

Monoterpene Indole Alkaloid from *Aframomum Meleguata*

Donatus Ebere Okwu*. Eunice Ego Njoku

Department of Chemistry, Michael Okpara University of Agriculture, Umudike,
P.M.B. 7267 Umuahia, Abia State, Nigeria

*Corres.author: okwudonatus@yahoo.com

ABSTRACT: As part of our study on bioactive agents from Nigerian medicinal plants, a new monoterpene indole alkaloid (10, 12-dihydroxy-18-ethenyl-4-pyrido- β -carboline **1**) was isolated from the leaves of *Aframomum meleguata*. The structure of **1** was elucidated using NMR spectroscopy in combination with IR and MS spectral data.

Key words: *Aframomum meleguata*, Monoterpenes, Indole alkaloid, herbal medicine.

INTRODUCTION

Species of the *Zingiberaceae*, (ginger family) have been used for various medicinal purposes in Nigeria (Gill 1992, Iwu 1993), and our systematic studies on Nigerian medicinal plants of this family have revealed marked diversity of the secondary metabolites present and their biological activities (Okwu 2007, Okwu and Njoku 2009). As part of our search for bioactive compounds from endemic species from Nigeria rainforest, *A. meleguata* was studied further after detection of antifungal activity in its aqueous and ethanol leaf extracts (Okwu and Njoku 2009).

A. meleguata is commonly known as guinea pepper, meleguata pepper, alligator pepper, guinea grains or grains of paradise. It is a herbaceous perennial plant, native to swampy habitats along the West African Coast (Iwu *et al* 1999). Its trumpet shaped purple flowers develop into 5 to 7 cm long pods containing reddish brown seeds. The seeds have a pungent, peppery taste due to aromatic ketones such as gingerol and paradol (Iwu 1993). *A. meleguata* is used as a fungicide. Antifungal effects of the leaf extract of *A. meleguata* on spore germination and mycelial reduction of the most occurring fungal pathogens causing soft rots of white yam (*Disoscorea rotundata*) tuber have been investigated (Okigbo and Ogbonnaya 2006). Fungi isolated from rotten yams were *Aspergillus flavus*, *Fusarium oxysporum*, *Rhizopus stoloniger*, *Botryodiplodia theobromae* and *Penicillium chrysogenum*. The ethanolic leaf extract followed by cold water and hot water extracts were

most effective on these pathogens (Okigbo and Ogbonnaya 2006). *A. meleguata* is very effective against major disease causing micro-organisms such as *Escherichia coli*, *Candida albicans*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Vibrio cholera*, *Salmonella spp*, *Streptococcus spp* and *Neisseria gonorrhoea* (Iwu *et al* 1999). The plant has been extensively used in herbal medicine not only for its oxytocic, analgesic, anti-inflammatory and antimicrobial properties but also provides relief in the treatment of human gastro-intestinal, hypermobility and peptic ulceration (Gill 1992). It is generally used as a stimulant and febrifuge. The decoction of the leaves is used for small pox and chicken pox (Gill 1992, Okwu 2005, 2007). The seeds of *A. meleguata* and fruits of *Spondias mombin* are powdered and mixed with cold pap as an anti-fertility agent (Gill 1992, Okwu 2007). It is also used for male sexual impotence (Bitti 2002). Both the leaves and the seeds of *A. meleguata* are used for inflamed condition of throat, fever and exanthemata when crushed and rubbed on the body. The plant is used in phytomedicine to cure rheumatoid arthritis, bronchitis, cough and bone treatment (Okwu 2007). They are also applied for headache as well as made into paste for wounds and sores (Okigbo and Ogbonnaya 2006). The other medicinal uses of *A. meleguata* include aphrodisiac, measles and leprosy (Iwu 1993). It is also taken for excessive lactation and hemorrhage, purgative, galactagogue and as haemostatic agent (Iwu 1993). In addition, the plant possesses anti-diabetic properties (Iwu 1993). *A. meleguata* is used as

