

Inulin is a naturally occurring polysaccharide found in many plants. It is not hydrolyzed by the endogenous secretions of the human digestive tract⁵⁰. It is metabolized in colon⁵¹⁻⁵². Inulin HP (high degree polymerization) was incorporated in eudragit RS film was evaluated as a possible biodegradable coating for colonic drug delivery. Vervoort *etal.*⁵³ Maris *etal.*⁵⁴ have done preliminary studies on synthesis and

characterization of various inulin hydro gels as carriers for colonic drug delivery system. Stubbe *etal.*⁵⁵ developed azo containing polysaccharide gels more specifically azo- inulin and azo dextran gels. The chemical structure of inulin is shown in Figure 2.

Guar gum:

Guar gum is a natural polysaccharide derived from the seeds of *Cyamopsis tetragonolobus*, having molecular weight of approximately 1,000,000, giving it a high viscosity in solution⁵⁶. Due to its high molecular weight it is metabolized in large intestine due to the presence of microbial enzymes⁵⁷⁻⁵⁸. Guar gum is hydrophilic in nature and swells in cold water forming viscous colloidal dispersions or sols⁵⁹. This gelling property retards release of the drug from the dosage form as well as it is susceptible to degradation in the colonic environment. To reduce the swelling properties of the guar gum it was reacted with glutaraldehyde under acidic conditions to obtain different products with increasing cross-linking densities. The products were characterized by measuring their swelling properties in simulated and intestinal fluids and their cross linking densities⁶⁰. Reduction in the enormous swelling by cross linking resulted in biodegradable hydrogel formation, which was able to retain poorly water-soluble drug. Krishnaiah *etal.*⁶¹ studied the influence of metronidazole and tinidazole on the usefulness of guar gum, a colon-specific drug carrier. Compression coated tablets⁶² of 5-ASA and matrix tablets of mebendazole have been prepared using guar gum as a carrier. Matrix tablets of guar gum with dexamethasone, indomethacin and budenoside have been investigated for colon targeted drug delivery^{63,64}. Matrix tablets containing various proportions of guar gum were prepared by wet granulation technique using starch paste as a binder^{65,66}. Das *etal.*⁶⁷ have studied cross-linked guar gum hydrogel discs for colon-specific delivery of ibuprofen: formulation and *in vitro* evaluation. Al-saidan *etal.*⁶⁸ studied *in vitro* and *in vivo* evaluation of guar gum-based matrix tablets of rofecoxib for colonic drug delivery. Krishnaiah *etal.*⁶⁹⁻⁷⁴ studied pharmacokinetics evaluation of various antiprotozoal drugs in healthy volunteers. Momin *etal.*⁷⁵ studied *in vitro* studies on guar gum based formulation for the colon-targeted delivery of Sennosides. Chourasia *etal.*⁷⁶ developed guar gum microspheres for target specific drug release to colon. Krishnaiah *etal.*⁷⁷ have tried guar gum as a carrier for colon specific delivery and they also studied the influence of metronidazole and tinidazole on *in vitro* release of albendazole from guar gum matrix tablets. Krishnaiah *etal.*⁷⁸ Studied of guar gum compression-coated 5-aminosalicylic acid tablets for colon-specific drug delivery. The chemical structure of guar gum is shown in Figure(3).

Amylose: Amylose is a polysaccharide obtained from plant extracts and is a component of starch. These are safe, nontoxic, and easily available. Colon-specific drug

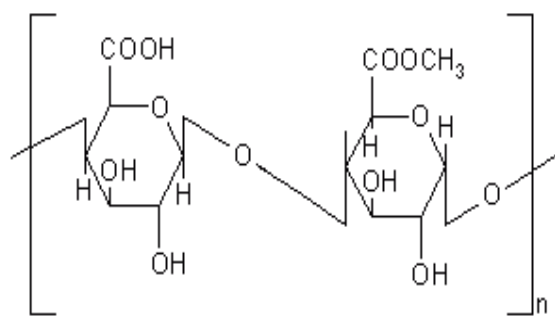


Figure1: Chemical structure of pectin.

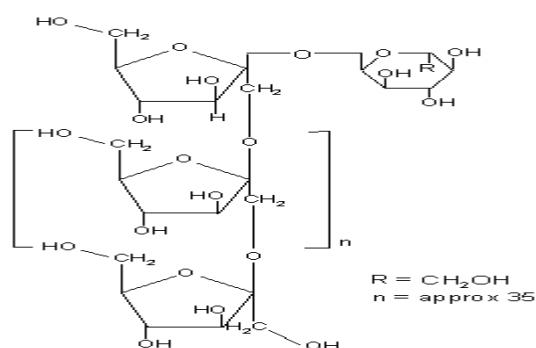


Figure 2: Chemical structure of inulin.

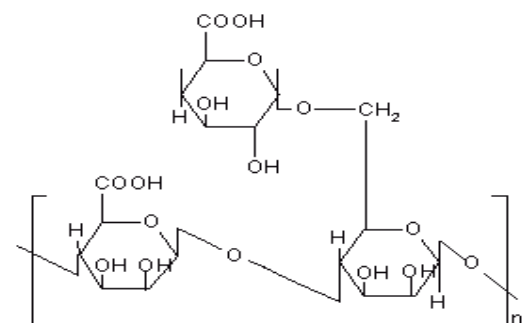


Figure 3: Chemical structure of guar gum

delivery may be possible by the application of dried amylose films to pharmaceutical formulations. Amylose, one of the major fractions of starch, possesses the ability to form films through gelation, when prepared under appropriate conditions. The microstructure of the film is potentially resistant to the action of pancreatic α -amylase but is digested by amylases of the colonic microflora. However, under simulated gastrointestinal conditions, coatings made solely of amylose will become porous and allow drug release. Incorporation of insoluble polymers into the amylose film, to control amylose swelling, provides a solution to this problem. A range of cellulose

and acrylate based copolymers were assessed, of which a commercially available ethyl cellulose (Ethocel) was found to control the swelling most effectively. Various works have been reported on formulation and evaluation of *in vitro* potential of amylose- ethocel coating system for colon-targeted delivery⁷⁹⁻⁸³. A mixture of amylose and ethocel (1:4) has been developed for colonic drug delivery using [¹³C] glucose as model drug⁸⁴. Epichlorhydrin treated cross-linked amylose was introduced as a matrix for controlled release of theophylline⁸⁵. The chemical structure of guar gum is shown in Figure 4.

