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# *In-vitro* antioxidant potential of various extracts from whole plant of *Bauhinia purpurea* (Linn)

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**Abstract:** The objective of the present investigation was to evaluate the *in-vitro* antioxidant potential various extracts of whole plant of *Bauhinia purpurea* with three *in-vitro* methods: DPPH scavenging activity, superoxide anion scavenging activity and estimation of total flavonoids. The ethyl acetate extract of *Bauhinia purpurea* was showed significant result in DPPH scavenging and superoxide anion scavenging activity methods. Similar result was not found in other two extracts. High amount of flavonoids were found in ethyl acetate extract of *Bauhinia purpurea* than that of other two extracts. The high antioxidant capacity observed for ethyl acetate extract of *Bauhinia purpurea* suggested that this plant could be used as an additive in the food industry providing good protection against oxidative damage. **Key words:** *Bauhinia purpurea*, DPPH scavenging activity, superoxide anion scavenging activity, total flavonoids.

## Introduction

Reactive oxygen species (ROS) including superoxide radicals, hydroxyl radicals, singlet oxygen and hydrogen peroxide are often generated as byproducts of biological reactions, or from exogenous factors<sup>1</sup>. *In vivo*, some of these ROS play positive roles such as energy production, phagocytosis, regulation of cell growth and intercellular signalling, or synthesis of biologically important compounds<sup>2</sup>. However, ROS may also be very damaging as they can induce oxidation of lipids, causing membrane damage, decreasing membrane fluidity, and leading to cancer via DNA mutation<sup>3</sup>. A potent scavenger of these ROS may serve as a possible preventative against free radical mediated diseases<sup>4</sup>.

*Bauhinia purpurea* Linn. (Leguminosae) is a medium sized deciduous tree, sparingly grown in India. This plant is used traditionally in dropsy, pain,

rheumatism, convulsions, delirium, and septicemia<sup>5</sup>. The bark of the plant is used as an astringent in the treatment of diarrhea. Its decoctions are recommended for ulcers as a useful was solution<sup>6</sup>. They are reported to exhibit various pharmacological activities such as CNS activity, cardiotonic activity, lipid-lowering activity, anti-oxidant activity, hepatoprotective activity, hypoglycemic activity, etc<sup>7</sup>. Even through, traditionally, leaves of *Bauhinia purpurea* (Linn) were extensively used for the treatment of variety of wounds<sup>8</sup>, and no scientific data in its support is available.

Literature survey revealed that there is a lack of enough scientific reports regarding the antioxidant properties of whole plant of *Bauhinia purpurea* (Linn). Hence the objective of the present investigation was to evaluate the *in-vitro* antioxidant potential of various extracts from whole plant of *Bauhinia purpurea* (Linn) with various *in-vitro* antioxidant methods.

#### **Material and Methods**

#### **Collection and Identification of Plant materials**

The whole plant of *Bauhinia purpurea* (Linn), were collected from Nagercoil, Kanyakumari District of Tamil Nadu, India. Taxonomic identification was made from Botanical Survey of Medical Plants Unit Siddha, Government of India. Palayamkottai. The whole plant of *Bauhinia purpurea* (Linn), were dried under shade, segregated, pulverized by a mechanical grinder and passed through a 40 mesh sieve.

#### **Preparation of Extracts**

The above powered materials were successively extracted with Petroleum ether  $(40-60^{\circ}C)$  by hot continuous percolation method in Soxhlet apparatus<sup>9</sup> for 24 hrs. Then the marc was subjected to Ethyl acetate (76-78°C) for 24 hrs and then marc was subjected to Methanol for 24 hrs. The extracts were concentrated by using a rotary evaporator and subjected to freeze drying in a lyophilizer till dry powder was obtained.

#### Determination of Antioxidant activity DPPH photometric assay<sup>10</sup>

The effect of extract on DPPH radical was assayed using the method of Mensor et al (2001)<sup>10</sup>. A methanolic solution of 0.5ml of DPPH (0.4mM) was added to 1 ml of the different concentrations of plant extract and allowed to react at room temperature for 30 minutes. Methanol served as the blank and DPPH in methanol without the extracts served as the positive control. After 30 min, the absorbance was measured at 518 nm and converted into percentage radical scavenging activity as follows.