carminative, as a cough remedy and lactation aid (Iwu *et al* 1999). Other uses are stomachache bronchitis and dysentery. It is used with lemon grass (*Cymbopogon citrates*) for female hygiene (Iwu *et al* 1999). The seeds, leaves and rhizomes are used in West African herbal medicine as stimulant, carminative and diuretic (Iwu *et al* 1999). It is used to treat snake bites and scorpion stings (Duke 2000). In spite of the various uses of *A. meleguata* in herbal medicine in Nigeria and as spices in food and drug production in Nigeria, the phytoconstituents of this herb have not been fully documented. In this paper, the details of the isolation and structure elucidation of a new monoterpene indole alkaloid (10, 12-dihydroxy-18-ethenyl-4-pyrido- β -carboline) from the leaves of *A. meleguata* was presented.

EXPERIMENTAL SECTION

General experimental procedure

The IR spectra were determined on a Thermo Nicolet Nexus 470 FT-IR spectrometer. The ^1H and ^{13}C NMR spectra were recorded on a Bruker Avance 400 FT NMR spectrometer using TMS as internal standard. Chemical shifts are expressed in parts per million (ppm). EIMS were performed with a Bruker APEX II mass spectrometer and ESIMS were recorded in the Q-STAR ESI-TOF-MS/MS Spectrometer. HREIMS were obtained on a GCT-MS instrument. Column chromatography was carried out with silica gel (200-300 mesh) and to monitor the preparative separations, analytical thin layer chromatography (TLC) was performed at room temperature on pre-coated 0.25mm thick silica gel 60 F₂₅₄ aluminum plates 20 x 20 cm Merck; Darmstadt, Germany. Reagents and solvents like ethanol, chloroform, diethyl ether, hexane were all of analytical grade and were procured from Merck, Darmstadt, Germany.

Plant materials

The fresh leaves of *A. meleguata* were harvested from Ibekuta Ibeku, Okwuato Aboh Mbaise Local Government of Imo State, Nigeria on 15th March 2007. The plant samples (fruits, seeds and leaves) were identified by Dr. A. Nmeregini of Taxonomy Section, Forestry Department, Michael Okpara University of Agriculture, Umudike, Nigeria. Voucher specimen No. AF/3355 has been deposited at the Forestry Department Herbarium of the University.

Extraction and isolation plant materials

Plant materials were treated and analyzed at the Chemistry laboratory, Michael Okpara University of Agriculture Umudike, Nigeria. The leaves (1kg) were dried on the laboratory bench for 10 days. The dry sample was milled and ground into powder (950g) using Thomas Wiley machine (model 5 USA). The

powdered plant materials were dried and stored in air tight bottles for chemical analysis. The powder plant sample (500g) was packed into a soxhlet apparatus (2 L) and extracted exhaustively with 1000 ml chloroform for 24 h. The chloroform extract was concentrated using a rotary evaporator at 45°C and in a hot air circulating oven to get dark brown oil (10.9g). The column chromatography of the extract was carried out using the chloroform fractions. The column was packed with silica gel and eluted with methanol: chloroform: petroleum ether (20: 30: 50) to get brown oil (0.86mg). The oil gave a single spot on TLC (R_f 0.4667). IR V_{max} 3401 cm⁻¹ (OH), 3008 cm⁻¹ (N-H), 2926 cm⁻¹ (-CH₂-), 1244 (-C-N). HEREIMS m/z 281.2439 [M⁺]; calculated for m/z 282 C₁₇H₁₈O₂N₂. ^1H NMR and ^{13}C NMR were presented in Table 1.

RESULTS AND DISCUSSION

Compound **1** was isolated as dark brown oil with strong fluorescence detected at 354 nm which is compatible with an aromatic 5,6-dihydro- β -carboline chromophore, as corroborated by an indole N-H IR band (3008 cm⁻¹). The IR spectrum showed absorptions of V_{max} 3401, 3008, 2926 and 1464 cm⁻¹, indicating the presence of -OH, NH, CH₂ and C=C-aromatic functional groups respectively.