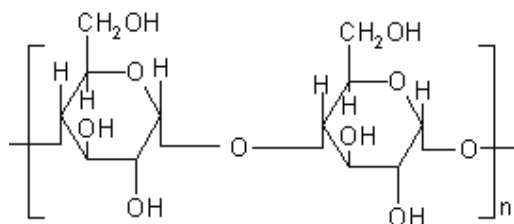


Figure 4: Chemical structure of Amylose

Chondroitin Sulfate:

Is a soluble mucopolysaccharide utilized as a substrate by the bacteriodes mainly by *Bacteriodes thetaiotaomicron* and *B.obvatus*^{86,87}. Natural chondroitin sulfate is water-soluble. However, cross-linked chondroitin sulfate is less hydrophilic and thus would provide a better shield. Colon Specific drug delivery systems based on chondroitin sulfate and cross-linked chondroitin sulfate were reported

by Rubinstein *et al*,^{88,89} have developed colonic drug delivery systems based on chondroitin sulfate and cross-linked chondroitin sulfate. Sintov *et al*⁹⁰ have developed indomethacin colon specific drug delivery using crosslinked chondroitin sulfate and studied for water uptake and release characteristics. The chemical Structure of chondroitin sulfate is shown in Figure 5.

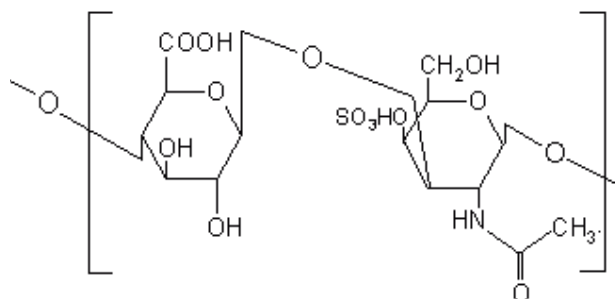


Figure 5: Chemical structure of Chondroitin Sulfate

Cyclodextrin:

Are cyclic oligosaccharides consisted of six to eight glucose units joined through a α -1,4 glucosidic bonds. They remain intact in stomach and small intestine, in the colon they under go fermentation due to the presence of colonic microflora and absorbed from these regions^{91,92}. Tanaka *et al*,⁹³ have prepared several cyclodextrin complexes for colon specific drug delivery systems. Hiramaya *et al*,⁹⁴ prepared two cyclodextrin conjugates i.e. ester and amide conjugates and it was shown that ester conjugate released the drug preferentially in colon than in stomach /small intestine. Yano *et al*,⁹⁵ have prepared the colon specific drug delivery system for prednisolone using α cyclodextrin. Various literatures available on formulation of prodrug of cyclodextrins with drug molecules, which provide a versatile means for construction of not only colon-targeted delivery systems but also delayed release systems⁹⁶⁻⁹⁷. The chemical structure of α -cyclodextrin is shown in Figure 6.

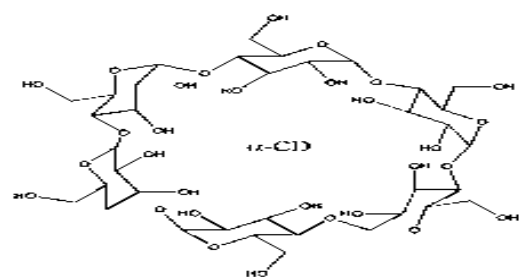


Fig 6: Chemical Structure of α -Cyclodextrin

Chitosan:

Chitosan is a high molecular weight polycationic polysaccharide derived from naturally occurring chitin by deacetylation. Chemically it is poly (N- glucosamine) and shows resistance to enzymes of upper GI tract⁹⁸. It is nontoxic, biocompatible and biodegradable. It is widely used as food ingredient⁹⁹. Tozazki *et al*,^{100,101} developed colon specific insulin delivery system with chitosan capsules. Tozaki *et al*,¹⁰² used rats to study the colon specificity of chitosan capsules R-68070, a thromboxane synthetase inhibitor used for chemically induced ulcerative colitis. Shimono *et al*,¹⁰³ developed new colon specific drug delivery system containing chitosan

dispersed drug delivery system composed of active ingredient, reservoir and drug release regulating layer dispersing chitosan powder in hydrophobic polymer. Orienti *et al*,¹⁰⁴ synthesized various salts of chitosan and evaluated for colon specific delivery system. Vandelli *et al*,¹⁰⁵, shu *et al*,¹⁰⁶ have developed a pH sensitive based chitosan hydro gels drug delivery system. Suzuki *et al*,¹⁰⁷ prepared hard capsules of chitosan with enteric polymers for colon targeted drug delivery. Jain *et al*,¹⁰⁸ developed albendazole microspheres for colon specific delivery using Chitosan HCl . The chemical structure of Chitosan is shown in Figure 7.