 $Scavengingactivity(\%) = \frac{A_{518} \text{ Control} - A_{518} \text{ Sample}}{A_{518} \text{ Control}} \times 100$ 

Where  $A_{518}$  control is the absorbance of DPPH radical+ methanol;  $A_{518}$  sample is the absorbance of DPPH radical+ sample extract/ standard.

#### Superoxide radical scavenging activity<sup>11</sup>

Superoxide radical ( $O_2^{-1}$ ) was generated from the photoreduction of riboflavin and was deducted by nitro blue tetrazolium dye (NBT) reduction method. Measurement of superoxide anion scavenging activity was performed based on the method described by Winterbourne et al (1975)<sup>11</sup>. The assay mixture contained sample with 0.1ml of Nitro blue tetrazolium (1.5 mM NBT) solution, 0.2 ml of EDTA (0.1M EDTA), 0.05 ml riboflavin (0.12 mM) and 2.55 ml of phosphate buffer (0.067 M phosphate buffer). The

control tubes were also set up where in DMSO was added instead of sample. The reaction mixture was illuminated for 30 min and the absorbance at 560 nm was measured against the control samples. Ascorbate was used as the reference compound. All the tests were performed in triplicate and the results averaged. The percentage inhibition was calculated by comparing the results of control and test samples.

#### **Total flavonoids**<sup>12</sup>

0.2g of the plant material was ground with ethanol-water in 2 different ratios namely 9:1 and 1:1 respectively. The homogenate was filtered and these 2 ratios were combined. This was evaporated to dryness until most of the ethanol has removed. The resultant aqueous extract was extracted in a separating funnel with hexane or chloroform. The solvent extracted aqueous layer was concentrated 0.5 ml of aliquot of extract was pipette-out in a test tube. 4 ml of the vanillin reagent (1% vanillin in 70% conc. H<sub>2</sub>SO<sub>4</sub>) was added and kept in a boiling water bath for 15 mins. The absorbance was read at 360 nm. A standard was run by using catechol (110 µg/ml).

#### **Results and Discussion**

Antioxidant compounds may function as free radical scavengers, initiator of the complexes of prooxidant metals, reducing agents and quenchers of singlet oxygen formation<sup>13</sup>. Phenolic compounds and flavonoids are major constituents of most of the plants reported to possess antioxidant and free radical scavenging activity<sup>14</sup>.

#### **DPPH scavenging activity**

DPPH is a stable free radical at room temperature often used to evaluate the antioxidant activity of several natural compounds. The reduction capacity of DPPH radicals was determined by the decrease in its absorbance at 517 nm, which is induced by antioxidants.

The percentage of DPPH radical scavenging activity of petroleum ether extract of *Bauhinia purpurea* was presented in Table 1. The petroleum ether extract of *Bauhinia purpurea* and Rutin (standard) showed DPPH free radical scavenging activity in a concentration range of 125-1000  $\mu$ g/ml and its inhibition ranged from 15-33% and 18-69% respectively. The IC<sub>50</sub> values of the petroleum ether extract of *Bauhinia purpurea* and Rutin were found to be 1440 $\mu$ g/ml and 480 $\mu$ g/ml respectively.

S.No	Concentration (µg/ml)	% of activity(±SEM)*			
5.110		Sample (Petroleum ether extract)	Standard (Rutin)		
1	125	$15.97 \pm 0.07$	$18.85 \pm 0.07$		
2	250	$19.78 \pm 0.03$	$22.08 \pm 0.05$		
3	500	$25.18 \pm 0.02$	$52.21 \pm 0.02$		
4	1000	$33.05 \pm 0.11$	$69.83 \pm 0.01$		
		$IC_{50} = 1440 \ \mu g/ml$	$IC_{50} = 480 \ \mu g/ml$		

Table 1: Effect of Petroleum ether extract of *Bauhinia purpurea* on DPPH assay

\*All values are expressed as mean  $\pm$  SEM for three determinations

The percentage of DPPH radical scavenging activity of ethyl acetate extract of *Bauhinia purpurea* was presented in Table 2. The ethyl acetate extract of *Bauhinia purpurea* and Rutin (standard) showed DPPH free radical scavenging activity in a concentration range of 125-1000  $\mu$ g/ml and its inhibition ranged from 24-69% and 18-69% respectively. The IC<sub>50</sub> values of the ethyl acetate extract of *Bauhinia purpurea* and Rutin were found to be 290 $\mu$ g/ml and 480 $\mu$ g/ml respectively.