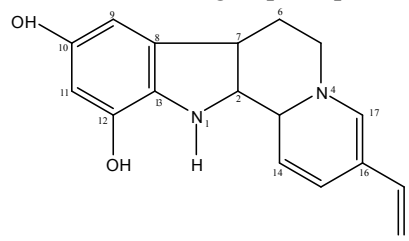


Figure 1: Compound **1** C₁₇H₁₈O₂N₂

The HRESIMS of **1** in the positive-ion mode, exhibited a molecular ion peak [M+H] at m/z 281.2439 calculated for C₁₇H₁₈O₂N₂ m/z 282. The ^1H NMR spectrum (Table 1) revealed the presence of a tetrahydro- β -carboline system due to the signals of δH 4.1675 (1Hs), 4.1397 (1Hs), 2.3275 (2Hs), 2.0204 (2Hd), 1.2775 (1Hs) and 7.4214 (1Hs). This spectrum also shows the presence of the pyridine protons at δH 5.3243 (1Hs) and 5.3281 (1Hs) and 4.1952 respectively.

The ^{13}C NMR spectrum (Table 1) confirmed a tetrahydro- β -carboline monoterpene structure for **1**. The ^{13}C NMR showed the presence of a terminal vinyl group for **1** at δC 128.134 and the presence of the signals for the aromatic carbon at δC 130.312, 129.798, and 129.798. Apart from the molecular ion peak at m/z 281.2439, the high resolution mass spectrum gave fragment peaks at m/z 280 and 279 respectively due to proton migration and

rearrangement. Detachment of the hydroxyl group produces the peak at m/z 264.2449 calculated for $C_{17}H_{15}ON_2$ (m/z 263). Further fragmentation afforded the base peak m/z 83.9525 and the peak m/z 81.0706 calculated for $C_5H_7N_2$ (m/z 81). The pattern of fragmentation of compound **1** is shown in figure 2. This analysis confirmed compound **1** isolated from the leaves of *A. meleguata* to be a monoterpene indole alkaloid 10,12-dihydroxy-18-ethenyl-4-pyrindo- β -carboline. Indole alkaloids containing a monoterpene moiety as in **1** is unusual. The isolation of

monoterpene indole alkaloid from *A. meleguata* supported the use of this herb in phytomedicine for the treatment of rheumatoid arthritis. This work therefore shows that monoterpene indole alkaloid is one of the physiologically bioactive constituents of *A. meleguata*. The occurrence of monoterpene indole alkaloid in *A. meleguata* is of significance because this is to the best of our knowledge the first report of its occurrence in any *Aframomum* species. These findings supported the use of *A. meleguata* as raw materials for food and pharmaceutical industries.

Table 1: 1H (400 MHz and ^{13}C NMR (400 MHz) Chemical shifts of Compound **1**

Position	δ H	δ C
1	4.1675	1Hs
2	1.2553	1Hs
3	4.1397	1Hs
4		
5	2.3275 – 2.7705	2Hd
6	2.0204 – 2.0573	2Hd
7	1.2775	1Hs
8		
9	7.4214	1Hs
10	4.1397	1Hbs
11	7.4214	1Hs
12	4.1531	1Hbs
13		
14	5.3243	1Hs
15	5.3281	1Hs
16		
17	5.3360	1Hs
18	5.3434	1Hs
19	4.1952	2Hs
		=CH ₂