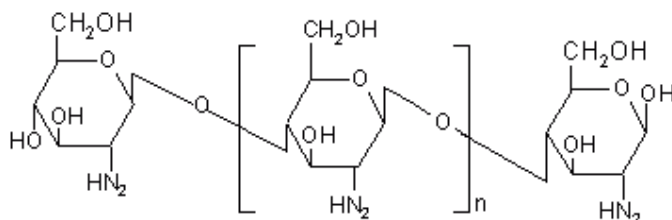


Fig 7:Chemical Structure of Chitosan

Dextran:

Dextran are colloidal, hydrophilic and water soluble substances, obtained from microorganisms of the family of Lactobacillus. Dextran of various molecular weights have been used as drug carriers. Various dextran ester prodrugs have been prepared and evaluated for their efficacy to deliver the drug to their target organ i.e. colon. Harboe, *et al*^{109,110} synthesized dextran ester prodrug. They formulated dextran T-70- naproxen ester and compared the bioavailability of prodrug form naproxen alone. They found that 100% bioavailability from prodrug form than the drug alone, they also found that the release of prodrug form of drug was released more in caecum homogenate than from the homogenate of small intestine of pig.¹¹¹⁻¹¹² Dextran ester prodrugs of metronidazole have been prepared and characterized for colon specific drug release¹¹³⁻¹¹⁵. Lee *et al*,¹¹⁶ developed a dextran-nalidixic acid ester with a varied degree of substitution for colon specific delivery. Jung *et al*,¹¹⁷ were

synthesized dextran ester prodrugs of 5-ASA and drug release rate study revealed that drug release was accelerated in large intestine . Bauer & Kesselhut¹¹⁸ synthesized dextran fatty acid ester and showed that lauroyl dextran esters with molecular weight of approximately 250000 and degree of substitution ranging from 0.11 to 0.3 were suitable for colon targeted drug delivery as film coatings. *In vitro* studies with lauroyl dextran esters bearing theophylline were carried out and it was shown that addition of dextranase accelerated the drug release¹¹⁹⁻¹²⁰. The side effects of steroid therapy, which are used in the treatment of chronic colitis, may be decrease by selectively delivering the drug to the colon using dextran¹²¹⁻¹²². Various dextran ester prodrugs viz:sulasalazine,budenoside,mesalazine,olsalazine etc., were formulated for colon delivery of steroids for local and systemic action¹²³⁻¹³⁰. The chemical structure of dextran is shown in Figure 8.

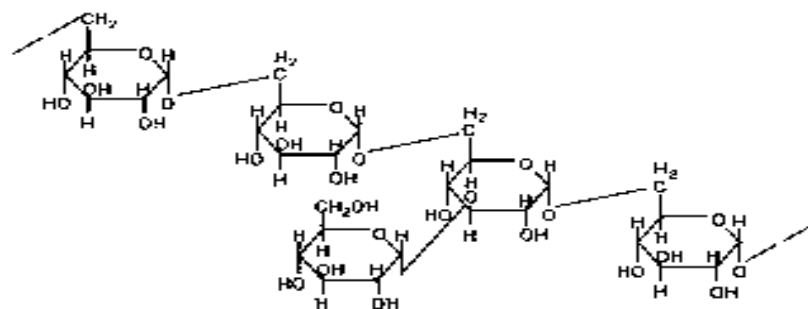


Fig 8: Chemical Structure of Dextran

Alginates:

Alginates are natural hydrophilic polysaccharide derived from seaweed and consist of 1-4, linked D- mannuronic acid and L- glucuronic acid residues. Alginates are easily gelled in presence of a divalent cation as calcium ion. The gelation /cross-linking is due to the stacking of the glucuronic acid blocks of alginate chains. Shun *etal.*^{131,132} developed calcium alginate beads as cores with a spray coat of 5-ASA on them. This system was prepared by coating calcium alginate beads with Aqua coat® that is a pH independent polymer followed by 2% w/v coating of eudragit L-30D. Being enteric polymer the release of drug in acidic medium was resisted and release was triggered in alkaline pH. Kiyoungh *etal.*¹³³ prepared alginate beads and coated with dextran acetate. In the absence of dextranase the release was minimal. The chemical structure of alginate is shown in Figure 9.

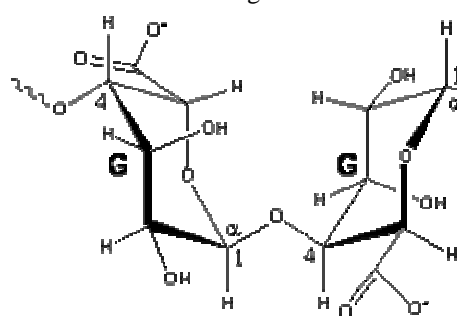


Fig 9:Chemical Structure of Alginate

Locust Bean Gum:

It is derived from carob (*Ceratonia siliqua*) seeds. It has as irregularly shaped molecule with -1,4- D-galactomanan units. This is neutral polymer slightly soluble in cold water. Raghav *etal.*¹³⁴ formulated and evaluated colon specific drug delivery systems based on polysaccharides; they used locust bean gum and chitosan in the ratio of 2:3, 3:2 and 4:1 were evaluated using *invitro* and *in vivo* methods. From *invitro* and *in vivo* studies revealed that locust bean and chitosan was capable of protecting the drug from being release in the stomach and small intestine and was susceptible to colonic bacterial enzymatic actions with resultant drug release in the colon.

CONCLUSION

Colon targeted drug delivery systems are exploited to selectively target the drug release to the colon. Several approaches have been investigated to achieve site specificity to colon. The polysaccharides based colon specific drug delivery is relatively easy due to the presence of various derivatizable groups, wide range of molecular weights, varying chemical compositions, low toxicity and high stability. The selection of suitable polysaccharide is a critical parameter in the fabrication of

colon specific drug delivery. The main limitation of this approach is their excessive water solubility. This high hydrophilicity cause to loose the strong network of polysaccharides and consequently drug is slowly released in the upper part of GIT. This can be over come by using cross-linking agents like glutaraldehyde, epichlorhydrin. This particular approach has brought in a break through in delivery system design and development.