The percentage of DPPH radical scavenging activity of methanolic extract of *Bauhinia purpurea* was presented in Table 3. The methanolic extract of *Bauhinia purpurea* and Rutin (standard) showed DPPH free radical scavenging activity in a concentration range of 125-1000  $\mu$ g/ml and its inhibition ranged from 19-67% and 18-69% respectively. The IC<sub>50</sub> values of the methanolic extract of *Bauhinia purpurea* and Rutin were found to be 570 $\mu$ g/ml and 480 $\mu$ g/ml respectively.

Table 2: Effect of Ethyl acetate extract of Bauhinia purpurea on DPPH assay

	¥	% of activity(±SEM)*			
S.No	Concentration (µg/ml)	Sample (Ethyl acetate extract)	Standard (Rutin)		
1	125	$24.04 \pm 0.08$	$18.85 \pm 0.07$		
2	250	$47.67 \pm 0.07$	$22.08 \pm 0.05$		
3	500	$63.68 \pm 0.04$	$52.21 \pm 0.02$		
4	1000	$69.22 \pm 0.03$	$69.83 \pm 0.01$		
		$IC_{50} = 290 \ \mu g/ml$	$IC_{50} = 480 \ \mu g/ml$		

\*All values are expressed as mean  $\pm$  SEM for three determinations

		% of activity(±SEM)*		
S.No	Concentration (µg/ml)	Sample (Methanolic extract)	Standard (Rutin)	
1	125	$19.51 \pm 0.01$	$18.85\pm0.07$	
2	250	$29.31 \pm 0.04$	$22.08\pm0.05$	
3	500	$45.62 \pm 0.03$	$52.21 \pm 0.02$	
4	1000	$67.23 \pm 0.02$	$69.83 \pm 0.01$	
		$IC_{50} = 570 \ \mu g/ml$	$IC_{50} = 480 \ \mu g/ml$	

 Table 3: Effect of Methanolic extract of Bauhinia purpurea on DPPH assay

\*All values are expressed as mean  $\pm$  SEM for three determinations

From the above result clearly indicated that the ethyl acetate extract of *Bauhinia purpurea* exhibits significant DPPH scavenging activity when compared with standard Rutin. Similar result was not found in other two extracts. The  $IC_{50}$  values of ethyl acetate extract of *Bauhinia purpurea* and Rutin were observed at 290µg/ml and 480µg/ml respectively.

#### Superoxide anion scavenging activity

Superoxide is a highly reactive molecule that reacts with various substances produced through metabolic processes. Superoxide dismutase enzymes present in aerobic and anaerobic organisms catalyses the breakdown of superoxide radical<sup>15</sup>. Superoxide scavenging ability of plant extract might primarily be due to the presence of flavanoids<sup>16</sup>

The percentage of superoxide anion scavenging activity of petroleum ether extract of *Bauhinia purpurea* was presented in Table 4. Maximum scavenging activity of plant extract and Quercetin at 1000  $\mu$ g/ml was found to be 44.35% and 98.01% respectively. The IC<sub>50</sub> value of plant extract

and Quercetin was recorded as  $1390\mu$ g/ml and  $60\mu$ g/ml respectively.

The percentage of superoxide anion scavenging activity of ethyl acetate extract of *Bauhinia purpurea* was presented in Table 5. Maximum scavenging activity of plant extract and Quercetin at 1000  $\mu$ g/ml was found to be 84.18% and 98.01% respectively. The IC<sub>50</sub> value of plant extract and Quercetin was recorded as 110 $\mu$ g/ml and 60 $\mu$ g/ml respectively.