s = singlet, d = doublet, m = multiplet, t = triplet, bs = broad singlet

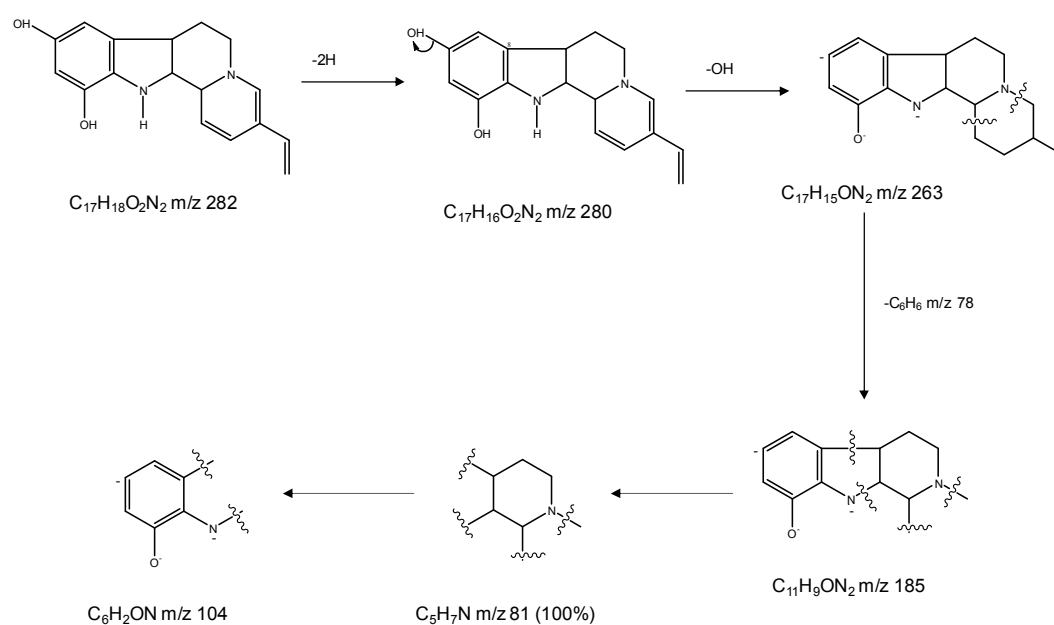


Figure 2: Fragmentation pattern of compound **1**

REFERENCES

- Agoha R. C. (1974) Medicinal plants of Nigeria. Offset Arakkenji, Raculfeider Wiskunde. The Netherlands Pp. 33, 4
- Apata L. (1979) The practice of herbalist in Nigeria. In Sofowara A Ed. African Medicinal plants. University of Ife Press Nigeria PP. 13 – 19.
- Betti M. (2002) Medicinal plants sold in Yaunde markets, Cameroon. African Study Monographs 23: 47 – 64.
- Duke J. A. (2000) Phytochemical data base USDA-ARS-NGRL, Research Center Beltsville Maryland.
- Gill L. S. (1992) Ethnomedical uses of plants in Nigeria University of Benin Press Benin City Edo State Nigeria Pp. 15
- Iwu M. M. (1993) Handbook of African medicinal Plants CRC Press Boca Raton Florida PP. 263 – 269.
- Iwu M. M.; Duncan A. R. and Okunji C. O. (1999) New anti-microbial of plant origin. Perspectives on new crops and new uses. ASHS Press Alexandria V. A. Pp. 457 – 462
- Okigbo RN, Ogbonnaya UO (2006) Antifungal effects of two tropical plant leaf extracts of (*Ocimum gratissimum* and *Aframomium meleguata*) on post harvest yam (*Dioscorea spp*) rot. *African Journal of Biotechnology* 5: 727 – 731.
- Okwu D. E. (2005) Phytochemicals, vitamins and mineral contents of two Nigerian Medicinal plants. *International Journal of molecular medicine and Advance Sciences* 1(1): 375 – 381.
- Okwu D. E. (2007) Nigerian medicinal plants 11. *Medicinal and Aromatic Plant Sciences and Biotechnology* 1(1): 97 – 102.
- Okwu DE, Njoku EE (2009) Chemical Composition and in vitro Antifungal Activity Screening of Seed and Leaf Extracts from *Aframomum meleguata* and *Monodora myristica* against *Sclectium rolfsii* of Cowpea plant *Vigna unguiculata* L. walp *Pest Technology* 3(1): (In Press).