REFERENCES:

1. Sarasija S, Hota A. Colon specific drug delivery systems. *Indian J Pharm Sci* 2000;62:1-6.
2. Saffran M, Bedra C, Kumar GS, Neckers DC. Vasopressin: a model for the study of effects of additives on the oral and rectal administration of peptide drugs. *J Pharm Sci* 1988;77(1):33-8.
3. Mackay M and Tomlinson E. "Colonic delivery of therapeutic peptides and proteins" In: *Colonic drug absorption and metabolism*, Bieck, P. Eds., Marcel Dekker, New York, 1993;159-176.
4. Lee V HL. "Changing needs in drug delivery" In: *Peptide and protein drug delivery*, Lee VHL., Eds., Marcel Dekker, New York, 1.
5. Ikesue K, Kopeckova P and Kopecek J. Degradation of Proteins by Enzymes of the Gastrointestinal Tract. *Proc Int Symp Control Rel Bioact. mater* 1991;18:580-581.
6. Digenis GA, Sandefer E. Gamma scintigraphy and neutron activation techniques in the *in vivo* assessment of orally administered dosage forms. *Crit Rev Ther Drug Carrier Syst.* 1991;7(4):309-45.
7. Taniguchi K, Muranishi S, Sezaki H. Enhanced intestinal permeability to macromolecules. II. Improvement of the large intestinal absorption of heparin by lipid-surfactant mixed micelles in rats. *Int J Pharm* 1980; 4:219-228.
8. Hardy JG, "Colonic transit and drug delivery." In: *drug delivery to the gastrointestinal Tract*, Hardy JG, Davis S S, Wilson, C.G, Eds., Ellis Horwood, Chichester, 1989:75.
9. Jay M, Beihn RM, Digenis GA, Deland FH, Caldwell L, Mlodozeniec AR. Disposition of radiolabelled suppositories in humans. *J Pharm Pharmacol.* 1985;37(4):266-8.
10. Wood E, Wilson CG, Hardy JG. The spreading of foam and solution enemas. *Int J Pharm* 1985; 25:191-197.
11. Wilson GC and Washington N. "Physiological pharmaceuticals, Biological Barriers to drug absorption". Ellis Horwood Ltd., Chichester, UK, 1989:22.
12. Antonin KH, Saano V, Bieck P, Hastewell J, Fox R, Lowe P, MacKay M. Colonic absorption of human calcitonin in man. *Clin Sci* 1992;83:627-631

13. Fara JW, In: Novel drug delivery and its therapeutic application, Presscot L.F., and Nimmo WS, Eds., Wiley Chichester, 989,103.
14. Friend DR. Colon-specific drug delivery. *Adv Drug Deliv Rev* 1991;7:149-199.
15. Van den Mooter, G, Kinget R. Oral colon specific drug delivery: A review. *Drug Deli* 1995; 2: 81-93.
16. Salyers A A, Vercellotti J R, West S H E, Wilkins TD. Fermentation of mucin and plant polysaccharides by strains of *Bacteroides* from the human colon. *Appl Environ Microbiol* 33:319-322.
17. Englyst HN, Hay S, Macfarlane GT. Polysaccharide breakdown by mixed populations of human faecal bacteria. *Microbiol* 1987; 95:163.
18. Ratner BD, Gladhill KW, Hobert T A. Analysis of *invitro* enzymatic and oxidative degradation of polyurethanes. *J Biomed Mater Res* 1988;22:509.
19. Hergenrother RW, Wabers HD, Cooper SL. The effect of chain extenders and stabilizers on the *invivo* stability of polyurethanes. *J Appl Biomater* 1992;3:17.
20. Park K, Shalaby S W W and Park H, In: Guillet J, Eds., *Biodegradable hydrogels for drug delivery*. Technomic Publishing Company, USA, 1993, 13.
21. Hovgaard L and Brondsted H. Current applications of polysaccharides in colon targeting. *Crit Rev Ther Drug Carr Syst* 1996;13:185.
22. Ashford M, Fell J, Attwood D, Sharma H, Woodhead P. An evaluation of pectin as a carrier for drug targeting to the colon. *J Control Rel* 1993;26: 213.
23. Towle GA and Christensen O, In: Whistler R L, Be Miller J N, Eds., *Industrial gums and their derivatives*. Academic Press, NY, 1973, 429.
24. Rubinstein A. Microbially controlled drug delivery to the colon. *Biopharm Drug Dispos* 1990;11: 465.
25. Rubinstein A, Radai R. Pectic salt as a colonic delivery system. *Proc Int Symp Control Rel Bioact Mater* 1991; 18:221.
26. Wakerly Z, Fell JT, Attwood D, Parkins DA. *Invitro* evaluation of pectin-based colonic drug delivery systems. *Int J Pharm* 1996;129:73.
27. Turkoglu M, Ugurlu T. *Invitro* evaluation of pectin-HPMC compression coated 5-aminosalicylic acid tablets for colonic delivery. *Eur J Pharm Biopharm* 2002; 53:65.
28. Munjeri O, Collett J H, Fell JT. Hydrogel beads based on amidated pectins for colon-specific drug delivery . *J Control Rel* 1997;46:272.
29. Wakerly Z, Fell J T, Attwood D, Parkins DA. Pectin/ethylcellulose film coating formulations for colonic drug delivery. *Pharma Res* 1996; 13: 1210.
30. Wakerly Z, Fell JT, Attwood D, Parkins D A. Studies on drug release from pectin/ethylcellulose film-coated tablets: a potential colonic delivery *Int J Pharm* 1997;153: 219.
31. Wakerly Z, Fell J T, Attwood D and Parkins D A. Pectin/ethylcellulose film coating formulations. for colonic drug delivery. *J Pharm Pharmacol* 1997; 49:1210.
32. Ashford M, Fell J, Attwood D, Sharma H and Woodhead P. Studies on pectin formulations for colonic drug-delivery. *J Control Rel* 1994; 30: 225.
33. Rubeinstein A, Radai R, Ezra M, Patahk S and Rokem JM. A Potential Colon Specific Drug Delivery Carrier. *Pharma Res* 1993;10: 258.
34. Atyabi F, Inanloo K and Dinarvand R. Bovine serum albumin loaded pectinate beads as colonic peptide delivery system: Preparation and *in vitro* characterization. *Drug Deliv* 2005;12: 367.
35. Ahmed I S. Effect of simulated gastrointestinal conditions on drug release from pectin/ethylcellulose as film coating for drug delivery to the colon. *Drug Dev Ind Pharm* 2005; 31:465.
36. Bourgeois S, Laham A, Besnard M, Andremonet A and Fattal E. *In vitro* and *in vivo* evaluation of pectin beads for the colon delivery of beta-lactamases. *J Drug Target* 2005;13: 277.
37. Hiorth M, Versland T, Heikkila J, Tho I and Sande SA. Immersion coating of pellets with calcium pectinate and chitosan. *Int J Pharm* 2006;308:25.
38. Kosaraju SL. Colon targeted delivery systems: review of polysaccharides for encapsulation and delivery. *Crit Rev Food Sci Nutr* 2005; 45: 251.
39. Sande SA. Pectin-based oral drug delivery to the colon. *Expert Opin Drug Deliv* 2005; 3: 441.
40. Xu C, Zhang JS, Mo Y, Tan RX. Calcium pectinate capsules for colon-specific drug delivery. *Drug Dev Ind Pharm* 2005;31:127-34.
41. Rao KP, Prabhashankar B, Kumar A, Khan A, Biradar SS, Srishail SP and Satyanath B F. Formulation and roentgenographic studies of naproxen-pectin-based matrix tablets for colon drug delivery. *Yale J Biol Med* 2003; 76:149.
42. Zambito Y, Baggiani A, Carelli V, Serafini MF and Di Colo G. Matrices for site-specific controlled-delivery of 5-fluorouracil to descending colon. *J Control Rel* 2005;102: 669.
43. Liu L, Fishman ML, Kost J and Hicks KB. Pectin-based systems for colon-specific drug delivery via oral route. *Biomaterials* 2003;24:3333.
44. Sinha VR, Mittal BR, Bhutani KK and Kumria R. Colonic drug delivery of 5-fluorouracil: an *in vitro* evaluation. *Int J Pharm* 2004;269:101.
45. Ahrabi SF, Madsen G, Dyrstad K, Sande SA, and Graffner C. Development of pectin matrix tablets for colonic delivery of model drug ropivacaine. *Eur J PharmSci* 2000 ;10:43.
46. Sriamornsak P, Nunthanid J, Wanchana S and Luangtana-Anan M. Composite film-coated tablets intended for colon-specific delivery of 5-aminosalicylic acid: using deesterified pectin. *Pharm Dev Technol* 2003;8:311.