The percentage of superoxide anion scavenging activity of methanolic extract of *Bauhinia purpurea* was presented in Table 6. Maximum scavenging activity of plant extract and Quercetin at 1000  $\mu$ g/ml was found to be 59.87% and 98.01% respectively. The IC<sub>50</sub> value of plant extract and Quercetin was recorded as 490 $\mu$ g/ml and 60 $\mu$ g/ml respectively.

 Table 4: Superoxide anion scavenging activity of Petroleum ether extract of Bauhinia purpurea

% of activity			(±SEM)*
S.No	Concentration (µg/ml)	Sample	Standard
		(Pet. ether extract)	(Quercetin)
1	125	$15.20 \pm 0.02$	$73.81 \pm 0.006$
2	250	$29.80 \pm 0.03$	$91.31 \pm 0.011$
3	500	$36.35 \pm 0.03$	$92.99 \pm 0.024$
4	1000	$44.35 \pm 0.02$	$98.01 \pm 0.012$
		$IC_{50} = 1390 \ \mu g/ml$	$IC_{50} = 60 \ \mu g/ml$

\*All values are expressed as mean  $\pm$  SEM for three determinations

Table 5: Sur	peroxide anion	scavenging	activity	of Ethvl	acetate extrac	t of <i>Bauhinia</i>	purpurea

		% of activity(±SEM)*		
S.No	Concentration (µg/ml)	Sample	Standard	
		(Ethyl acetate extract)	(Quercetin)	
1	125	$51.65 \pm 0.04$	$73.81 \pm 0.006$	
2	250	$59.35 \pm 0.02$	$91.31 \pm 0.011$	
3	500	$63.39 \pm 0.03$	$92.99 \pm 0.024$	
4	1000	$84.18 \pm 0.01$	$98.01 \pm 0.012$	
		$IC_{50} = 110 \ \mu g/ml$	$IC_{50} = 60 \ \mu g/ml$	

\*All values are expressed as mean  $\pm$  SEM for three determinations

		% of acti	vity(±SEM)*
S.No	Concentration (µg/ml)	Sample	Standard
		(Methanolic extract)	(Quercetin)
1	125	$19.93 \pm 0.02$	$73.81 \pm 0.006$
2	250	$29.54 \pm 0.01$	$91.31 \pm 0.011$
3	500	$51.41 \pm 0.03$	$92.99 \pm 0.024$
4	1000	$59.87 \pm 0.02$	$98.01 \pm 0.012$
		IC <sub>50</sub> =490 μg/ml	$IC_{50} = 60 \ \mu g/ml$

Table 6: Superoxide anion scavenging activity of Methanolic extract of Bauhinia purpurea

\*All values are expressed as mean  $\pm$  SEM for three determinations

Table 7: The total flavonoids content of various extracts of whole plant of Bauhinia purpurea

	S.No	Extracts	Total flavonoids content (mg/g) (±SEM)*
Γ	1	Petroleum ether extract of Bauhinia purpurea	$2.17 \pm 0.08$
Γ	2	Ethyl acetate extract of Bauhinia purpurea	$4.19 \pm 0.96$
	3	Methanolic extract of Bauhinia purpurea	$2.86 \pm 0.05$

\*All values are expressed as mean  $\pm$  SEM for three determinations

Based on the above results the  $IC_{50}$  values and percentage scavenging capacity, it was found that ethyl acetate extract of *Bauhinia purpurea* is more effective in scavenging superoxide radical than that of other two extracts.

## **Total phenol**

The total amount of flavonoids content of various extract of whole plant of *Bauhinia purpurea* was summarized in Table 7. Flavonoids present in food of plant origin are also potential antioxidants<sup>17, 18</sup>. Most beneficial effects of flavonoids are attributed to their antioxidant and chelating abilities<sup>19</sup>. Recent studies showed that many flavonoids & related polyphenols contribute significantly to the total antioxidant activity of many plants<sup>20</sup>. The higher content of flavonoids were found in ethyl acetate extract of *Bauhinia purpurea* than that of other two extracts.