47. El-Gibaly I. Oral delayed-release system based on Zn-pectinate gel (ZPG) microparticles as an alternative carrier to calcium pectinate beads for colonic drug delivery. *Int J Pharm* 2002; 232:199.
48. Turkoglu M and Ugurlu T. In vitro evaluation of pectin-HPMC compression coated 5-aminosalicylic acid tablets for colonic delivery. *Eur J Pharm Biopharm* 2002; 53:65-73.
49. Mura P, Maestrelli F, Cirri M, Gonzalez Rodriguez ML and Rabasco Alvarez AM. Development of enteric-coated pectin-based matrix tablets for colonic delivery of theophylline. *J Drug Target* 2003; 11:365.
50. Dysseler B S and Hoffen M J. Inulin, an alternative dietary fibre. Properties and quantitative analysis. *Eur J Clin Nutr* 1995; 49:145.
51. Wang X and Gibson G R. Effects of the in vitro fermentation of oligofructose and inulin by bacteria growing in the human large intestine. *J appl Bacteriol* 1993; 25:373.
52. Gibson GR and Roberfroid MR. The bifidogenic nature of chicory inulin and its hydrolysis products. *J Nutr* 1995;125:1401.
53. Vervoort L, van den Mooter G, Augustijn P, Busson R, Toppet S and Kinget R. Inulin hydrogels as carriers for colonic drug targeting: I. Synthesis and characterization of methacrylated inulin and hydrogel formation. *Pharm Res* 1997; 14: 1730.
54. Maris B, Verheyden L, Reeth K V, Samyn C, van den Mooter G, Augustijn P and Kinget R. Synthesis and characterisation of inulin-azo hydrogels designed for colon targeting. *Int J Pharm* 2001; 213:143.
55. Stubbe B, Maris B, van den Mooter G and Demeester J. The in vitro evaluation of 'azo containing polysaccharide gels' for colon delivery. *J Control Rel* 2001; 75: 103.
56. Goldsten A M, Alter E N and Seaman JK. In: Whistler R.L., Eds., Oral colon specific drug delivery and their derivatives. Academic Press, Florida, 1992,1.
57. Tomlin J, Taylor J S and Read NW. The effects of mixed bacteria on a selection of viscous polysaccharides in vitro. *Nutr Rep Int* 1989;39: 121.
58. Baylis C E and Houtson AP. Degradation of guar gum by faecal bacteria. *Appl Environ Microbiol* 1986; 48: 626.
59. Johnson J C and Gee J M. Effect of gel-forming gums on the intestinal unstirred layer and sugar transport in vitro. *Gut* 1981; 22:398-403.
60. Gliko-Kabir I, Yagen B, Baluom M and Rubinstein A. Phosphated crosslinked guar for colon-specific drug delivery. II. *In vitro* and *in vivo* evaluation in the rat. *J Control Rel* 2000; 63: 129.
61. Krishnaiah Y S R, Seetha Devi A, Nageswara Rao L, Bhaskar Reddy P R, Karthikeyan RS and Satyanarayana V. Guar gum as a carrier for colon specific delivery; influence of metronidazole and tinidazole on *in vitro* release of albendazole from guar gum matrix tablets. *J Pharm Pharmaceut Sci* 2001;4: 235.
62. Krishnaiah Y S R, Veer Raju P, Dinesh Kumar B, Bhaskar P and Satyanarayana V. Development of colon targeted drug delivery systems for mebendazole. *J Control Rel* 2001; 77:87.
63. Wong D, Larrabeo S, Clitford K, Tremblay J and Friend D R. USP dissolution apparatus III (reciprocating cylinder) for screening of guar-based colonic delivery formulation. *J Control Rel* 1997;47: 173.
64. Ramaprasad Y V, Krishnaiah Y S R and Satyanarayana S. In vitro studies on guar gum based formulation for the colon targeted delivery of Sennosides. *J Control Rel* 1998; 51: 281.
65. Krishnaiah Y S R, Satyanarayana S, Dinesh Kumar B and Karthikeyan RS. Studies on the development of colon-targeted delivery systems for celecoxib in the prevention of colorectal cancer. *J Drug Target* 2002;10: 247.
66. Krishnaiah Y S R, Satyanarayana S, Dinesh Kumar B, and Karthikeyan RS. In vitro drug release studies on guar gum-based colon targeted oral drug delivery systems of 5-fluorouracil. *Eur J Pharm Sci* 2002; 16:185.
67. Das A, Wadhwa S and Srivastava AK. Cross-linked guar gum hydrogel discs for colon-specific delivery of ibuprofen: formulation and in vitro evaluation. *Drug Deliv* 2006;13 :139.
68. Al-Saidan SM, Krishnaiah YS, Satyanarayana V and Rao GS. In vitro and in vivo evaluation of guar gum-based matrix tablets of rofecoxib for colonic drug delivery. *Curr Drug Deliv* 2005; 2:155.
69. Krishnaiah YS, Indira Muzib Y and Bhaskar P. In vivo evaluation of guar gum-based colon-targeted drug delivery systems of ornidazole in healthy human volunteers. *J Drug Target* 2003;11:109-15.
70. Krishnaiah YS, Satyanarayana V, Dinesh Kumar B, Karthikeyan RS, Bhaskar P. Pharmacokinetic evaluation of guar gum-based colon-targeted oral drug delivery systems of metronidazole in healthy volunteers. *Eur J Drug Metab Pharmacokinet* 2002; 27:273.
71. Krishnaiah YS, Veer Raju P, Dinesh Kumar B, Satyanarayana V, Karthikeyan RS, and Bhaskar P. Pharmacokinetic evaluation of guar gum-based colon-targeted drug delivery systems of mebendazole in healthy volunteers. *J Control Rel* 2003; 88: 95.
72. Krishnaiah YS, Satyanarayana V, Dinesh Kumar B, Karthikeyan RS and Bhaskar P. In vivo pharmacokinetics in human volunteers: oral administered guar gum-based colon-targeted 5-fluorouracil tablets. *Eur J Pharm Sci* 2003 ;19 :355.
73. Krishnaiah YS, Muzib YI, Bhaskar P, Satyanarayana V and Latha K. Pharmacokinetic evaluation of guar gum-based colon-targeted drug delivery systems of tinidazole in healthy human volunteers *Drug Deliv* 2003;10:263-8.
74. Krishnaiah YS, Veer Raju P, Dinesh Kumar B, Jayaram B, Rama B, Raju V and Bhaskar P.

Pharmacokinetic evaluation of guar gum-based colon-targeted oral drug delivery systems of metronidazole in healthy volunteers. *Eur J Drug Metab Pharmacokinet* 2003; 28: 287.

75. Momin M and Pundarikakshudu K. In vitro studies on guar gum based formulation for the colon targeted delivery of Sennosides. *J Pharm Pharm Sci* 2004; 7:325.

76. Chourasia MK and Jain SK. Potential of guar gum microspheres for target specific drug release to colon. *J Drug Target* 2004;12: 435.

77. Krishnaiah YS, Seetha Devi A, Nageswara Rao L, Bhaskar Reddy PR, Karthikeyan RS and Satyanarayana V. Guar gum as a carrier for colon specific delivery; influence of metronidazole and tinidazole on in vitro release of albendazole from guar gum matrix tablets. *J Pharm Pharm Sci* 2001; 4:235.

78. Krishnaiah YS, Satyanarayana S and Prasad YV. Studies of guar gum compression-coated 5-aminosalicylic acid tablets for colon-specific drug delivery. *Drug Dev Ind Pharm* 1999; 25: 651.

79. Milojevic S, Newton J M, Cummings J H , Gibson GR, Botham R L , Ring, S C, Stockham M and Allwood M.C. Amylose as a coating for drug delivery to the colon: Preparation and in vitro evaluation using 5-aminosalicylic acid pellets. *J Control Rel* 1996; 38: 75.

80. Wilson PJ and Basit AW. Exploiting gastrointestinal bacteria to target drugs to the colon: an in vitro study using amylose coated tablets. *Int J Pharm* 2005; 300: 89.

81. Siew LF, Basit AW and Newton JM. The properties of amylose-ethylcellulose films cast from organic-based solvents as potential coatings for colonic drug delivery. *Eur J Pharm Sci* 2000 ;11(2):133-9.

82. Siew LF, Basit AW and Newton JM. The potential of organic-based amylose-ethylcellulose film coatings as oral colon-specific drug delivery systems. *AAPS Pharm SciTech* 2000 ;1(3): E22.