# **References**

- 1. Cerutti PA, Oxidant stress and carcinogenesis, Eur J Clin Invest, 1991, 21:1–11.
- Halliwell, B Antioxidants and human diseases: a general introduction, Nutr Rev, 1997, 55: S44– S52.
- 3. Cerutti P, Oxy-radicals and cancer. Lancet, 1994, 344:862-863.

## **Conclusion**

The results of the present study was clearly indicated that the ethyl acetate extract of *Bauhinia purpurea* can be used as easily accessible source of natural antioxidants and as a possible food supplement in pharmaceutical industry. However, the ethyl acetate extract of *Bauhinia purpurea* was found high content of flavonoids. These in vitro assays indicate that this plant extracts is a significant source of natural antioxidant, which might be helpful in preventing the progress of various oxidative stresses. Therefore, further investigations need to be carried out to isolate and identify the antioxidant compounds present in the plant extract.

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- 4. Ames BN, Gold LS and Willet WC, The causes and prevention of cancer, Proceedings of the National Academy of Sciences USA,1995, 92: 5258–5265.
- Asolker LV, Kakkar KK, Chakre OJ. Supplement to glossary of Indian medicinal plants. Part-I (A-K). National Institute of Science Communication, New Delhi: 2000. P. 116-7.

- Kirthikar KR, Basu DB. Indian Medicinal Plants. Vol. 4.2n ed. Dehradun: Oriental Enterprises; 2000.p. 1255-7.
- Rajanarayana K, Reddy MS, Chaluvadi MR, Krishna DR. Bioflavonoids classification, pharmacological, biochemical effects and therapeutic potential. Indian J Pharmacol, 2001; 33:2-16.
- 8. Chopda MZ, Mahajan RT. Wound Healing Plants of Jalgaon District of Maharashtra state, India. Ethnobotanical Leaflets, 2009; 13:1-32.
- Harborne J.B. Phytochemical methods 11 Edn In Chapman &, Hall. New York: 1984, 4-5.
- Mensor LL, Meneze FS, Leitao GG, Reis AS, Dos santor JC, Coube CS and Leitao SG. Screening of Brazilian plant extracts for antioxidant activity by the use of DPPH free radical method. Phytother.Res, 2001,15: 127-130.
- 11. Winterbourne, C.C., Hawkins, R.E., Brain, M and Carrel, R.W (1975). The estimation of red cell superoxide dismutase activity. J. Lab.chem.Med. 85, 337-341.
- 12. Cameron GR, Milton RF and Allen JW. Measurement of flavonoids in plant samples. Lancet, 1943, 179.
- 13. Andlauer W and Furst P.Antioxidative power of phytochemicals with special reference to cereals. Cereal Foods World,1998, 43, 356-359

- Christensen Lars P. Tuliposides from Tulipa sylvestris and T. turkestanica, *Phytochemistry*, 1999, 51 (8), 969-974.
- 15. Shirwaikar, A, Punitha, ISR (2007). Antioxidant studies on the methanol stem extract of *Coscinium fenestratum*, Natural Product Sciences. 13 (1), 40-45.
- Zheng W, Wang SY (2001). Antioxidant activity and phenolic compounds in selected herbs. J. Agri. Food. Chem. 49, 5165-5170.
- Salah N, Miller NJ, Paganga G, Tijburg L, Bolwell GP and Rice Evans C, Polyphenolic flavonoids as scavengers of aqueous phase radicals and as chainbreaking antioxidants, Arch Biochem Biophys, 1995, 322(2), 339-346.
- Van Acker SABE, Van den Vijgh WJF and Bast F, Stuctural aspects of antioxidant activity of flavonoids, Free Rad Bio Med, 1996, 20(3), 331-342.
- 19. Hassig A, Liang WX, Shwabl K and Stampfl K, Flavonoids and tannins: plant based antioxidants with vitamin character, Med Hypotheses, 1999, 52, 471-481.
- Luo XD, Basile MJ, Kennelly EJ, Polyphenolic antioxidants from fruits of chrysophylolum cainito L. (Star apple), J Agric Food Chem, 2002, 50, 1379-1382.

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