83. Tuleu C, Basit AW, Waddington WA, Ell PJ and Newton JM. Colonic delivery of 4-aminosalicylic acid using amylose-ethylcellulose-coated hydroxypropylmethylcellulose capsules. *Aliment Pharmacol Ther* 2002;16:1771.

84. Cummings J H, Milojevic S, Newton JM, Cummings J H, Gibson GR, Botham R L, Ring S C, Stockham M and Allwood M C. In vivo studies of amylose- and ethylcellulose-coated [¹³C]glucose microspheres as a model for drug delivery to the colon. *J Control Rel* 1996;40: 123.

85. Lenaerts V, Dumoulin Y and Mateescu M A. Controlled-release of theophylline from cross-linked amylose tablets. *J Control Rel* 1991;15: 39-46.

86. Salyers AA and O'Brein M. Cellular location of enzymes involved in chondroitin sulfate breakdown by *Bacteroides thetaiotaomicron*. *J Bacteriol* 1980;143: 772.

87. Salyers, A.A. Energy sources of major intestinal fermentative anaerobes. *Am J Clin Nutr* 1979; 32:158.

88. Rubinstein A, Nakar D and Sintov A. A Potential Biodegradable Carrier for Colon Specific Drug Delivery. *Int J Pharm* 1992;84:141.

89. Rubinstein A, Nakar D and Sintov A. Colonic Drug Delivery: Enhanced Release of Indomethacin from Cross-Linked Chondroitin Sulfate Matrix in Rat Cecal Content. *Pharm Res* 1992; 9: 276.

90. Sintov A, Di-capua N and Rubinstein A. Cross-linked chondroitin sulphate: characterization for drug delivery purposes. *Biomaterials* 1995;16: 473.

91. Antenucci R and Palmer J K. Enzymatic degradation of alpha- and beta cyclodextrins by bacteroides of the human colon. *J Agric Food Chem* 1984;32:1316-1321.

92. Flourie B , Molis C, Achour L, Dupas H , Hatat C and Rambaud JC. Fate of beta -cyclodextrin in the human intestine. *J Nutr* 1993;123: 676.

93. Tanaka H, Kominato K and Yamamoto R. Synthesis of doxorubicin cyclodextrin conjugates. *J Antibio* 1994; 47:1025.

94. Hiramaya F, Minami K and Uekama K. In vitro evaluation of biphenyl acetic acid-beta-cyclodextrin conjugates as colon-targeting prodrugs: Drug release behaviour in rat biological media. *J Pharm Pharmacol* 1996;48: 27.

95. Yano H, Hirayama F., Kmada M, Arima H, Uekama K and Uekama K. Colon-specific delivery of prednisolone-appended alpha-cyclodextrin conjugate: alleviation of systemic side effect after oral administration. *J Control Rel* 2002;79: 103.

96. Zou MJ, Cheng G, Okamoto H, Hao XH, An F, Cui FD, Danjo K. Colon-specific drug delivery systems based on cyclodextrin prodrugs: in vivo evaluation of 5-aminosalicylic acid from its cyclodextrin conjugates. *World J Gastroenterol* 2005;11: 7457.

97. Minami K, Hirayama F and Uekama K. Colon-specific drug delivery based on cyclodextrin prodrug: release behaviour of biphenyl acetic acid form its cyclodextrin conjugates in rat intestinal tract after oral administration. *J Pharm Sci.* 1998; 87: 715.

98. Omrod D J, Holmes C C and Miller TE. Dietary chitosan inhibits hypercholesterolemia and atherosclerosis in the apolipoprotein E-deficient mouse model of atherosclerosis. *Atherosclerosis* 1998;138: 329.

99. Weiner M L, In: Brine, C J, Sandford PA, Zikakis J P, Eds., *Advances in chitin and chitosan*, Elsevier applied sciences, London, 663-672.

100. Tozaki M, Komoike J, Tada C, Maruyama T, Terabe A, Suzuki T , Yamamoto A and Muranishi S. Chitosan capsules for colon-specific drug delivery: Improvement of insulin absorption from the rat colon. *J Pharm Sci* 1997;86:1016.

101. Tozaki M, Odoriba T, Okada N, Fujita T, Okabe S, Terabe A, Suzuki T, Yamamoto A and Muranishi S. Chitosan capsules for colon-specific drug delivery: enhanced localization of 5-aminosalicylic acid in the

103. large intestine accelerates healing of TNBS-induced colitis in rats. *J Control Rel* 2002; 82: 51.
102. Tozaki M, Odoriba T and Fujita T. Chitosan capsules for colon-specific drug delivery: enhanced localization of 5-aminosalicylic acid in the large intestine accelerates healing of TNBS-induced colitis in rats. *Life Sci* 1999; 64:1155.
103. Shimono N, Takatori T, Masumi T, Ueda M, Mori M, Higashi Y and Nakamura Y. Chitosan dispersed system for colon-specific drug delivery. *Int J Pharm* 2002;245: 45.
104. Orienti I, Cerchiara T, Luppi B, Bigucci F, Zuccari G and Zecchi V. Influence of different chitosan salts on the release of sodium diclofenac in colon-specific delivery. *Int J Pharm* 2002; 238: 51.
105. Vandelli M A, Leo E, Forni F and Bernatei M T. *In vitro* evaluation of a potential colonic drug delivery system that releases drug after a controllable lag-time. *Eur J Pharm Biopharm* 1996;43:148.
106. Shu X Z, Zhu K J and Song W. Novel pH-sensitive citrate crosslinked chitosan film for drug controlled release. *Int J Pharm* 2001; 212: 19.
107. Suzuki T, Matsumoto T, Hagino Y, Tozaki H, Yamamoto A and Muranishi S. In: Prasad P, Eds., *Biodegradation and medical applications of chitosan hard capsules*, Plenum Press, Ny, 1998, 567.
108. Rai G, Jain SK, Agrawal S, Bhadra S, Pancholi SS and Agrawal GP. Chitosan hydrochloride based microspheres of albendazole for colonic drug delivery. *Pharmazie* 2005;60:131.
109. Harboe E, Larsen C, Johansen M and Olesen H P. Macromolecular prodrugs. XV. Colon targeted delivery. Bioavailability of naproxen from orally administered dextran ester prodrugs varying molecular size in the pig. *Int J Pharm* 1989; 213: 93.
110. Harboe E, Larsen C and Johansen M. Macromolecular prodrugs. XIV. Absorption characteristics of naproxen after oral administration of a dextran T-70-naproxen ester prodrugs in pigs. *Farmaci Sci* 1988;16: 73.
111. Larsen C, Harboe E, Johansen M and Olesen H.P. Macromolecular prodrugs. XVI. Colon targeted delivery. Comparison of the rate of release of naproxen from dextran ester prodrug in homogenates of various segments of the pig gastrointestinal tract. *Pharm Res* 1989; 6: 995.
112. Larsen C, Harboe E, Johansen M and Olesen H.P. Macromolecular prodrugs. XV. Colon-targeted delivery--bioavailability of naproxen from orally administered dextran-naproxen ester prodrugs varying in molecular size in the pig. *Pharm Res* 1989; 6: 919.
113. Johansen M and Larsen C. Stability kinetics and of hydrolysis of metronidazole monosuccinate in aqueous solution and in plasma. *Int J Pharm* 1984; 21: 201.
114. Johansen, M and Larsen C. A comparison of the chemical stability and the enzymatic hydrolysis of a series of aliphatic and aromatic ester derivatives of metronidazole. *Int J Pharm* 1985; 27: 219.
115. Vermeersch J, Vandoorne F, Permentier D and Schacht E. Macromolecular prodrugs of metronidazole 1. Esterification of hydroxyl containing Polymers with metronidazole monosuccinate. *Bull Soc Chim Belg* 1985; 94: 591.
116. Lee J S, Jung YJ, Doh M J and Kim Y M. Synthesis and properties of dextran-nalidixic acid ester as a colon-specific prodrug of nalidixic acid. *Drug Dev Ind Pharm* 2001; 27: 331-336.
117. Jung Y J, Lee J S, Kim Y M, Kim Y K and Kim H H. Synthesis and properties of dextran-5-aminosalicylic acid ester as a potential colon-specific prodrug of 5-aminosalicylic acid. *Arch Pharm Res* 1998; 21:179.
118. Bauer K H and Kesselhut J F. Novel pharmaceutical excipients for colon targeting. *STP Pharm Sci* 1995; 5: 54.
119. Hirsch S, Binder V, Kolter K, Kesselhut J F and Bauer K H. Lauroyldextrin as a coating material for site-specific drug delivery to the colon: In vitro dissolution of coated tablets. *Proc Int Contrl Bioact Mater* 1997; 24: 379.
120. Brondsted H, Andersen C and Hovgaard L. Crosslinked dextran - a new capsule material for colon targeting of drugs. *J Control Rel* 1998;53:7.
121. Friend D R, Philips S, McLeod A and Tozer T N. Relative anti-inflammatory effect of oral dexamethasone-b-D-glucoside and dexamethasone in experimental inflammatory bowel disease in guinea pig. *Proc Int Contrl Bioact Mater* 1991; 18: 612.
122. Tozer T N, Rigod J, McLeod A, Gungon R, Hong M K and Friend D R. Colon-specific delivery of dexamethasone from a glucoside prodrug in the guinea pig. *Pharm Res* 1991; 8: 445.
123. McLeod A D, Friend DR and Tozer T N. Dextran Esters: Potential Prodrugs for Colon Specific Delivery. *Int J Pharm* 1993; 92:105.
124. McLeod A D, Friend D R, Fedorak R N, Tozer T N and Cui N. A glucocorticoid prodrug facilitates normal mucosal function in rat colitis without adrenal suppression. *Gastroenterology* 1994;106:405.
125. McLeod A D, Tozer T N and Tolentino L. Colon-specific delivery of dexamethasone from a glucoside prodrug in the guinea pig. *Biopharm Drug Dispos* 1994;15:151.
126. Zhou SY, Mei QB, Liu L, Zhang BL, Li C and Zhou J. Different molecular weight dextran - linking of dexamethasone in vitro release of targeting. *Yao Xue Xue Bao* 2003;38:388.
127. Zhou SY, Mei QB, Zhou J, Liu L, Li C and Zhao DH. Glucocorticoid precursor drugs in the rat gastrointestinal position to address the pharmacokinetics. *Yao Xue Xue Bao.*, 2001, 36, 325.

128. Pang YN, Zhang ZR, Pang QJ and Li TL. Site-specific drug release in vitro of prednisolone-dextran conjugates with different molecular mass. Yao Xue Xue Bao 2001; 36:625.
129. Shrivastava SK, Jain DK and Trivedi P. Dextrans--potential polymeric drug carriers for suprofen. Pharmazie 2003; 8: 804.
130. Shrivastava SK, Jain DK and Trivedi P. Dextrans--potential polymeric drug carriers for flurbiprofen. Pharmazie 2003; 58:389.
131. Shun YL and Ayres J W. Calcium alginate beads as core carriers of 5-aminosalicylic acid. IntJPharm1992; 212:19.
132. Lin SY, Ayres JW. Calcium Alginate Beads as Core Carriers of 5-Aminosalicylic Acid", Pharm.Res.,1992;9:1128.
133. Kiyoun L, kun N and Yueim K. Polysaccharides as a drug coating polymer. Polym Prep1999;40: 359.
134. Raghavan CV, Muthulingam C, Leno L, Jenita JAJ and Ravi TK. An in vitro and in vivo investigation into the suitability of bacterially triggered delivery system for colon targeting. Int J Pharm 2002;238:51
